

1949

Some factors influencing the slicing quality and the palatability of canned beef

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SOME FACTORS INFLUENCING
THE SLICING QUALITY AND THE PALATABILITY
OF CANNED BEEF

by
Mary Eloise Green

A Dissertation Submitted to the
Graduate Faculty in Partial Fulfillment of
The Requirements for the Degree of

DOCTOR OF PHILOSOPHY

Major Subject: Foods

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1949

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INTRODUCTION

Good slicing quality in meat is a desirable characteristic not usually secured in the canned product. The long processing time at high temperature during canning causes changes to occur in the muscle fibers and in the connective tissue so that the meat tends to fall apart and be stringy. These changes are not well understood and are suitable for investigation from many standpoints. For this study some of the physical aspects of the slicing quality and the palatability of canned beef have been considered.

The addition of table salt to meat during cooking is known to affect the flavor. Some experimental work has shown that immersion of cubes of beef in various brining solutions prior to cooking improves the tenderness, juiciness, and texture as well as the flavor. The hydrogen ion concentration of meat is a factor of importance in the catalytic changes that occur during aging of raw meat and during the cooking or canning process. The lactic acid content of the meat influences to considerable extent the pH under various storage and cooking conditions. Since substances added to the surface of meat, unless given a long time for penetration, tend to affect only the surface portions, it was decided to inject certain solutions into the meat. The substances

selected for experimentation were sodium chloride, lactic acid, and a mixture of the two substances.

In order to have cuts of meat from the same anatomical position, matching pairs of muscles were separated from the carcass and divided into cuts. The cuts from one side of the animal were injected and those from the other side served as controls. Four animals of the same carcass grade and one animal of a lower grade were used. After a suitable aging period the cuts of beef were canned. The processing time for the meat from three of the animals was a period considered to be safe from a bacteriological standpoint; a longer processing, typical of that often used in canning meat, was used for the other two animals.

The principal characteristics compared in the control and in the injected samples follow: (1) slicing quality, as indicated by number and character of slices obtained, by weight of unsliceable meat, and by judges' scores for sliceability, (2) palatability, as determined by judges' scores for aroma, flavor of meat and of liquid, tenderness, juiciness, and texture, and (3) microscopic appearance of muscle fibers and connective tissue of the rib portion of the longissimus dorsi muscle.

REVIEW OF LITERATURE

References to the slicing quality of beef in the literature are practically nil. Indirectly this quality has been considered from the standpoint of tenderness and texture of various meats. Much of the work pertaining to the palatability of beef is related to fresh or to frozen beef prepared by roasting, broiling, stewing, or cooking in deep fat. Canned beef or other canned meat has been studied primarily in regard to the bacteriological aspects and the adequacy of processing methods or to the retention of nutrients.

Considerable attention has recently been directed to the physical and chemical changes taking place in living muscle and to post-mortem changes occurring after slaughter of animals for food.

Structure and Composition of Skeletal Muscle

In its physical aspects skeletal muscle is made up of bundles of muscle fibers held together with connective tissue. This arrangement is apparent even to the unaided eye, but becomes increasingly evident on microscopic examination. Maximow and Bloom (20) state that skeletal muscle

fibers are from 10 to 100 or more microns in thickness and the length is usually shorter than that of the muscle. The boundary of the fiber is a transparent film (sarcolemma) and within it are long parallel threads (myofibrils), interfibrillar substances (sarcoplasm), and nuclei. The myofibrils have a maximum diameter of 1 to 2 microns and extend parallel to the long axis of the fiber giving the appearance of longitudinal striations within the fiber. Along the length of each fibril are alternate dark (A) and light (I) disks or bands which appear as crosswise striae in the muscle fiber. The myofibrils are the contractile part of the fiber.

The connective tissue meshwork of muscle is designated as endomysium if it surrounds the fibers within a bundle, as perimysium if it surrounds the bundle, and as epimysium if it forms the sheath around the muscle. Some fat globules are located within the connective tissue, as are blood and lymph vessels and nerves. The connective tissue has both collagenous and elastic fibers, the former being long, straight or wavy and containing fibrils, whereas the latter are usually branched and are lacking in fibrils. Collagenous fibers usually have a parallel arrangement; the elastic ones often appear singly. Tendons contain much collagenous tissue and ligaments are high in elastic connective tissue.

Proteins of Muscle

The proteins making up the different parts of the muscle have received considerable study. Bailey (5) refers to collagen and elastin as extracellular proteins and to the protein components of the sarcoplasm and fibrils as intracellular.

Intracellular proteins

Bailey (5) states that the fibrils consist of myosin (a globulin), a concentrated gel, and that sarcoplasm contains some proteins that are globulin-like and others that are albuminous in nature. Bate-Smith (6) in 1937 listed four intracellular proteins, two of which, myosin and globulin X, were said to be globulins and two, myogen and myoalbumin, were albumins. He listed the percentage composition of each in muscle as the following: myosin, 63 per cent; globulin X, 9 per cent; and myogen and myoalbumin together, 10 per cent. The isoelectric points were reported as: myosin, pH 5.5; globulin X, pH 5.2; myogen, pH 6.5-6.7; and myoalbumin, pH 3.0-3.5.

Myosin has been studied more extensively than the other intracellular proteins. Astbury and Bell (3, pp. 696-697) state that the pattern of structure of myosin is similar to the keratin of mammalian hair and both belong to the large

group of fibrous proteins, the characteristics of which are summarized in these words:

. . . that the structural unit of the group is a 'grid' consisting of long polypeptide chains crosslinked by means of their side-chains, that the main-chains of this grid are not normally in the extended configuration but are thrown into a sequence of folds transverse to the side-chains, and that when the fibers are stretched the grid is pulled out flat, only to return to its folded configuration when the tension is released.

The reversible transformation within the molecule from folded to extended form is said to be the basis of the long-range elastic properties of the fibrous proteins. Astbury and Dickinson(4) describe the myosin in muscle as being in the form of long chain-bundles of submicroscopic size extending approximately parallel to the axis of the muscle fiber. They state that the protein chains are normally in the folded configuration. They refer to "supercontraction" as the condition thought to exist when a muscle is placed in hot water or steam. More or deeper folds exist.

Astbury (2) reports the following values for the approximate number of amino acid residues (total 576) in what he calls the more reliable amino acids of rabbit myosin:

cystine/2 plus cysteine, 8; methionine, 15; serine, 23; threonine, 21; tyrosine, 13; tryptophan, 3; aspartic acid plus amide, 45; glutamic acid plus amide, 101; arginine, 27; lysine, 47; histidine, 7; amides, 57. A small proportion of

phosphorus is part of the permanent structure of myosin. He emphasizes the packing together of side chains in triads on alternate sides of the main chain and the alternate polar and nonpolar arrangement of individual side chains. He states that this kind of structural arrangement is in harmony with the concept that myosin "is both the working elastic mechanism in muscle and also a principal enzyme (adenosine triphosphatase) in the elastic cycle" (2, pp. 85-86).

Szent-Györgyi (33, 34, 35) published work in 1945, 1947, and 1948 in which myosin is recognized not as a single protein but as a complex made of two proteins. He assigned the name of actomyosin to the complex, retained the name of myosin for one of the proteins, and gave the new name of actin to the second protein. Since actin exists in both globular and fibrous forms, he designated the former as G-actin and the latter as F-actin. The complex has both globular and fibrous forms, G- and F-actomyosin, respectively. Szent-Györgyi enumerated several physical and chemical characteristics for each of these proteins. The protein he called myosin has an isoelectric point of pH 5.2, is soluble in water giving a limpid solution, behaves like a globulin except for solubility in water, has a tendency to form threads when slightly denatured, and is quantitatively precipitated from watery solution by small concentrations of neutral salts, such as KCl, but the precipitate dissolves

again if the concentration of salt is increased. In regard to the action of myosin with adenosine triphosphate, he states (33, p. 13): "The ATP adsorption is an exceedingly labile function of myosin" He points out that as K is adsorbed, myosin becomes more and more capable of binding ATP also. Ca or Mg alone in low concentration does not cause ATP to be adsorbed, but in the presence of KCl they greatly increase the adsorption of ATP.

Actin is more stable than myosin, according to Szent-Györgyi (33). It has an isoelectric point of pH 4.7, is readily denatured by heat, and is not precipitated by alkali salts. The F- and G- forms have widely different properties and the transformation from one form to the other is thought to occur in each contraction cycle in living muscle. F-actomyosin is formed from F-actin and myosin and is described by Szent-Györgyi as "a typically fibrous colloid with very long particles" (33, p. 24). G-actomyosin can be prepared by bringing about contraction of the F- form. In explaining the action of these proteins in muscle action, he pictures actomyosin as an elongated particle composed of two parallel parts, one of which is long and continuous (myosin) and the other of which (actin) is discontinuous and capable of becoming globular. The elongated part shrinks more rapidly than its partner, which causes the particle to bend into a circular shape. He represents the fibril as a spiral rod.

This spiral nature of myofibrils is questioned by Speidel (32). By the use of striated muscle from the sea spider, and, to a limited extent, muscles of shrimp, frog, and rabbit, Speidel was able to show by means of photomicrographs that the cross striae in the myofibrils are arranged transversely, not spirally.

Extracellular proteins

The principal extracellular or structural proteins in muscle are collagen and elastin. The chief characteristics of collagen, as stated by Bull (11), are the inelasticity at body temperature, the high content of proline and hydroxyproline, the ability to contract spontaneously to about one-fourth of its length when heated to about 60°C., long range elasticity of the contracted fiber while hot, and the spontaneous partial recovery of length on immersion of the fiber in cold water after contraction. The thermally contracted collagen has an amorphous X-ray diffraction pattern.

Astbury (2) states that few chemical analyses have been carried out on pure collagen fibers; gelatin has been analysed much more extensively. He reports that the X-ray patterns of collagen fibers and of oriented gelatin have the same main features but differ in certain details such as high spacings in true collagen that are lacking in the gelatin pattern. The general molecular plan of the collagen group is

a repetition of glycine and imino residues along the polypeptide chains at intervals of three residues.

An outstanding characteristic of elastin is its elastic quality at room temperature. Bull (11) states that the unstretched elastin from ligamentum nuchae has a completely amorphous diffraction pattern. Stretching to 200 per cent extension results in some orientation of X-ray spacings but no new spacings occur. He mentions the high proline content of elastin.

Denaturation of proteins

Mirsky (21) discusses denaturation of proteins in general, then considers a special case, that of denaturation of myosin. He points out that the most frequent signs of instability of protein, i.e., tendency to denature, are loss of solubility and loss or impairment of a specific property. Among the properties affected by denaturation, according to this author, are: crystal form, solubility, viscosity, chemical changes, reversibility, formation of fibers, and configuration. He states in connection with these properties that many native forms of proteins have been crystallized but not the denatured forms. He says that at the isoelectric point a denatured protein cannot be dissolved in neutral salts as can native globulins; that denatured proteins are more viscous than native ones; the number of SH, S-S, and

phenol groups is increased by denaturation; and that denaturation is a change from a compact to an extended configuration. In writing of the reversibility of denaturation, he cites evidence that coagulation takes place in two steps, the first being reversible, the second not reversible, the latter occurring at a slightly higher temperature than the former. Anson (1) enumerates the following agents which can cause denaturation of protein to occur: heat, surface action, ultra-violet light, high pressure, organic solvents such as alcohol, and reagents which can dissolve coagulated protein such as acid, alkali, urea, detergents, and others. He states that the molecule of protein opens up and changes its shape during denaturation.

Variations among Muscles and among Animals

The manner in which meat is divided into retail cuts in the United States often results in the inclusion of several muscles in each steak, roast, or piece of meat to be cooked. Differences in tenderness, texture, and other characteristics of individual muscles make it difficult to secure uniformly well prepared meat. Ramsbottom and Strandine (28) made a systematic study of some of the physical differences among the major muscles constituting the beef carcass. Although they were primarily interested in the factors responsible for tenderness, they reported data on the identification,

location, composition, weight, pH, tenderness, and histological rating of 50 representative muscles in three heifer carcasses of the same grade. The variation in fat content of the muscles was found to be from an average of 1.5 per cent in the extensor carpi radialis of the foreshank to an average of 18.1 per cent for the intercostal muscles. The average moisture content of the first named muscle was 76 per cent, whereas the latter contained 62.5 per cent moisture. The average fat content of all muscles was 5.7 per cent; the moisture, 72.2 per cent. The average weight of the smallest muscle studied (sartorius) was 0.4 pound; of the largest (biceps femoris) 10.2 pounds. Comparisons of tenderness were made by means of (1) histological rating of collagenous and of elastic connective tissue content, (2) shear force of both raw and cooked beef, and (3) judges' scores for tenderness of the cooked meat. The lowest average shear force for a 1/2-inch cylinder of raw beef was 3.8 pounds for the longissimus dorsi muscle; the highest, 26.0 pounds for the cutaneous muscle. For the cooked beef, the average shear force values varied from 7.1 pounds (psoas muscles) to 16.3 pounds (rhomboideus). Correlations that were statistically significant were found between the histological rating of the raw beef and the shear force of the cooked meat.

Prudent (27) used a chemical method for determination of collagen and elastin in four beef muscles from each of two

animals. She found differences among the muscles and between the animals in the percentages of the collagen and elastin nitrogen in relation to total nitrogen. The dairy cow (carcass grade Cutter) had a higher collagen nitrogen but a lower elastin nitrogen content than the steer (carcass grade Good). She stated that the muscles of the cow were smaller than those of the steer and that the fat and connective tissue of the cow were bright yellow.

Paul (23) found a significant difference in the diameters of muscle fibers in various muscles as revealed by histological study. The differences among muscles were highly significant after the meat was cooked. She grouped the muscles according to diameter of fibers as follows: smallest, semitendinosus, psoas major, adductor; medium, semimembranosus and biceps femoris; largest, vasti. The gastrocnemius muscle had the largest fibers but they shrank the most, hence did not fit well into the arbitrary groups. She found there was no high correlation between the number of muscle fibers per bundle and the tenderness of the meat.

Harrison (18) compared the histological characteristics of four muscles from three grades of animals and found the psoas major muscle, which is the most tender of the four, had slender fibers with distinct cross striae. There was little connective tissue in the psoas muscle compared to moderate or

large amounts in the others. The muscle fibers of the older animal were more gnarled and worn than those in the younger animals. Fat located between the muscle fibers was noticeable in the aged animal. The sections from two animals contained larger amounts of collagen than sections from the other two.

Deatherage and Harsham (15) found differences in the initial tenderness levels of beef cattle as revealed by judges' scores for tenderness of broiled steak the second or third day after slaughter of the animals. Paul and McLean (25) used roasts from veal calves of three different carcass weights, approximately 50, 125, and 200 pounds. After roasting to internal temperatures of 71°, 77°, 82°, or 88°C., the cuts from the smallest animal were judged to be the most tender, even though this meat had no apparent intramuscular fat and very little fat between the muscles.

In another phase of the study Paul and McLean (26) observed variations in the histological structure of the muscles from the hind quarter of veal. Differences in amounts of connective tissue and in size of fasciculi were marked. The semitendinosus and the biceps femoris muscles were judged after cooking as the juiciest of the muscles studied. The former of these muscles (judged to be tender as well as juicy) had much connective tissue with abundant elastic fibers

and the fasciculi were medium in size. The latter muscle had thick masses of connective tissue in the perimysium but most of the fibers were collagenous rather than elastic. The semimembranosus was described as the driest and one of the toughest of the muscles studied. The connective tissue was relatively small in amount and most of the fibers were of the collagenous type.

Influence of pH

According to Fenn and Maurer (16), the pH of intercellular fluids in living tissue (frog muscle) is 7.3 to 7.5 and the pH of the interior fibers is 6.9 or lower. They state that post mortem a pH of 5.6 to 6.0 is common and a value of 5.3 is possible. Bate-Smith (7) states that he found a range of pH values of 5.36 to 5.80 for the psoas muscles of beef from 29 carcasses and a range of 5.5 to 6.0 for the pH of thigh muscles from 6 other beef animals. He makes the statement that a variation in pH between 5.4 and 6.0, except for occasional values outside this range, may be expected for beef in full rigor. Ramsbottom et al. (29) in their study of 25 representative beef muscles found a range of pH values from 5.5 to 5.8. In a later work on 50 beef muscles, Ramsbottom and Strandine (28) reported a range of 5.5 to 6.0 and an average pH of 5.7.

Hall, Latschar, and Mackintosh (17) in studying the problem of "dark cutting" beef found this kind of beef to have an abnormally high pH. Among the other characteristics observed were "low glucose, practically no glycogen, high inorganic phosphate, low oxidation potential, and rapid oxygen uptake" (17, p. 81). They give the value of pH 6.4 as characteristic of black beef and 5.4 for light beef. They found approximately 900 milligrams lactic acid per 100 grams bright rib eye meat having a pH of 5.50, whereas the addition of approximately 450 milligrams of lactic or pyruvic acid was required to bring the pH of 100 grams of beef rib eye from pH 6.50 to 5.50. They conclude that the difference of primary importance between bright and dark beef is the difference in the amount of lactic acid present. They say the cause of dark cutting beef appears to be a deficiency of glycogen in the tissues at the time of slaughter.

Winkler (39) was able to show a relationship between the color of beef and its pH. He injected samples of minced pork and of beef with solutions of dilute lactic acid or of ammonia, stored the samples at 0°C. for 3 to 5 days, and by means of a color comparator determined differences in color and in the scatter of the wave bands. Choosing the curve for the scatter of the red band for pork as typical (although the pH curve for beef is said to be similar), he points out that from pH about 4.5 to 5.5 the meat becomes lighter

and beyond 5.5 darker in color. In another phase of the study Winkler (40) tested the work required to shear raw beef and pork samples (loin of beef from three animals) in which the pH values had been adjusted by injection of lactic acid or ammonia solutions. He concluded that the "addition of sufficient lactic acid or ammonia to raw pork or beef made the meat more tender" (40, p. 13). He pointed out a greater variability in tenderness in beef from different animals than in pork, even though the samples were at the same pH.

Bate-Smith (7) in reviewing the significance of pH in the ripening of beef enumerates these characteristics of meat at the upper end of the pH range: dark color, slimy or sticky feel, flabbiness, juice not readily expressed, high electrical resistance; salt does not readily penetrate it from curing pickle. He refers to them as defects in the meat and believes they are related to the substance of which the fibrils of the muscle is composed. The main protein constituent, myosin, has an isoelectric point a little above pH 5.3 at which point there is minimum of swelling. He describes the myosin of living muscle as a weak jelly, and states that as the pH falls this jelly shrinks; at pH 6.5 to 6.0 the fibrils shrink apart and scatter light. He attributes high electrical resistance and high pH to swelling of the fibers and the accompanying narrowness of the channels through which ions can move freely. The stickiness at high pH's he

says is due not only to swelling of the fibers but also to some dissolution of myosin.

Post-mortem Changes during Aging of Beef

The outstanding change which occurs soon after slaughter of the animal is a stiffening of the muscles, i.e., the development of rigor mortis. Bate-Smith (7) in writing of rigor mortis and aging of beef relates rigor to glycolysis and the breakdown of adenosine triphosphate. The lactic acid produced from the glycogen acidifies the muscle so that the pH steadily falls from near 7.4 as circulation stops. He emphasizes that the production of acid is not the cause of rigor mortis even though a change in pH greatly affects the physical characteristics of myosin. He states (7, p. 7) that "when sufficient acid is produced, rigor always sets in when the muscle reaches a pH in the neighborhood of 6.3." The removal of adenosine triphosphate from muscle is cited by him as the immediate cause of the stiffening in muscle during rigor. He does not state how rigor is resolved, but says (7, p. 33):

The actual cause of the increase in tenderness during the ripening process has not been elucidated. The most likely theory is that it is due to proteolysis by tissue proteinase, such as cathepsin.

Paul, Lowe, and McClurg (24) noted a difference in the

fat of roasts cooked without storage and of those aged 1 day or longer. In the former the fat was soft and oily, whereas the fat of the latter was firm and brittle. The surface of roasts stored for 0 or 1 day was dry; those stored 2 and 4 days were moist; by the 9th day moisture collected in the paper about the roasts but the surface of the meat was fairly dry; and at 18 days the surface was sticky. The roasts cooked without storage developed rigor during the roasting process, with the exception of the psoas major which had become stiff before it was put into the oven. The psoas was out of rigor by the time the roasting was completed, but the other muscles were stiff, hard, and very difficult to cut. The authors (24, p. 224) describe the procedure of trying to cut the meat in rigor as "similar to cutting a rubber cork." The stored roasts were out of rigor when cooked and the tenderness increased up to the 9th day of storage. With 18 days of storage the results were variable. Deatherage and Harsham (15) compared the tenderness of beef at various intervals of storage up to 41 days. They found a fairly consistent tenderizing effect with increased storage up to 17 days; after that time some carcasses continued to increase steadily in tenderness and others became more tough at one or more of the aging periods.

Harrison (18) compared the tenderness of three grades of beef after aging periods of 1, 2, 5, 10, 20, and 30 days.

The average scores for all roasts indicated a gradual increase in tenderness as aging progressed, with the greatest increase occurring in the first 10 days. For individual muscles the relation of tenderness to time of aging was not always linear. She studied the histological structure of muscles at the different storage periods. Disintegration of muscle fibers was evident at about the 10th day of storage in three of the four animals and became increasingly evident at the 20th and 30th days of aging. This disintegration was described as consisting of destruction of the striae in strips of the muscle fibers, resulting in increased fragility of fibers.

Prudent (27) analyzed samples of meat from two of the animals of Harrison's study to determine the amount of chemical breakdown occurring in collagen and elastin during storage of the meat. She found that the length of the storage period had little if any effect on the degradation of collagen and elastin of the muscles studied. The tenderizing effect which had occurred, as shown in the findings of Harrison (18), could not be explained on the basis of the degradation of collagen and elastin during storage. — *Method at fault.*

The length of the storage period has an effect on other factors of beef than tenderness. Paul (23) reported an increase in electrical conductivity with storage. Juiciness, as indicated by judges' scores, gradually increased with storage time; the amount of press fluid dropped markedly from

the 2d to the 9th day of storage, then rose to a high figure at the 18th day. The aroma scores increased up to 9 days, but dropped after 18 days of storage. The flavor of fat became less desirable with the increased length of storage period; by 18 days it had developed rancidity.

Paul measured the pH of raw beef from various muscles during aging periods of 0, 1, 2, 4, 9, and 18 days. She states that the changes during storage followed the usual pattern, namely, "a drop followed by a slow rise" (23, p. 69). The changes in pH during storage were computed to be highly significant and the differences between muscles to be significant. The muscles having the highest pH were also said to have had the strongest odor of any of the aged roasts before cooking.

Harrison (18) reported acidification of muscle post mortem and described the change as rapid during the first 1 to 2 hours. There was a slow rise in the pH of the muscles during storage. The average values for all muscles for 1, 2, 5, 10, 20, and 30 days of aging were 5.47, 5.45, 5.47, 5.49, 5.51, and 5.88, respectively.

The weight lost by roasts during various aging periods was determined by Harrison (18). The loss was greatest for the meat from the carcasses graded Good; intermediate, for the meat from the carcass graded Commercial; least, for that

from the carcass of Cutter grade. The roasts lost slightly more weight as the aging time increased. Muscles differed in the amount of weight lost during aging.

Changes during the Cooking of Beef

The changes that occur during the cooking of beef are so complex that it is very difficult to determine the role of the individual factors. Some studies have been made of the effect of heating on portions of animal tissue composed principally of collagenous or of elastic tissue, i.e., of tendons and of ligaments, respectively.

Changes in tendons and ligaments

Harrison (18) used strips of tendon from around the anterior end of the longissimus dorsi muscle and from the Achilles tendon. The strips were heated in distilled water at 60°, 65°, 70°, and 95°C. for periods varying from 15 seconds to 30 minutes. She reported a progressive decrease in length of tendons as the temperature was increased. With increased length of time of heating, the length was found to decrease in the first stages, with no further shortening occurring as heating was continued. Softening occurred that was evident in the lower shear force values for the heated than for unheated samples. Strips of ligamentum nuchae were heated in distilled water at 70° and at 95°C. for 30 minutes,

and for 1 and 2 hours. The strips became thicker and wider on heating than when raw. They tended to decrease in length with cooking but the changes were small compared to those in the tendons. Considerable variation in results was noted among animals and even among samples from the same animal. The shear force values for the ligaments heated at 95° or 70°C, showed evidence of a tenderizing effect on the elastic tissue. This is contrary to the belief that cooking does not affect elastic tissue.

Ramsbottom et al. (29) compared the relative tenderness of raw connective tissue from the infraspinatus and biceps femoris muscles and found that more than 120 pounds (the capacity of the shearing device) were required to shear it. After cooking, the force required was 21.5 pounds. Yellow elastic tissue (ligamentum nuchae) had a shear force value of 81.1 pounds raw and 42.3 pounds cooked. Fatty tissue improved materially in tenderness during cooking irrespective of the content of connective tissue. The rectus femoris muscle increased in toughness on cooking. Hence these authors concluded that factors other than connective tissue and fat have a considerable effect on tenderness of meat. They state that coagulation and denaturation of protein are among the factors which have a greater negative effect than the positive effect of partial hydrolysis of collagen.

Changes in meat

Ramsbottom and Strandine (28) tested the theory that cooking tenderizes beef. In their study of 50 representative muscles from three heifer carcasses graded Good, they found that cooking the meat to an internal temperature of 76.7°C. (170°F.) in lard at 121.1°C. (250°F.) resulted in shear force values that were higher than for the raw meat for 35 of the 50 muscles. These results indicated a decrease in tenderness with cooking. Ramsbottom et al. (29) reported that in preliminary tests the beef cooked in lard at 121.1°C. (250°F.) cooked more quickly to 170°F. than in the oven at 162.8°C. (325°F.). The oven-cooked meat was consistently more tender and had less variability between muscles than the meat cooked in deep fat.

Cover (12) studied the effect of extremely low rates of heat penetration in relation to tenderness of beef. Two oven temperatures were used, 80°C. (176°F) for the experimental roasts and 125°C. (257°F.) for the controls. She found that the well-done stage as judged by subjective methods was reached at an internal temperature of 70°C. for the experimental roasts compared to an internal temperature of 80°C. for the control roasts. Similarly, the rare stage was reached at 58° or 59°C. with extremely slow roasting and at 63°C. with roasting at 125°C. A very much longer time in the oven was

required at 80°C. than at 125°C. to reach a given internal temperature. The roasts cooked slowly were reported to be more tender by all measures than those cooked at the higher temperature. Without exception, the roasts which required as much as 30 hours (in the 80°C. oven) to lose their pink color were tender, but the roasts cooked a shorter time (in either oven) were not always tender. Roasts from the extremely slow oven lacked the usual plump appearance; those cooked to the well-done stage were so tender as to offer no resistance to cutting or chewing and the texture was described as mealy or powdery in the mouth. The actual moisture content was not determined, but the meat seemed dry as it was chewed.

In the study of veal roasts cooked in ovens at 163°C. (325°F.), Paul and McLean (25) found that in general the color and the palatability factors (flavor, texture, and tenderness) improved with each increase of internal temperature (71°, 77°, 82°, and 88°C.) for the three sizes of animals. Juiciness decreased with increased internal temperature. Cooking losses and the cooking time increased as the internal temperature was increased. The amount of soluble nitrogen in the meat, contrary to the trend expected if collagen is changed to gelatin, showed a decrease from the raw meat through the various stages of cooking. The histological changes observed in the connective tissue of the cooked veal were of two types: (1) an apparent swelling or

"spreading" of the collagenous fibers particularly in the semitendinosus muscle, but no change in the elastic fibers, and (2) a disruption in the fine connective tissue sheath (endomysium) of the individual muscle fibers. Both of these changes would contribute to increased tenderness in the meat. They further point out that there was some evidence of disruption in the sarcolemma of the muscle fibers during cooking. This was shown as a change in the edges of the muscle fibers from a smooth, straight appearance in the raw veal to a rough, irregular outline in the roasted meat. The staining reaction of the cooked connective tissue was observed to be less intense than that of the raw tissue. These authors describe in some detail the distribution and character of the collagenous and elastic tissue in each of the several muscles studied. (See Paul and McLean, 26.)

Bell et al. (9) used a chemical method for measuring the tenderizing effect of cooking on meat. For beef boiled in an excess of water to 85°C., they reported a conversion of collagen to gelatin to the extent of about 22 per cent. The percentage of protein represented by gelatin in the raw meat was 10.86 and that in the cooked meat was 8.37. Prudent (27) secured results of an opposite nature. She found by chemical determinations that cooking to 70°C. by immersion in fat at 96° to 98°C. resulted in little if any changes in the collagen and elastin content of the muscles studied.

Ref. 27

The pH values for cooked meat have been reported to be slightly higher than for uncooked meat. Harrison (18) found average pH values of 5.76, 5.74, 5.77, 5.78, 5.73, and 5.87, respectively, for the cooked roasts aged for 1, 2, 5, 10, 20, and 30 days. Bendall (10) cooked beef at 100°C. for 1 hour and found considerable shift to the alkaline side.

Aroma and Flavor of Beef

Crocker (14) has made a critical analysis of the flavor of unaged beef and of the influence of boiling and of low-temperature heating on the flavor. He states that the flavor of raw meat is mostly in the juice, not the fiber, and that the flavor is "weak, sweetish, salty and generally blood-like" (14, p. 180). He says that most of the flavor of cooked meat comes from the meat fiber. He describes the flavor of cooked beef in these words (14, p. 180):

This meaty flavor, typical of cold roast beef or pot roast, was apparently due to volatile substance detected by the sense of smell, even though chewing was needed to release it. It was fragrant, moderately acidic, only slightly burnt, and distinctly caprylic. It was definitely sulfury.

He says that cooked meat flavor is essentially an odor, for only a trace of sweetness was evident when the nostrils were held. Some astringency in the mouth was noted. The effect of length of time of boiling was studied and the flavor of cooked beef was found to increase up to 3 1/2 hours of

boiling, whereas after that time a gradual loss of flavor occurred. Beef boiled in a citric acid solution (2 per cent strength based on the weight of the meat) had an acetic-like odor and caprylic flavor resembling corned beef. The pH of the raw beef was 6, that of the meat cooked in citric acid solution was 3.5. Beef boiled in 1 per cent sodium bicarbonate solution developed a pH of 8.87 and was said (14, p. 180) to have "an aminic and sulfury odor, suggestive of both clams and eggs, with some piperidine evident" and the taste was described as "very weak." Beef bones were found to contribute almost no flavor but gave gelatin and tallow to preparations. Marrow and tissue fats were said to contribute little meat flavor.

Effect of Added Substances

In some preliminary investigations Sair and Cook (30) observed a relationship between the amount of drip from meat (prior to freezing) and the pH of the tissues. They decided to extend the study of this relationship by artificially adjusting the pH of meat samples. They injected 20 milliliters of lactic acid or ammonium hydroxide of suitable concentration into 800- to 900-gram pieces of pork, beef, and mutton with a calibrated 5-milliliter hypodermic syringe. The injections were made at numerous uniformly distributed points. Those samples not requiring an adjustment of pH were

injected with 20 milliliters of distilled water. After storage of the injected samples for 3 days at 0°C., the samples were minced and divided into experimental lots, some to be frozen and others to be left unfrozen as controls. These investigators found a marked relation between the amount of drip and the pH of the meat. The maximum drip was obtained between pH 5.0 and pH 5.2 for all three kinds of meat. They plotted curves for the drip from the frozen (total) and unfrozen (control) samples. The curves descended slightly on the acid side to pH 4.4 (the lowest pH value tested) and also descended, but more sharply, on the alkaline side to about pH 6.4. At pH 6.4 the difference in the drip from the control and frozen samples (net drip, difference between amount for control and frozen samples) became zero and the amount of drip was very small. The shape of the drip curves was nearly identical for the three kinds of meat, but the quantity of liquid exuding from unfrozen beef was slightly less than the amount from pork or mutton.

Snyder (31) studied the effect on cooking losses of the addition of salt to the surface of beef roasts before roasting and of the addition of salt to beef stew before cooking. She reported the formation of a crusty outside layer and a penetration of salt flavor to the depth of less than 1/2 inch below the surface of the roasts. There was a development of gray color within the meat which corresponded to the depth of

penetration of salt flavor. In the case of the stews there was said to be no texture or color differences in the salted and unsalted cubes, but there was an improvement in flavor due to the addition of salt.

Crain (13) and Tofte (37) treated 3-inch cubes of beef with three types of brining solutions: (1) salt, 10 gm. to 100 cc. water, (2) vinegar, 25 cc. to 75 cc. water, and (3) salt and sugar, 7.5 gm. of each to 100 cc. water. After standing in the brining solution for 72 hours in the refrigerator, the cubes of meat were seared, then cooked in covered stew pan or Dutch oven to an interior temperature of 205°F. (200°F. in the case of the vinegar-treated meat). The brined samples gained in weight prior to cooking; the controls lost weight. Cooking losses were higher, however, for the brined than for the unbrined samples. The vinegar-treated meat had a lower pH than the controls, but the difference was more marked in surface samples than in those from the interior. The pH values were generally higher for cooked than for raw meat. Tofte reported low pH's for vinegar-treated samples even after cooking, but Crain found no significant differences after cooking due to vinegar treatment. All three brining treatments resulted in tenderization of the meat, except for one animal reported by Tofte, the meat of which was moderately tender before brining. The judges generally preferred the brined meat to the controls in flavor and in

juiciness, but the differences were much greater for salt or salt-sugar treatment than for vinegar. Scores for aroma were variable but in general were slightly higher in brined than in control samples.

Both investigators (13, 37) described color changes in the brined samples. The vinegar-treated meat became purplish gray in the outer portions and was dark on cooking; salt-treated meat was slightly gray. Many samples immersed in the salt or the salt-sugar solution developed iridescence on the surface; only a few control or vinegar-treated samples had this characteristic. Brined samples failed to develop a brown exterior during searing. Tofte (37) studied the histological effect of brining the beef cubes. She found a decrease in the distinctness of outline of the muscle fibers and decreased clarity in differential staining of connective tissue and muscle fibers as a result of the brining treatment. In the slides of cooked meat the collagen was barely apparent; the elastin showed no perceptible change. The amorphous appearance of brined meat and the differences between surface and interior samples of vinegar-treated meat were less marked after cooking than while raw.

Bate-Smith (8) reported the use of phosphate solution to aid in tenderization of beef. The concentration giving the greatest effect was 0.2 M, but an appreciable effect was

noted with a fourth of that concentration. A suitable mixture of mono- and di-hydrogen phosphate to give a pH value between 6 and 7 was used. The addition of concentrated meat stock from previous cooking was recommended by this investigator as an alternative method of securing a tenderizing effect. Lowe (19) describes short studies in which 0.2 M and 0.4 M mixtures of NaH_2PO_4 and Na_2HPO_4 were added to stews or in which the beef was brined in the phosphate solutions. The addition of broth to stews was also studied. Results for any of these treatments varied with the size of the beef cubes. In small cubes, the salt solution penetrated a larger proportion of the cube and tenderizing occurred. There was less effect on large cubes. The flavor of the treated samples was generally preferred by judges to that of the meat left untreated.

The immersion of cuts of beef in solutions of calcium chloride was also reported by Lowe (19). The lower concentrations (0.1 M and 0.2 M) had no appreciable effect on shearing strength, press fluid, or the qualities tested subjectively. The higher concentrations (0.4 M and 0.5 M) imparted a bitter flavor. Some weight gains and a higher content of calcium in the brined than in the unbrined meat were found, but the differences were less marked in cooked than in raw samples.

Canning Procedures

Directions for the commercial preparation and processing of various fruits and vegetables are available in the literature, but the references for commercially canned meats are very limited. A bulletin of the National Canners Association Research Laboratory (22), which outlines canning methods for several low-acid foods in metal containers, provides no time or temperature tables for meat and meat products but refers the reader to directions from research laboratories connected with the canning industry.

The Bureau of Human Nutrition and Home Economics (38) has published a bulletin for use in the home canning of meats. The directions for the raw packing of beef, veal, pork, or lamb as large pieces include the following selected points: trimming away excess fat, placing pieces of meat in the cans with the grain of the meat running lengthwise, filling tin cans to the top with raw meat, preheating the open cans of meat in water extending to about 2 inches below the rim for a period of about 50 minutes in tin cans or to 170°F. for the meat at the center of the jar, sealing tin cans, and processing at once in a steam-pressure canner at 10 pounds pressure (240°F.) for 65 minutes for number 2 tin cans (75 minutes for pint and 90 minutes for quart glass jars).

A technical bulletin by Toepfer et al. (36) reports the findings from experiments in processing inoculated packs of low-acid foods by home canning methods. Recommendations were formulated for various foods in certain sizes of cans based on the data from the heat penetration studies. For pork and beef, the recommended processing temperature is 240°F. and the time periods for certain sizes of cans are the following: beef in quart jars, 90 minutes; pork in quart jars, pint jars, number 3 or number 2 1/2 cans, and number 2 cans, respectively, 90, 90, 75, and 65 minutes. These investigators point out that the processes may be more severe than necessary for home canning, but no reduction is advisable until sufficient data on heat resistance and thermal-death-time curves of spoilage organisms in food media are available.

EXPERIMENTAL PROCEDURE

Preliminary Investigations

In the preliminary part of the study, several different substances were tested for their effect on the slicing quality and palatability of canned beef. Solutions of phosphate, of chloride, and of acid in various concentrations and mixtures were used, and the injected meat was stored for different lengths of time before canning. For example, a solution of mono- and di-sodium phosphate in 0.1 M and 0.2 M concentrations was used for injection of beef cuts and the meat was aged for 4, 9, or 14 days. Some other solutions tested were sodium chloride and a mixture of sodium chloride with lactic acid or with mono-sodium phosphate for three different aging periods. A few cuts of meat were injected with calcium hydrogen phosphate solution or with this substance in combination with sodium chloride. A few experiments were done in which the solution used for injection was one of the following: ascorbic acid, ascorbic acid and sodium chloride, or ascorbic acid, sodium chloride, and lactic acid.

Little or no improvement in slicing quality was noted for injected samples in comparison with control samples with phosphate or ascorbic acid treatment. Samples treated with

sodium chloride, even though they showed no improvement in sliceability, were found to be much more tender than the controls. An improvement in the texture of the meat was noted and the flavor was more desirable than in the untreated samples. Sodium chloride-lactic acid-treated samples showed slicing quality that was slightly improved or similar to that of the control samples without impairment in palatability.

On the basis of these preliminary findings, the treatments selected for further investigation were the use of the following three kinds of solutions for injection of the raw beef prior to aging and canning: (1) sodium chloride, (2) lactic acid, and (3) a mixture of sodium chloride and lactic acid.

Selection of Animals

Five animals were used in the study after a period in which meat from the same number of animals had been used in preliminary investigations. All the animals were procured by the Animal Husbandry Department of the College. Four animals were steers of beef type purchased from a farmer near Ames, and the fifth was an aged dairy cow purchased from another farmer. The four steers were identified as Animals VI, VII, VIII, and IX, respectively, and the cow, as Animal X. The carcass grade of each of the steers was Commercial and

that of the cow was Cutter in accordance with U. S. Government standards.

Slaughter of Animals, Separation of Muscles,
and Division into Cuts

Each of the animals was slaughtered in the Animal Husbandry abattoir by E. A. Kline, Instructor in Animal Husbandry, and Julius T. Jensen, Meat Laboratory Assistant, or by students under their direction. The carcass was divided into halves and the two sides of beef were allowed to hang in the cooler at 34° to 36°F. until the following day. The sides of beef were divided into quarters between the 12th and 13th ribs and matching pairs of muscles were separated from the carcass. Belle Lowe, Professor of Foods and Nutrition, Alma R. Plagge, Research Associate, and the author separated the muscles from the carcass, divided the muscles into cuts, and carried on the preparation of the meat for canning and subsequent testing.

The muscles selected for use because of their suitable size, shape, and structure were the following: (1) longissimus dorsi, loin portion, (2) longissimus dorsi, rib portion, (3) psoas major and psoas minor, (4) semitendinosus, (5) semimembranosus, and (6) biceps femoris.

Before division into cuts, the extraneous fat and heavy

connective tissue were trimmed from the muscle. From three positions within each muscle, small samples of meat were taken for histological and for pH determinations. The location of each position is indicated in the diagram of the respective muscle (Figures 1, 2, 3, 4, and 5). The approximate size of each histological sample was 3 cm. x 1 cm. x 1 cm. The sample was cut in half crosswise and the two pieces were placed in an individual bottle of formalin-physiological salt solution for fixation. Each sample for pH determination was cut to a weight of 10 grams, wrapped individually in moisture-vapor-proof cellophane, and frozen at -30°F . to be kept for later assay.

The size and general shape of the beef muscles used in the study were suited to canning in the number 2 size of tin can (diameter, $3 \frac{7}{16}$ inches; height, $4 \frac{9}{16}$ inches). Some advantages were that pieces of meat approximately the size of the container could be cut from the muscles, the canning procedures could be carried out to advantage, and suitable samples for testing could be secured from the canned meat. For the experimental work of the problem, cuts of beef were needed which would be large enough to fit the container after removal of pH and histological samples and which would provide for losses in weight during aging. By dividing each of the smaller muscles into three parts and the larger muscles into six parts, pieces of approximately the desired

Figure 1. Longissimus Dorsi Muscle.

- A, B, and C - Cuts of beef for canning.
- a and b - Histological and pH samples, respectively, from muscle before division into cuts and after aging one day.
- c and d - Histological and pH samples, respectively, from beef cuts aged eight days.
- e and f - Histological and pH samples, respectively, from canned beef.
- g, h, and i - Slices for scoring by judges.

Division of the Longissimus Dorsi
Between 12th and 13th rib

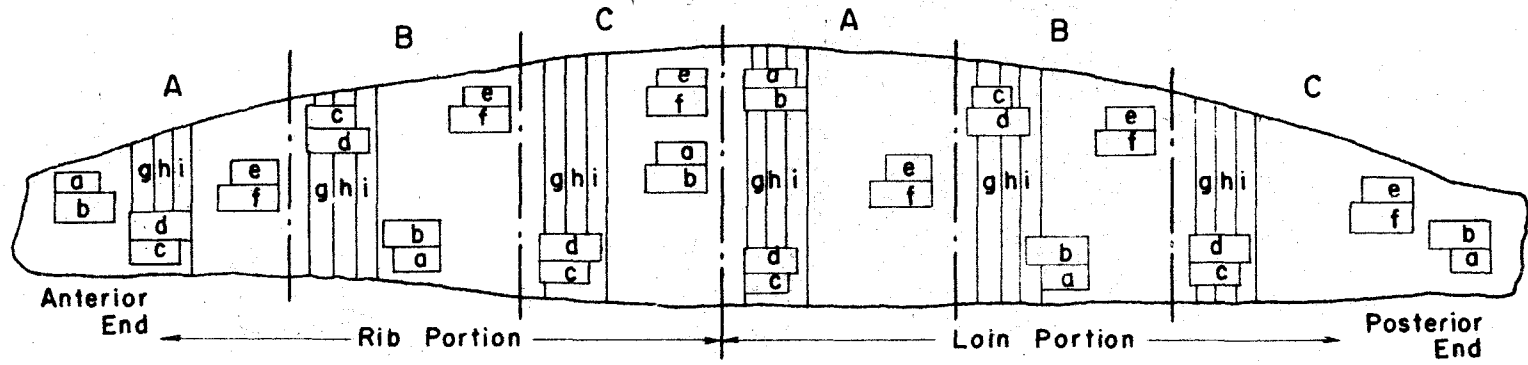


Figure 1.

Longissimus Dorsi Muscle.

Figure 2. Psoas Major and Psoas Minor Muscles.

- A, B, and C - Cuts of beef for canning.
- a and b - Histological and pH samples, respectively, from muscle before division into cuts and after aging one day.
- c and d - Histological and pH samples, respectively, from beef cuts aged eight days.
- e and f - Histological and pH samples, respectively, from canned beef.
- g, h, and i - Slices for scoring by judges.

Division of the Psoas Major and Psoas Minor

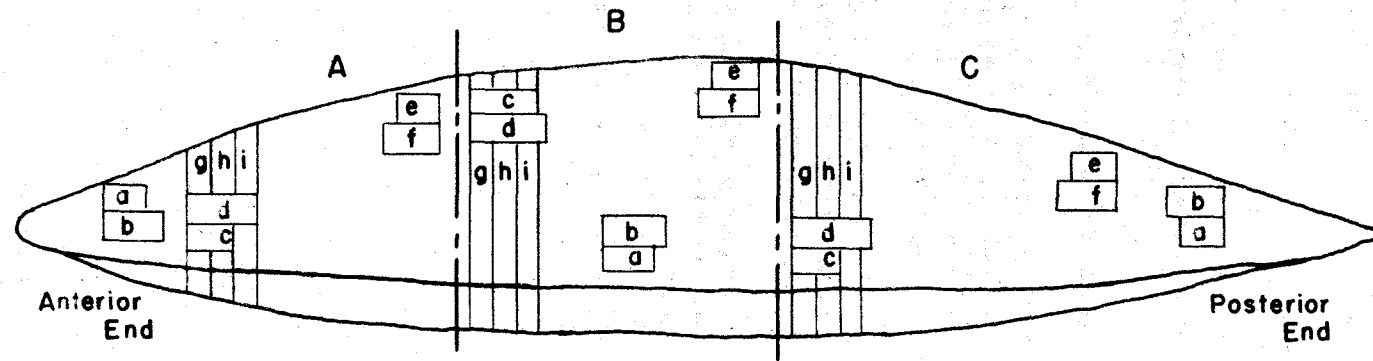


Figure 2.

Psoas Major and Psoas Minor Muscles.

Figure 3. Semitendinosus Muscle.

- A, B, and C - Cuts of beef for canning.
- a and b - Histological and pH samples, respectively, from muscle before division into cuts and after aging one day.
- c and d - Histological and pH samples, respectively, from beef cuts aged eight days.
- e and f - Histological and pH samples, respectively, from canned beef.
- g, h, and i - Slices for scoring by judges.

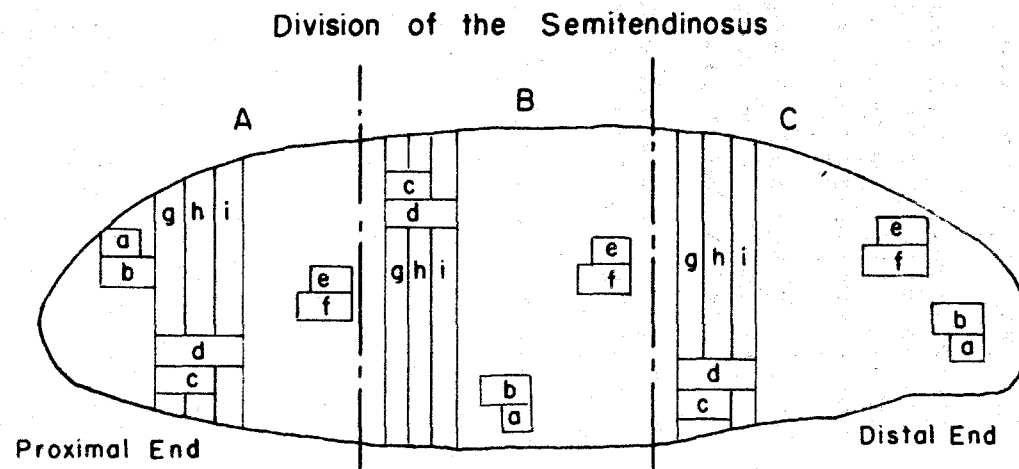


Figure 3.
Semitendinosus Muscle.

Figure 4. Semimembranosus Muscle.

A, B, C, D,

E, and F - Cuts of beef for canning.

a and b - Histological and pH samples, respectively, from muscle before division into cuts and after aging one day.

c and d - Histological and pH samples, respectively, from beef cuts aged eight days.

e and f - Histological and pH samples, respectively, from canned beef.

g, h, and i - Slices for scoring by judges.

Division of the Semimembranosus

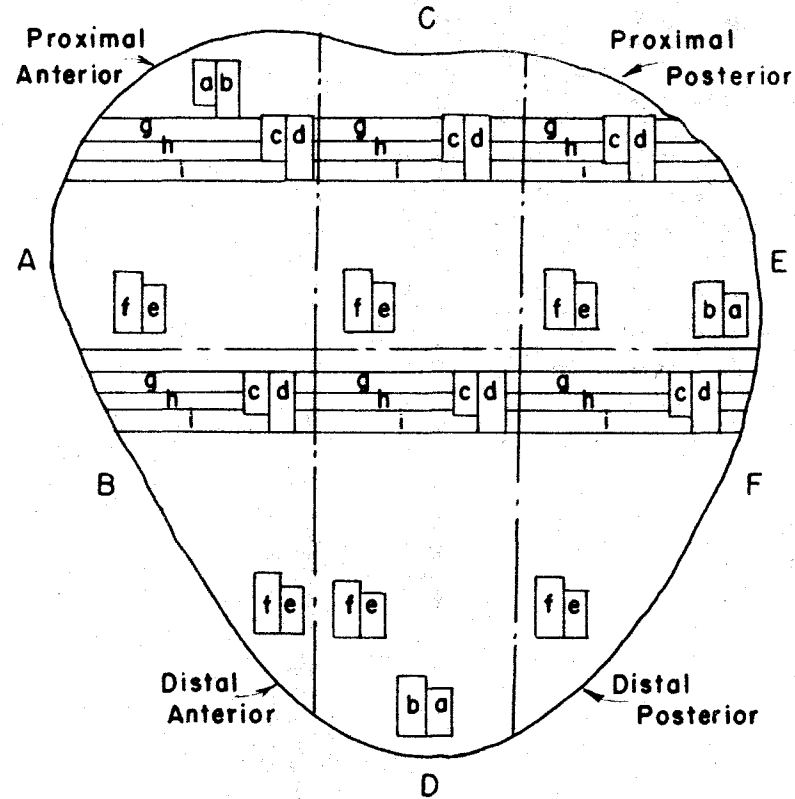


Figure 4.

Semimembranosus Muscle.

Figure 5. Biceps Femoris Muscle.

- A, B, C, D,
E, and F - Cuts of beef for canning.
- a and b - Histological and pH samples,
respectively, from muscle
before division into cuts
and after aging one day.
- c and d - Histological and pH samples,
respectively, from beef
cuts aged eight days.
- e and f - Histological and pH samples,
respectively, from canned
beef.
- g, h, and i - Slices for scoring by judges.

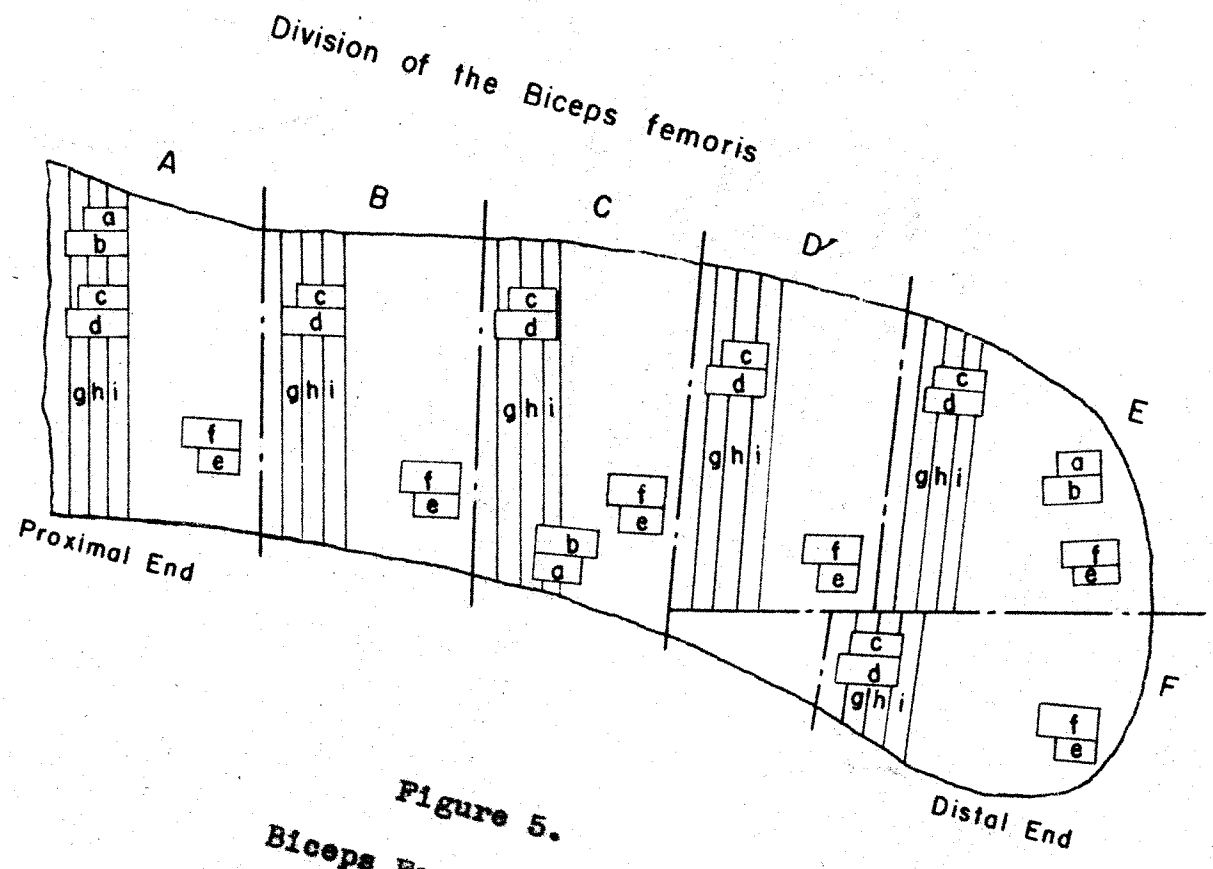


Figure 5.
Biceps Femoris Muscle.

size were obtained. The pieces were trimmed to approximately 600-gram weight (600 to 610 grams). When the piece was already smaller than the desired weight (in only a few cases), a second piece or "filler" of meat from the same animal was included to make a suitable total weight.

The weight of each of the matching cuts of meat, trimmed to size as indicated, was recorded and each cut was labeled to indicate sample number and orientation within the muscle. Glass beakers of 600-milliliter capacity had been provided for storage of the individual cuts of meat. The piece of meat in each case was put into the beaker so the anterior (or proximal) end was uppermost. Control samples (those from the left side of the carcass) were covered with moisture-vapor-proof cellophane held in place with a rubber band, and were placed in the cooler at 34° to 36°F. for aging. Samples to be injected (those from the right side of the carcass) were placed in the beakers and the injection was carried out, as described in another part of this paper, with the beaker serving as container for any solution that separated from the meat. The control samples were left uninjected so they would be representative of beef as it is usually canned.

Injection of Cuts of Meat

Equipment

The needles used for injection of the meat were made at the College Instrument Shop and consisted of 4 hypodermic needles mounted $3/4$ inch apart in the shape of a square. Each needle was approximately 3 inches long and had a bore of approximately $1/16$ inch. The instrument was screwed into the nozzle of a pressure pump (one regularly used by the Animal Husbandry Department for injecting hams and shoulders of pork). The pressure exerted by the pump was 30 pounds per square inch when the hand-control lever was pressed to the limit. By insertion of a wedge below this lever, the device was operated at a pressure slightly under 30 pounds.

Solutions used for injection

Three kinds of solutions were used for injection of the meat. The sodium chloride solution was made in 15 per cent strength by using the proportion of 150 grams of chemically pure sodium chloride made to 1000 milliliters with distilled water. The solution was stirred until the salt dissolved and was then filtered. Salimeter and temperature readings were taken. In each case the temperature was 22°C . (71.6°F .) and the salimeter reading showed a saturation of 51 per cent. (According to the statement on the instrument, 100 per cent

saturation equals 26.395 per cent sodium chloride, and the instrument is calibrated to show the saturation at 60°F.)

The lactic acid solution was made by adding lactic acid (85 per cent strength) to distilled water until the solution gave readings of pH 3.4 when tested with the pH meter. Very small amounts of lactic acid were added at one time and repeated readings were taken until the desired pH had been reached.

The solution containing a mixture of sodium chloride and lactic acid was made by using a portion of the 15 per cent sodium chloride solution already described and adding very small amounts of lactic acid (85 per cent strength) to it until repeated readings of the pH meter indicated a pH of 3.4 for the solution.

The quantity of each of the three solutions made for the treatment of the meat samples from each animal was 3500 milliliters (approximately 1 gallon). The solutions were stored in glass jugs in the refrigerator at 4°C. for 1 or 2 days. Several hours prior to use for injection of samples, the containers of solution were removed from the refrigerator and allowed to come to laboratory temperature (17° to 19°C.).

Method of injection

As has been stated, beef cuts from the right side of the

carcass were injected and the matching cuts from the left side were used uninjected as controls. Since three or six cuts had been secured from each muscle, it was possible to treat one cut from each muscle, or two cuts in the case of the large muscles, with each of the three solutions. For Animal VI, the particular cut from each muscle to receive a particular kind of treatment was determined by random selection; for succeeding animals, the order was rotated. The design for treatment of the cuts from each animal is shown in Table 1.

Preliminary experiments had indicated that injection to approximately 10 per cent increase in weight per cut would be a suitable working standard. Since the samples had been trimmed to weights within the range of 600 to 610 grams, it was decided to inject the samples so that the weight of sample plus injecting solution was 60 grams above that of the sample itself. Preliminary work had also shown that even though the liquid was injected into the meat, some of it tended to flow from the cut surfaces. By having the piece of meat in the beaker while it was being injected and during subsequent storage, this liquid was held in close contact with the meat, even though all of it did not stay within the interior.

With the beaker of meat resting on one pan of a laboratory balance and weights sufficient for a 60-gram increase on the other pan, the 4-needle hypodermic instrument was inserted

Table 1. Design for Injection of Beef Cuts. Position of sample in muscle is indicated in Figures 1, 2, 3, 4, and 5.

Animal no.	Sodium chloride		Lactic acid		Sodium chloride and lactic acid	
	Posi- tion	Sample no.	Posi- tion	Sample no.	Posi- tion	Sample no.
Longissimus dorsi, loin portion						
VI	A	162	B	163	C	164
VII	C	188	A	186	B	187
VIII	B	211	C	212	A	210
IX	A	234	B	235	C	236
X	C	260	A	258	B	259
Longissimus dorsi, rib portion						
VI	C	167	B	166	A	165
VII	A	189	C	191	B	190
VIII	B	214	A	213	C	215
IX	C	239	B	238	A	237
X	A	261	C	263	B	262
Psoas major and psoas minor						
VI	A	168	C	170	B	169
VII	B	193	A	192	C	194
VIII	C	218	B	217	A	216
IX	A	240	C	242	B	241
X	B	265	A	264	C	266
Semitendinosus						
VI	B	172	A	171	C	173
VII	C	197	B	196	A	195
VIII	A	219	C	221	B	220
IX	B	244	A	243	C	245
X	C	269	B	268	A	267
Semimembranosus						
VI	D	177	A	174	B	175
	F	179	E	178	C	176
VII	B	199	D	201	A	198
	C	200	F	203	E	202
VIII	A	222	B	223	D	225
	E	226	C	224	F	227
IX	D	249	A	246	B	247
	F	251	E	250	C	248
X	B	271	D	273	A	270
	C	272	F	275	E	274

(continued)

Table 1 (continued)

Animal no.	Sodium chloride		Lactic acid		Sodium chloride and lactic acid	
	Posi- tion	Sample no.	Posi- tion	Sample no.	Posi- tion	Sample no.
Biceps femoris						
VI	E	184	A	180	B	181
	F	185	C	182	D	183
VII	B	205	E	208	A	204
	D	207	F	209	C	206
VIII	A	228	B	229	E	232
	C	230	D	231	F	233
IX	E	256	A	252	B	253
	F	257	C	254	D	255
X	B	277	E	280	A	276
	D	279	F	281	C	278

lengthwise of the grain of the meat. Operation of the lever of the pressure pump as the instrument was slowly withdrawn permitted the solution to enter the meat. The needles were inserted three times in each cut of meat so that the resulting points of injection were about equally spaced. Usually it was necessary to add a little more solution at the top of the beaker to bring the weight to the desired figure. Occasionally it was necessary to pour off a small amount of the liquid to secure the proper weight of sample plus injecting solution. After injection of the sample, the container of meat was covered with moisture-vapor-proof cellophane and stored in the cooler beside the respective control sample.

Aging the Cuts

The control and injected cuts of beef were permitted to remain in the cooler at 34° to 36°F. for a period of 1 week after cutting (8 days after slaughter of the animal).

Canning the Meat

Equipment

The canning equipment was of commercial type and was located in the Food Processing Laboratory. The steamer, can sealer, and processing retort were operated by R. G. Tischer, Research Associate Professor of Horticulture. Plain tin cans, number 2 size, were used as containers for the meat. The lids had a rubber-like sealing compound at the rim.

Preparation of samples

The cuts of meat, after aging 8 days, were taken from the cooler, removed from the beakers, and individually weighed. The "drip" and/or surplus solution were discarded without weighing. From the anterior end of each cut, samples for histological study and for pH determinations were taken as previously described.

Since it was desired to have a minimum of 20 ounces (567 grams) of raw meat to put into each can, the piece was trimmed

to a weight within the range of 568 to 578 grams. In a few instances the "filler" of meat from the same animal was used to make up for the difference in weight. After a record of its weight was made, the cut of meat was put into the can with anterior end of the cut uppermost. A thermometer was inserted in each of four pairs of representative cans so that the bulb of the thermometer was near the center of the cut of meat. The square of cellophane which had been over the top of the beaker during storage was transferred to the top of the can to remain during the preheating of the meat before sealing. From each animal 48 cans of meat were used for processing plus some reserve cans, making a total of 56 for a retort load. This number of cans was sufficient to fill the retort to approximately one-third of its capacity.

Preheating the meat

The cans of meat resting on metal trays were loaded into a three-compartment steamer for preheating. The doors of the steamer were left slightly ajar so that atmospheric pressure would be maintained. A preliminary test had shown that approximately 1 hour of steaming was necessary for the meat at the center of the can to reach a temperature of 170°F. or higher. At the end of an hour of heating the cans were removed from the steamer, the thermometers read, the cellophane covers removed, and the cans closed on the automatic sealing machine.

Processing the meat

The cans of meat were immediately transferred to the processing retort and arranged with metal racks separating the tiers of cans. The retort was closed, the steam turned on, and the automatic device for recording temperature and time within the retort was adjusted. Processing was carried out at 10 pounds pressure (240°F.) for 65 minutes for the meat from animals VI, VII, and VIII; for 90 minutes in the case of Animals IX and X. Within a minute after opening the steam valve, the pressure had reached 10 pounds within the retort.

At the end of the processing period the steam was turned off and the valves were adjusted so that running cold water was circulated through the retort to cool the cans. The cooled cans were dried and two of the reserve cans were placed in an incubator at 100°F. for observation of keeping quality at an elevated temperature. All the other cans remained at room temperature overnight and then were transferred to the cooler for storage at 34° to 36°F. until they were opened for evaluation of the meat. The cans were stored in the cooler because it was the only convenient storage space available.

Evaluation of the Canned Meat

Preparation for evaluation

During an 8-day evaluation period, six cans of meat were

opened daily. The six cans opened on any particular day were from the same pair of muscles and represented the three kinds of injection. The order in which the meat from the various muscles was tested was determined by drawing lots. The design for the order of evaluation of canned beef samples is shown in Table 2. Each day the six cans to be opened on the following day were removed from storage in the cooler and were allowed to come to room temperature.

Table 2. Design for the Evaluation of Canned Beef Samples

Animal no.	Control and injected samples evaluated each day							
	1st	2d	3d	4th	5th	6th	7th	8th
VI	168	180	165	174	162	171	181	175
	169	183	166	176	163	172	182	177
	170	185	167	179	164	173	184	178
VII	192	189	186	200	198	195	204	206
	193	190	187	202	199	196	205	207
	194	191	188	203	201	197	208	209
VIII	213	228	216	210	219	224	230	222
	214	229	217	211	220	226	231	223
	215	232	218	212	221	227	233	225
IX	243	234	253	240	237	246	252	247
	244	235	254	241	238	248	255	249
	245	236	256	242	239	251	257	250
X	278	261	272	270	267	258	276	264
	279	262	274	271	268	259	277	265
	281	263	275	273	269	260	280	266

Before a can was opened, a measurement was taken of the vacuum developed within the can by inserting a vacuum gauge through the lid. Following this measurement the can was opened and the liquid was drained into a bowl for weighing. A portion of the liquid (approximately 35 milliliters) was poured into a screw-topped glass jar and put into the refrigerator for storage at 4°C. until the following day when it was observed for the amount of gelation which had occurred. The remainder of the liquid was poured into a 400-milliliter beaker for observation of the color and general appearance, and for sampling by the judges when scoring flavor of the liquid. The meat from the can was weighed in the same bowl that had been used for the juice, samples for histological study and for pH determination were removed from the piece of meat, and the meat was ready for slicing.

Slicing the meat

A hand-operated mechanical slicer with rotary blade was used to slice the meat from each can. Slices were cut 3/8 inch in thickness. Slicing was begun at the anterior end of the cut of meat, the end uppermost in the can. Individual squares of waxed paper were placed on the platform of the slicer to receive each slice as it came from the blade. The papers were numbered to show the sequence of the slices. Even when the meat was so crumbly as to not give a true slice,

whatever amount of meat came through the machine with one revolution of the blade was placed upon the respective paper and was later counted in determining the "possible number of slices." The slicings from each can of meat were arranged consecutively on a tray for counting the number of slices obtained and for observation of the quality of the slices.

In determining the count of slices, the meat was considered to be a slice if it held together for approximately three-fourths or more of its area. Doubtful cases of "holding together" were tested by picking up the slice with a wire clamp and suspending the slice vertically for about 5 seconds. Distinction was made between slices as to quality, i.e., if they held together for almost the entire area they were classed as firm slices; if they crumbled for about one-fourth of their area they were classed as crumbly slices. Crumbliness more extensive than this meant that the meat was not counted as a "slice obtained" but was considered to be part of the "unsliceable portion." Weight of the total unsliceable portion was recorded for each can and the count of these very crumbly slices was considered in determining the "possible number of slices" from the can. While the slicings from each can of meat were still spread out on the trays, observations were made of the color, iridescence, and general appearance of the meat.

Scoring the meat

Slices obtained from certain relative positions in the cut of meat were presented to each of the three judges for scoring. Judge number 1 received the second slice (counting from the anterior end of the cut), judge number 2 the third slice, and judge number 3 the fourth slice (indicated as g, h, and i, respectively, in Figures 1, 2, 3, 4, and 5). Each slice was individually wrapped in waxed paper, marked with a code number, and placed in an individual plastic bag. One such slice from each of the six cuts of meat to be judged that day was presented to each judge for scoring and each judge poured some of the liquid from the beakers into a cup for tasting. Both the meat and the liquid were at room temperature when scored. Sliceability and the desirability of the following six palatability factors were scored by the judges: aroma, flavor or meat, flavor of liquid, tenderness, juiciness, and texture. The scoring range for each factor was a score of 10 for extremely good quality to a score of 1 for extremely poor quality. A copy of the score sheet is given in the Appendix.

Gelation of liquid

Observations were made of the amount of gelation that had occurred in the liquid during refrigeration for approximately 24 hours. The jars were taken from the refrigerator, the screw

cap was removed and the contents of the jar were turned onto a plate. If gelation had taken place, a thin-bladed knife was run once around the interior of the jar to loosen the jellied material from the glass. The rigidity of the gel as it rested on the plate and as a knife was cut through it was observed. If the material was still liquid, the consistency was observed.

Measurement of pH

At three different stages there were samples removed for pH determinations, namely, (1) from three positions in the muscle 1 day after slaughter of the animal and before the muscle was divided into cuts, (2) from the anterior end of the beef cuts on the 8th day after slaughter, and (3) from the posterior end of the piece of canned meat before the slicing was done. The positions within the muscle from which these samples and also those for histological study were taken are shown in the diagrams of the respective muscles (Figures 1, 2, 3, 4, and 5). Each 10-gram sample for pH determination was wrapped in moisture-vapor-proof cellophane and the samples for 1 day were placed in a bag. Samples were stored temporarily (about 2 hours) in the refrigerator at 4°C., transferred to the quick-freezing compartment at the Meats Laboratory for freezing at -30°F., and finally stored at 0°F. until the 13th or 14th day, at which time the samples were removed for measurement of the pH value.

A Coleman pH meter was the instrument used for this determination. The samples were prepared for testing by removing from zero storage, placing a few at a time at room temperature (leaving the remainder in the refrigerator), cutting four or five times through the 10-gram sample with a sharp knife, and exposing the cut pieces to the air for a few minutes until the meat was partially thawed. The pieces were placed in the stainless steel cup of a Waring blender. Distilled water was added (25 milliliters for the raw meat, 30 milliliters for the canned) and the blender was run for 30 seconds. This procedure resulted in maceration of the sample and elevation of the temperature to approximately 25°C. Some of the mash was placed in the cup of the pH meter and the reading for the sample was taken.

Histological study

Samples from the longissimus dorsi muscle, rib portion, were selected for histological study. The judges' ratings of palatability factors had shown greater differences between control and injected samples for this muscle than for the other muscles. Histological sections were cut either 25 or 30 microns thick on the freezing microtome. Weigert's stain for elastic connective tissue and Van Gieson's stain for collagen were used to selectively stain these compounds. The muscle fibers appeared yellow-green to reddish orange, the

elastic tissue blue-black, and the collagen bright pink with this staining technique. The nuclei were not differentiated; fat deposits were not stained but could be detected by the shape of the colorless areas within the connective tissue.

RESULTS AND DISCUSSION

The purpose of the investigation, as stated earlier in this paper, was to study the effects on certain physical and organoleptic properties of canned beef of injecting the raw meat with solutions of sodium chloride, of lactic acid, or of sodium chloride and lactic acid. For each experimental cut of beef injected with appropriate solution, there was an uninjected control sample from the corresponding position on the opposite side of the animal. Comparisons are made between the results for the matching pairs of samples.

Detailed tables of data are given in the Appendix as follows: Table 1, Palatability of Canned Beef; Tables 2 and 3, Slicing Quality of Canned Beef; Tables 4 and 5, pH of Beef; Table 6, Weight Changes before Canning; Table 7, Weight of Canned Meat and of Liquid; Tables 8 to 22, inclusive, analyses of variance of canned beef scores for flavor, tenderness, juiciness, texture, and slicing quality. Summary tables and graphs of the results are included under the heading, Results and Discussion.

Palatability of Canned Beef

The palatability of the canned beef was determined by judges' scores for six characteristics of the meat: aroma, flavor of meat and of liquid, tenderness, juiciness, and texture. The judges were experienced in evaluating meat without the addition of table salt during or after cooking. The three judges were consistent in their ratings of palatability of the meat. This was shown by statistical analysis by individual judges of some of the scores given to samples of the canned beef. The uniformity of scoring was sufficiently high that for the remainder of the statistical part of the study, the results were analyzed on the basis of total scores, rather than by scores of individual judges.

The results for four of the palatability factors (flavor, tenderness, juiciness, and texture) and for the judges' rating of slicing quality were selected for statistical analysis. Other results are considered from the standpoint of trends shown, but not on the basis of statistical significance. For purposes of treating the results statistically, Animals VI, VII, and VIII, the meat of which was processed for 65 minutes, were grouped together; Animals IX and X, although of different carcass grade but with the same processing time of 90 minutes, were placed in a second group.

Aroma

The averages of the aroma scores for the canned beef from Animals VI, VII, and VIII were nearly identical for control and injected samples for the three kinds of injection. These average values for aroma, as shown in Table 3, lie within the narrow range of 7.7 to 8.1. For the meat of Animal IX, which was processed for 90 minutes, the average scores for aroma were slightly lower than for the meat for the three animals processed 65 minutes. These data were not analyzed statistically and the differences may be too small to be significant. The meat of Animal X was judged to be less desirable in aroma than that of the other four animals. The average values for control and injected samples for the three kinds of treatment, as shown in Table 3, were within the range of 6.1 to 6.6. Animal X was the aged dairy cow (carcass grade Cutter) and the meat, even before canning, was noted to have a stronger odor than that of the other animals. The injected samples for Animal X had slightly higher average scores for aroma than the control for each of the three treatments, but the differences are probably too small to be significant.

Thus, on the basis of judges' scores, it is evident that injection of the meat had little or no effect on the aroma of the canned meat. The samples processed 90 minutes had

Table 3. Averages of Palatability Scores.

Animal* no.	No. of muscles	Palatability scores					
		Aroma	Flavor		Tender- ness	Juici- ness	Tex- ture
			Meat	Liquid			

SODIUM CHLORIDE INJECTION

Control samples

VI	6	7.9	7.3	6.4	7.1	5.7	5.7
VII	6	8.0	7.5	6.6	7.4	6.4	6.3
VIII	6	7.7	7.6	6.1	7.2	5.9	5.8
Av. (3 animals)		7.9	7.5	6.4	7.2	6.0	5.9
IX	6	7.5	7.2	5.7	7.8	4.5	5.7
X	6	6.2	4.9	4.2	7.3	4.4	4.7
Av. (2 animals)		6.8	6.0	5.0	7.6	4.4	5.2
Av. (5 animals)		7.5	6.9	5.8	7.4	5.4	5.6

Injected samples

VI	6	7.8	8.0	7.2	8.1	6.7	7.1
VII	6	7.8	8.2	7.5	8.6	6.6	7.2
VIII	6	7.9	8.1	7.2	8.0	6.6	6.9
Av. (3 animals)		7.8	8.1	7.3	8.2	6.6	7.1
IX	6	7.4	7.9	7.1	8.7	4.8	6.8
X	6	6.6	5.7	5.2	8.7	5.4	6.1
Av. (2 animals)		7.0	6.8	6.2	8.7	5.1	6.4
Av. (5 animals)		7.5	7.6	6.8	8.4	6.0	6.8

(continued)

*Animals VI, VII, and VIII were steers, carcass grade Commercial, processed 65 minutes; Animal IX was a steer, carcass grade Commercial, processed 90 minutes; Animal X was a cow, carcass grade Cutter, processed 90 minutes.

Table 3 (continued)

Animal no.	No. of muscles	Palatability scores					
		Aroma	Flavor		Tender-ness	Juici-ness	Tex-ture
			Meat	Liquid			
SODIUM CHLORIDE AND LACTIC ACID INJECTION							
Control samples							
VI	6	7.9	7.5	6.5	7.7	6.3	6.2
VII	6	8.0	7.6	6.8	7.6	6.3	6.3
VIII	6	7.7	7.6	6.2	7.0	6.2	6.0
Av. (3 animals)		7.9	7.6	6.5	7.4	6.3	6.2
IX	6	7.4	7.2	5.6	7.6	4.5	5.7
X	6	6.2	5.2	4.0	7.4	4.4	4.8
Av. (2 animals)		6.8	6.2	4.8	7.5	4.4	5.2
Av. (5 animals)		7.4	7.0	5.8	7.5	5.5	5.8
Injected samples							
VI	6	8.0	8.0	7.4	8.4	6.9	7.6
VII	6	7.9	8.2	7.8	8.6	6.8	7.1
VIII	6	8.0	8.2	7.5	8.3	6.7	7.4
Av. (3 animals)		8.0	8.1	7.6	8.4	6.8	7.4
IX	6	7.5	8.0	7.2	8.9	5.1	7.0
X	6	6.4	5.8	5.3	8.9	4.9	6.2
Av. (2 animals)		7.0	6.9	6.2	8.9	5.0	6.6
Av. (5 animals)		7.6	7.6	7.0	8.6	6.1	7.1

(continued)

Table 3 (continued)

Animal no.	No. of muscles	Palatability scores					
		Aroma	Flavor		Tender-ness	Juici-ness	Tex-ture
			Meat	Liquid			
LACTIC ACID INJECTION							
Control samples							
VI	6	8.1	7.4	6.4	7.6	6.3	6.6
VII	6	7.8	7.6	6.9	7.6	6.2	6.0
VIII	6	7.7	7.5	6.1	6.7	5.6	5.6
Av. (3 animals)		7.9	7.5	6.5	7.3	6.0	6.1
IX	6	7.5	7.3	5.5	7.8	4.8	5.9
X	6	6.1	5.0	3.9	7.2	4.2	4.5
Av. (2 animals)		6.8	6.2	4.7	7.5	4.5	5.2
Av. (5 animals)		7.4	7.0	5.8	7.4	5.4	5.7
Injected samples							
VI	6	8.0	7.7	6.6	7.7	6.5	6.8
VII	6	7.9	7.4	6.7	7.4	6.1	6.2
VIII	6	7.8	7.6	6.1	6.5	5.7	5.4
Av. (3 animals)		7.9	7.6	6.5	7.2	6.1	6.1
IX	6	7.4	7.2	5.6	7.6	4.5	5.7
X	6	6.4	5.2	4.5	7.7	4.8	5.2
Av. (2 animals)		6.9	6.2	5.0	7.6	4.6	5.4
Av. (5 animals)		7.5	7.0	5.9	7.4	5.5	5.9

slightly lower average scores for aroma of the canned beef than those processed 65 minutes. The scores for meat from the aged dairy cow were markedly lower than for the other animals, and only a slight improvement in aroma was noted as a result of injection with any of the three solutions.

Flavor

An improvement in the flavor of the meat injected with either sodium chloride solution or with sodium chloride and lactic acid solution, in comparison with the control samples, is shown by the data in Table 3 and Figures 6 and 7. An analysis of variance of the results shows that the differences were highly significant.

The flavor differences between muscles were not significant for either group of animals, nor between animals of the first group. Within the second group, composed of Animals IX and X, the flavor differences between the meat of the two animals were highly significant. The meat of Animal X, the aged dairy cow, was strong in flavor. The average scores for this factor, as shown in Table 3, are 5.7 and 4.9, respectively, for sodium chloride-injected and the control samples, whereas, for Animal IX the values are 7.9 and 7.2, respectively. Similarly, for samples treated with combined sodium chloride and lactic acid, Animal X had values of 5.8 and 5.2, respectively, for injected and control samples, compared to

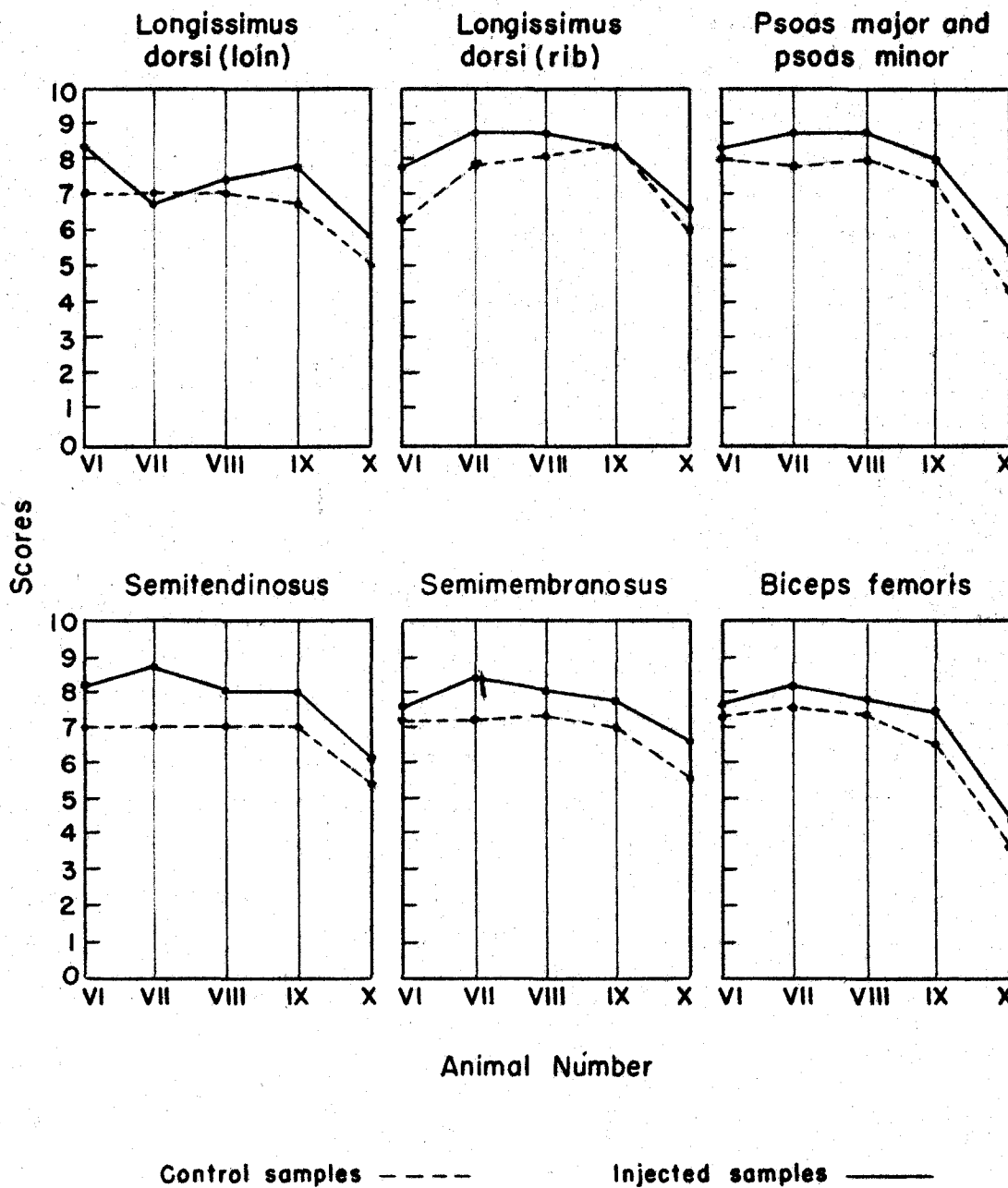


Figure 6. Scores for Flavor of Canned Beef.
Sodium chloride injection.

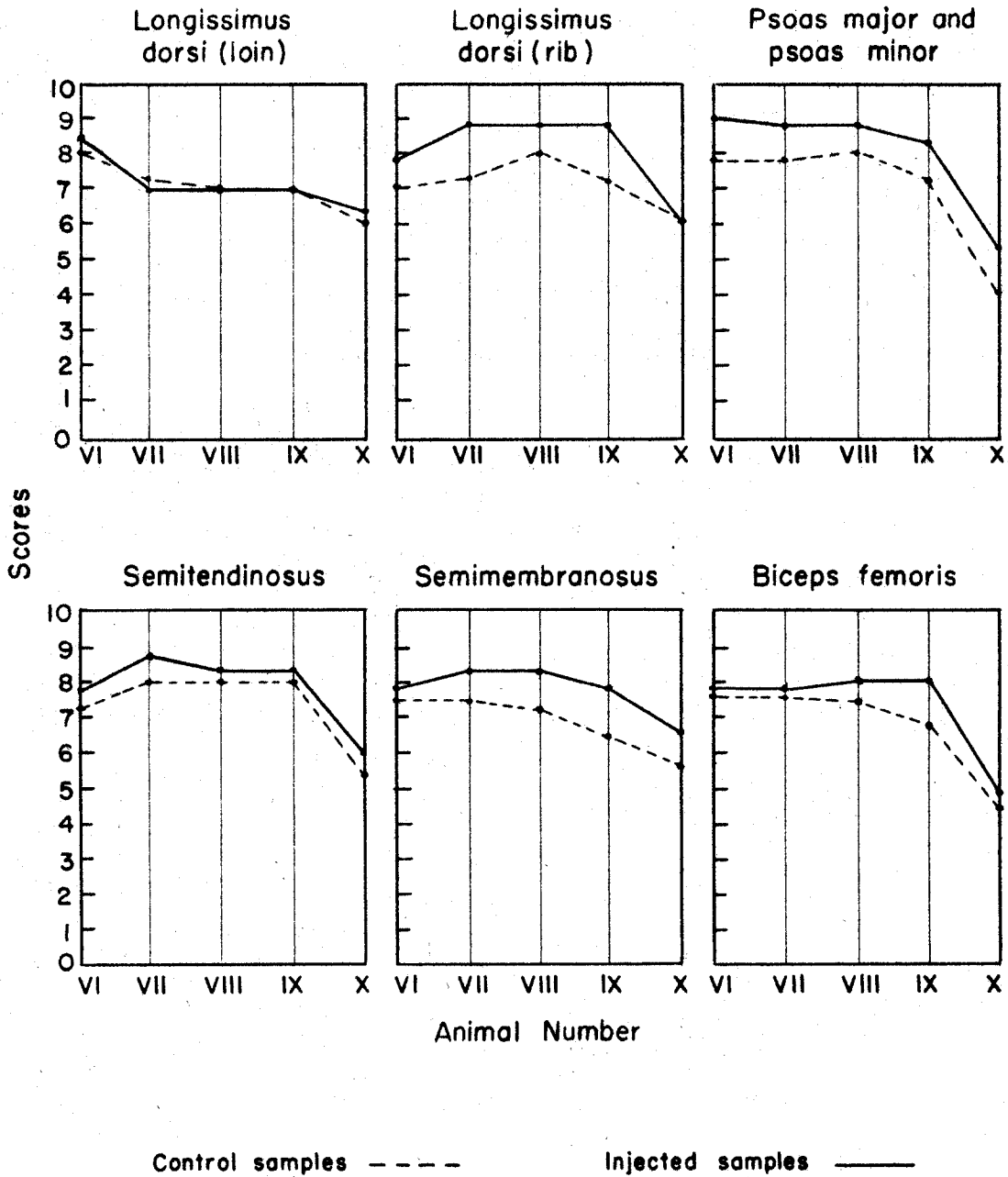


Figure 7. Scores for Flavor of Canned Beef.
Sodium chloride and lactic acid injection.

8.0 and 7.2 for Animal IX.

Lactic acid injection made no significant difference in the flavor of control and injected samples of beef for either group one or group two (Figure 8). The flavor differences between Animal IX and Animal X were highly significant, as was true in the results for the other two kinds of injection.

Thus, injection of the raw meat with either sodium chloride or with sodium chloride-lactic acid solution was shown to have a favorable effect on the flavor of the canned meat, the differences between injected and control samples being highly significant. Injection with lactic acid, on the other hand, made no significant difference in the judges' rating of the flavor of the canned meat.

Flavor of the liquid

The results for the flavor of the liquid show the same general pattern as those for the flavor of the meat, but the level of scores is markedly lower for the liquid than for the meat, as shown by the average values in Table 3. Comments of the judges as to flavor of the liquid included such descriptive terms as "metallic" and "bitter." For the older animal, the liquid was said to have a "very strong flavor." For the samples treated with either solution containing sodium chloride, the judges repeatedly commented that the juice was

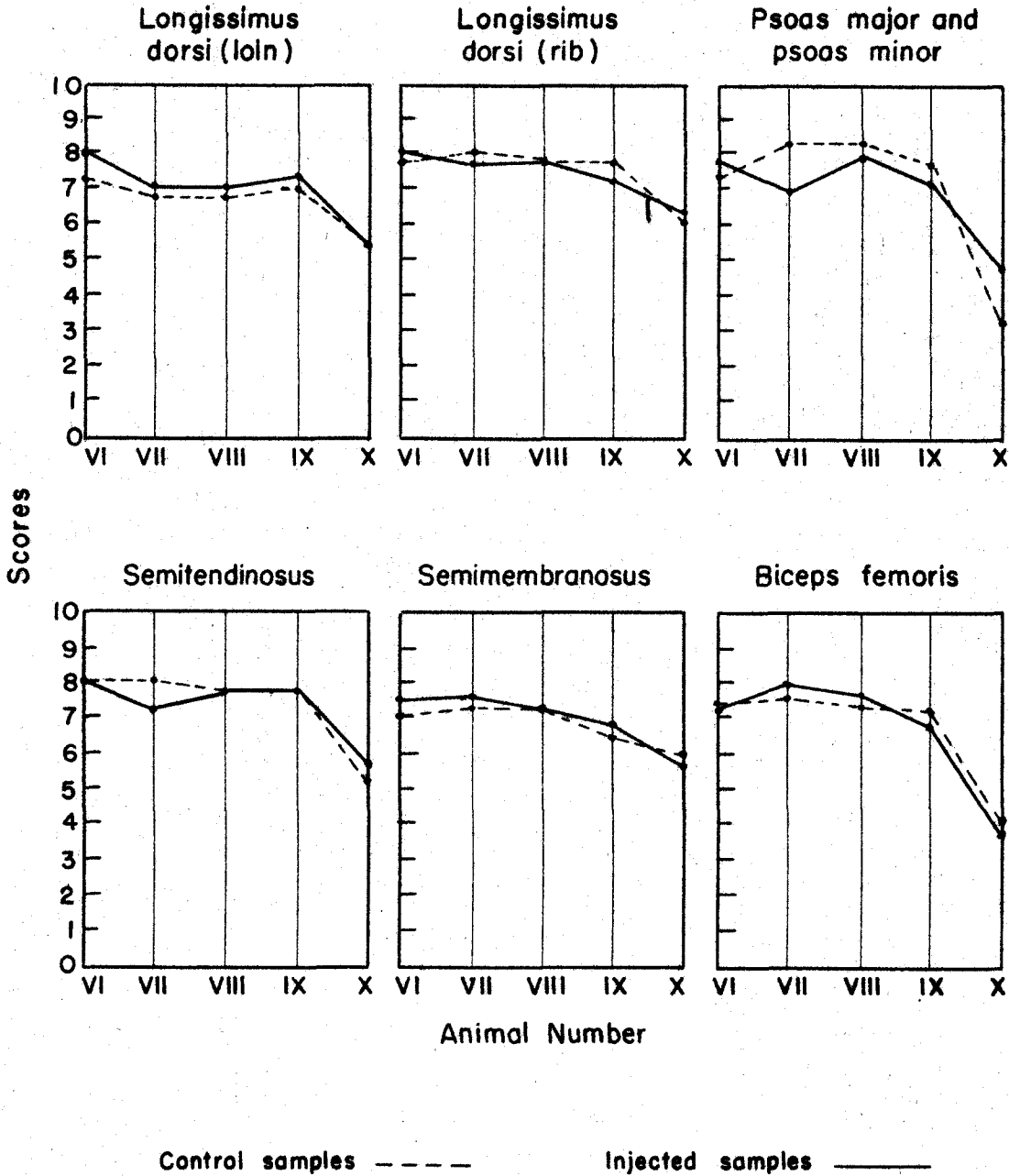


Figure 8. Scores for Flavor of Canned Beef.
Lactic acid injection.

"salty" or "a little too salty," whereas the meat was not similarly designated. Even though the flavor of the liquid was often indicated as salty, the average scores for flavor of the liquid (Table 3 and Figures 9 and 10) were higher for injected than for control samples for either of the solutions containing salt. The lactic acid treatment had no consistent effect on the flavor of the liquid, as shown in Table 3 and Figure 11.

Tenderness

The tenderizing effect of both sodium chloride and sodium chloride-lactic acid treatment is shown by judges' scores for tenderness of the canned beef (Table 3). The analysis of variance indicates that the differences between injected and control samples were highly significant. Lactic acid injection had no significant tenderizing effect on the meat.

For samples from Animals VI, VII, and VIII with sodium chloride injection, there was no significant difference in tenderness among animals, but there was a difference, significant at the .05 level, among the muscles. The psoas muscles received high scores for tenderness; the longissimus dorsi, loin portion, was rated lower, in general, than the other muscles of these three animals. The average tenderness scores for control samples of Animals IX and X (Table 3 and Figure 12) were lower for the aged dairy cow than for the

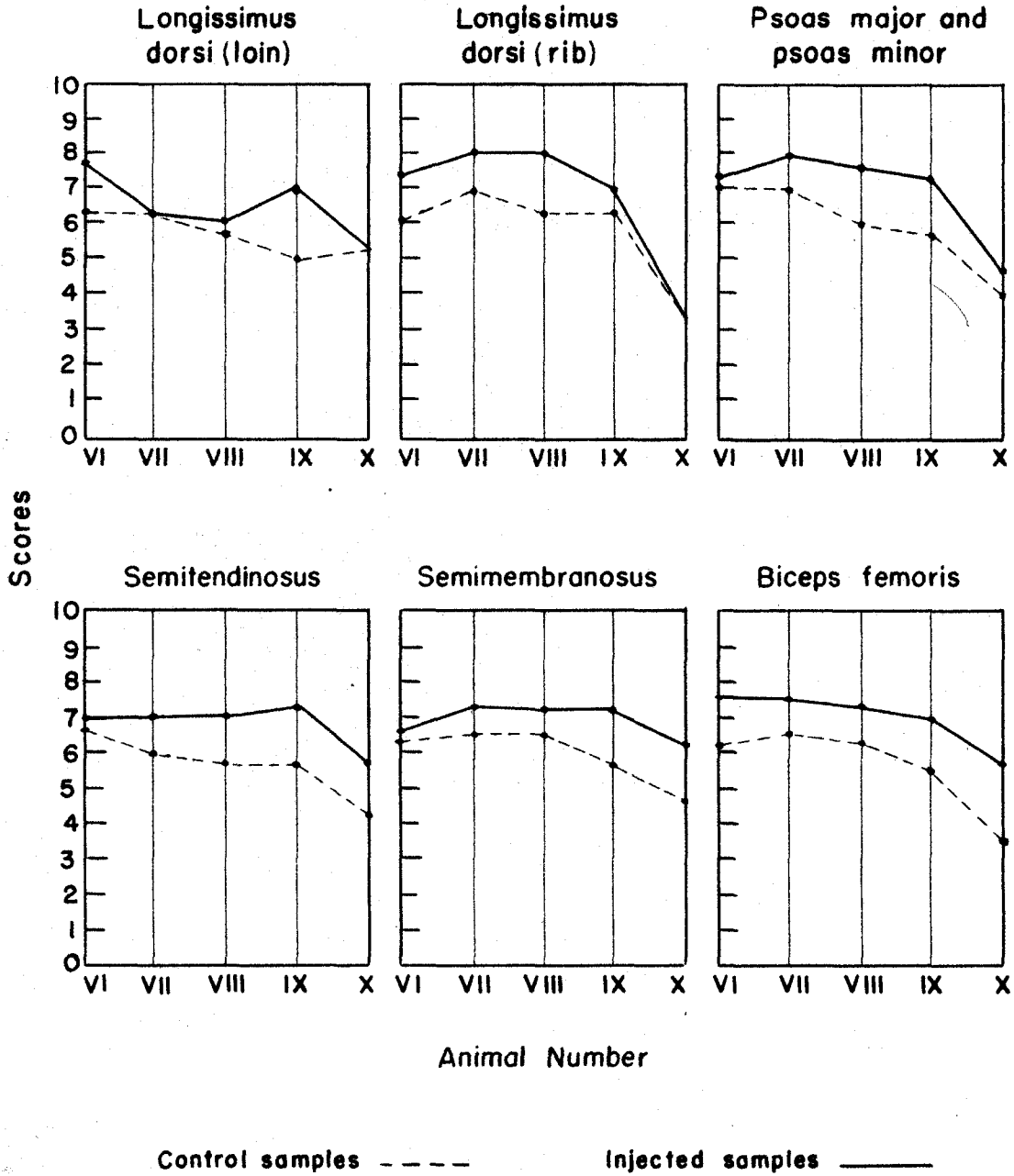


Figure 9. Scores for Flavor of Liquid from Canned Beef. Sodium chloride injection.

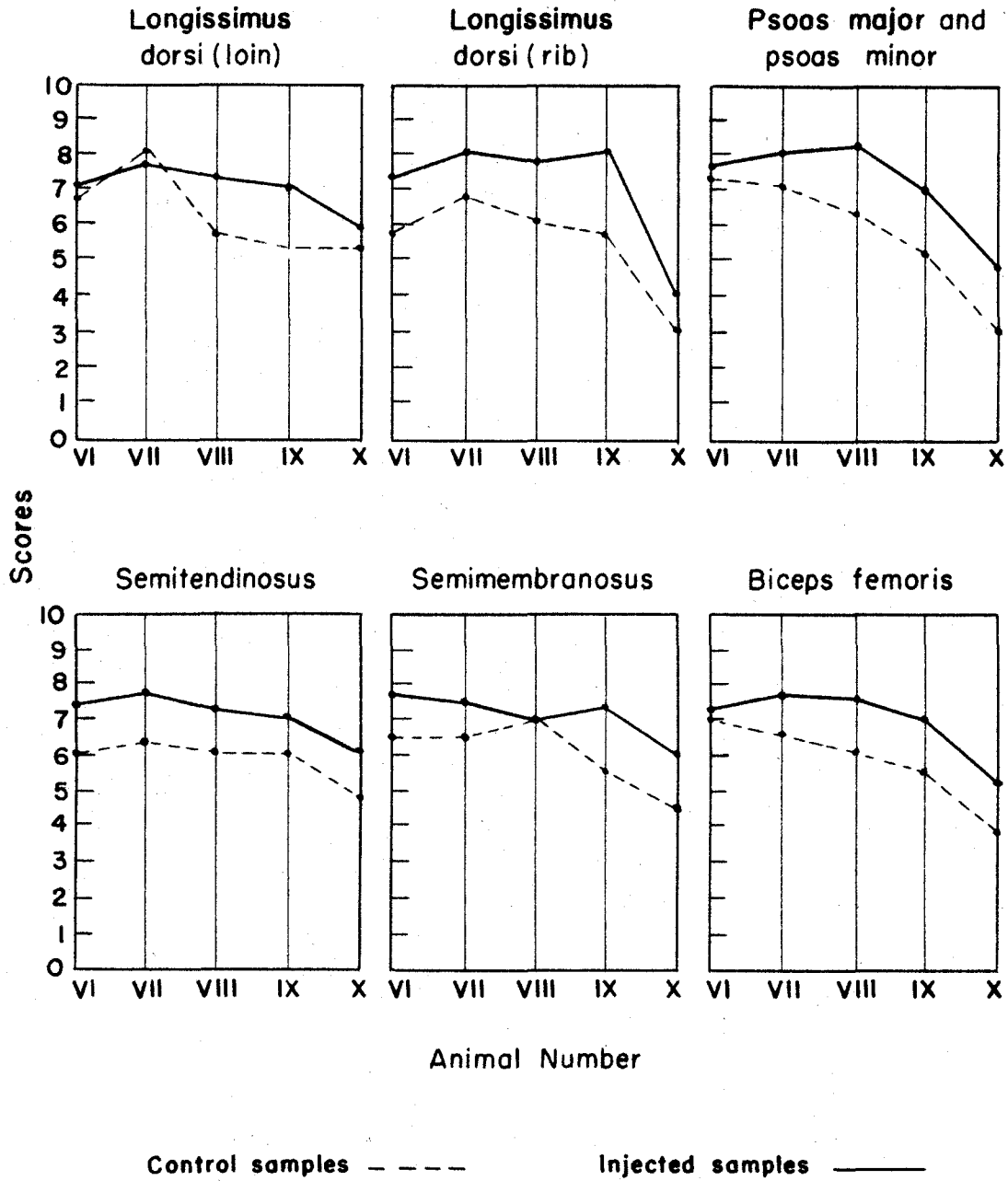


Figure 10. Scores for Flavor of Liquid from Canned Beef. Sodium chloride and lactic acid injection.

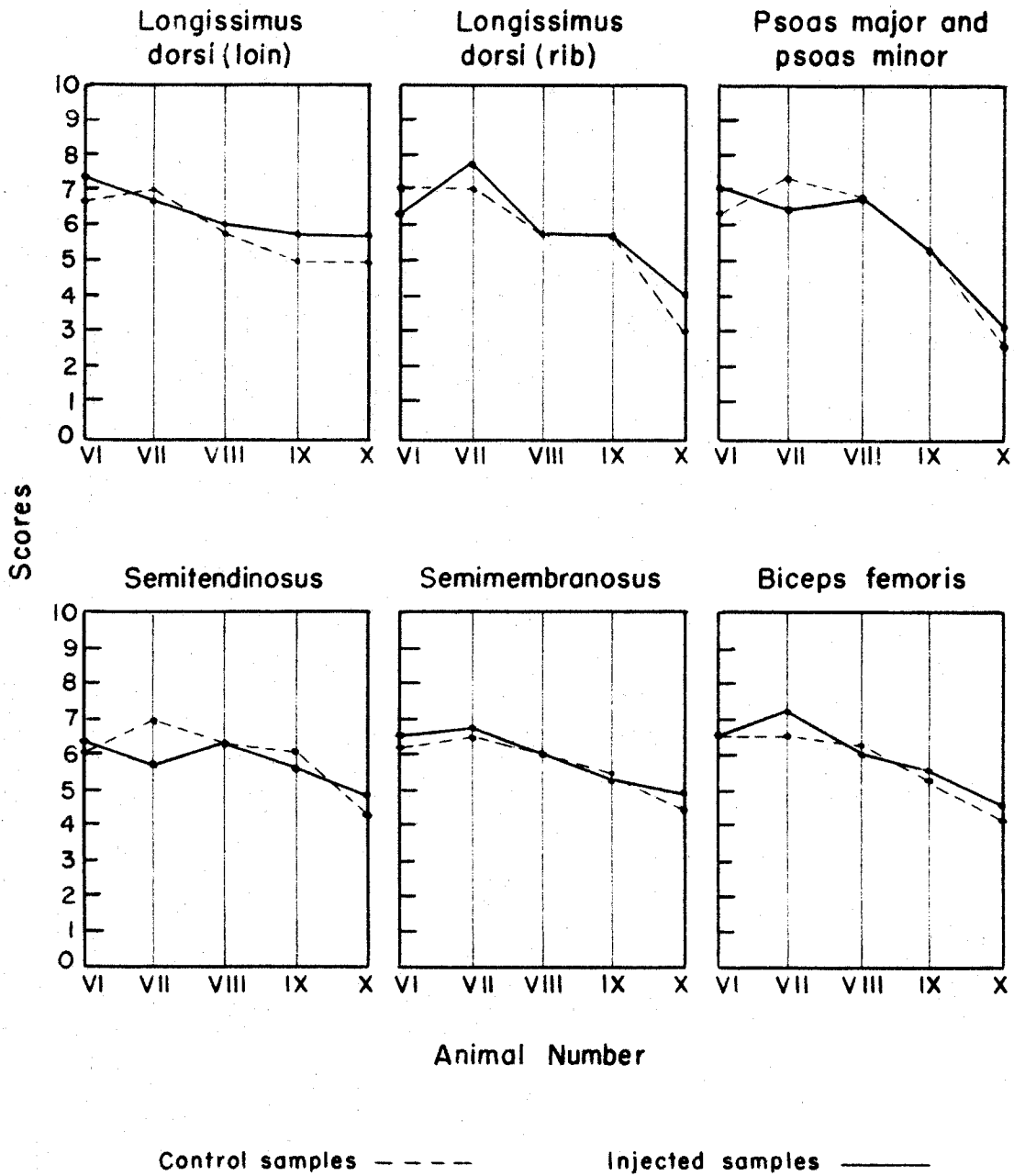


Figure 11. Scores for Flavor of Liquid from Canned Beef. Lactic acid injection.

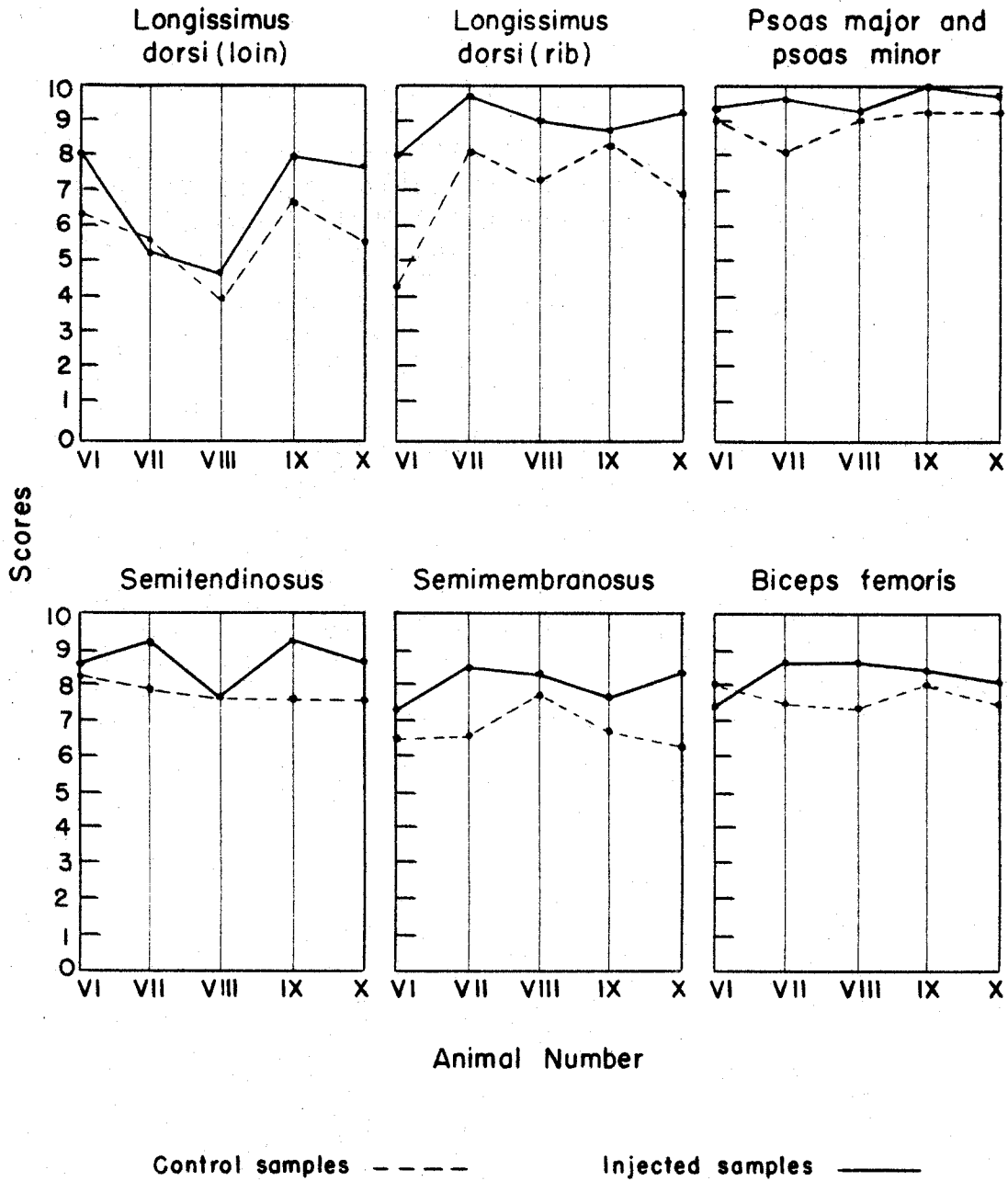


Figure 12. Scores for Tenderness of Canned Beef. Sodium chloride injection.

beef steer. The meat from both animals was made more tender by injection with sodium chloride solution and the differences owing to treatment were highly significant. The tenderizing effect was more marked for the meat from the older animal than for the meat from the younger animal.

Results for sodium chloride and lactic acid in combination were similar to those for sodium chloride alone; i.e., the injected cuts for both groups of animals received higher average tenderness ratings than the control samples (Table 3 and Figure 13. These differences were highly significant. The differences in tenderness of muscles for the first group of animals were significant at the .05 level; for the second group, at the .01 level. The psoas muscles ranked highest, and the loin portion of the longissimus dorsi muscle lowest, in the average tenderness scores among the five animals. The meat of the aged dairy cow would be expected to be less tender than the meat of the steer, but for the samples included in the sodium chloride-lactic acid treatment, no significant differences in tenderness between animals were shown. This may be explained on the basis of the long processing period (90 minutes) for Animals IX and X during which time the connective tissue may have become softened in the meat of both animals. A second factor would be the small number of animals in the group. The differences would have to be large

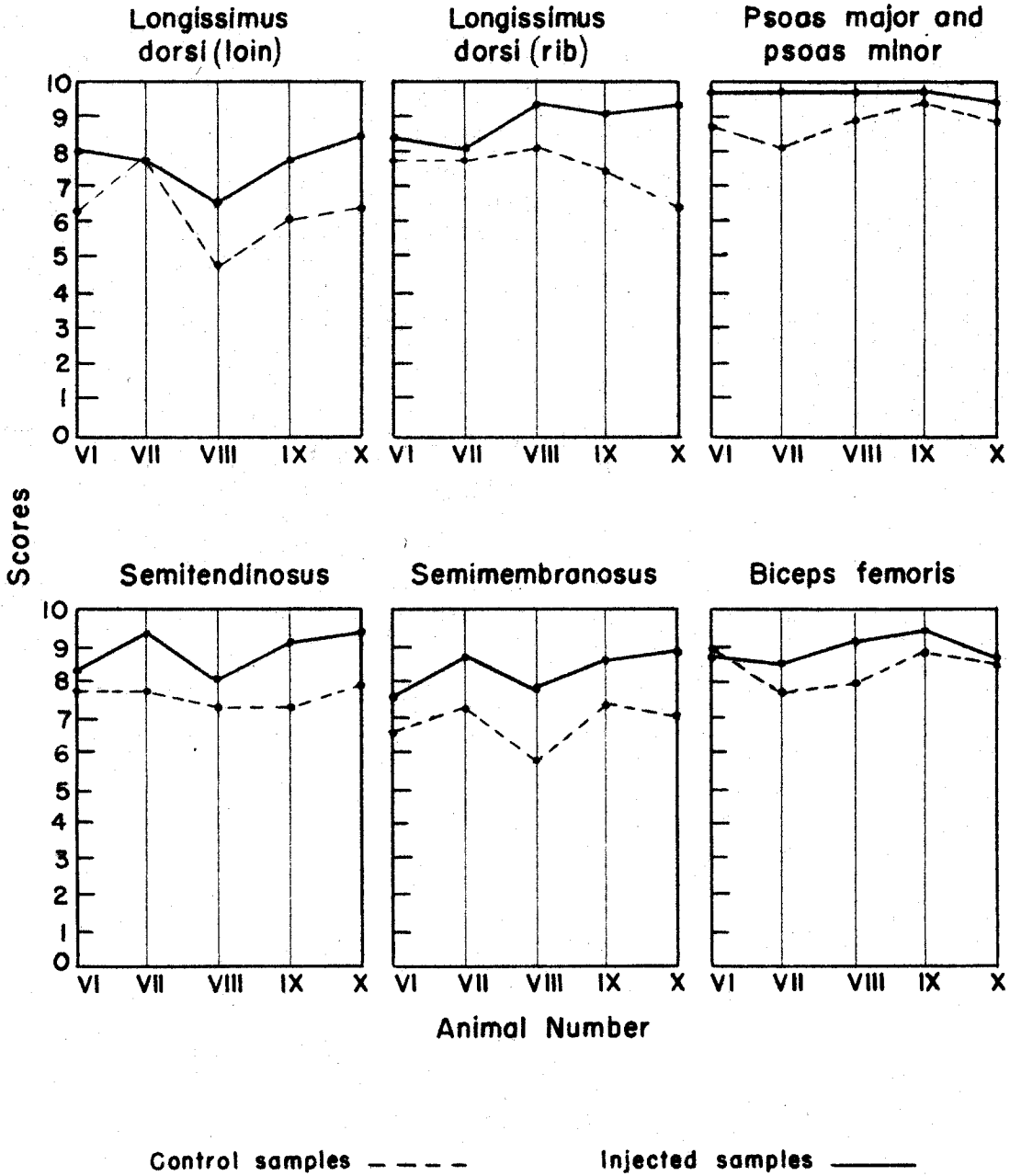


Figure 13. Scores for Tenderness of Canned Beef. Sodium chloride and lactic acid injection.

to be classed as significant with only two animals in the group.

Although the lactic acid injection of beef did not have a significant effect on the tenderness of the meat (Figure 14), the data showed there were differences in tenderness among muscles that were significant at the .01 level for Animals VI, VII, and VIII, but only at the .05 level for Animals IX and X. Differences among animals were highly significant for the first group of animals, but not significant for the second group. The same factors, i.e., the long processing period and the small number of degrees of freedom in the statistical analysis of the results, may explain the fact that the differences in tenderness between Animals IX and X were too small to be significant.

In general, it may be said that the tenderness of meat, as revealed by judges' scores, indicated differences among muscles and to some extent among animals. Injection of the raw meat with sodium chloride solution or with combined sodium chloride and lactic acid solution resulted in improved tenderness of the canned beef.

Juiciness

Judges' scores for juiciness of the canned beef showed no significant differences due to animal variation for either

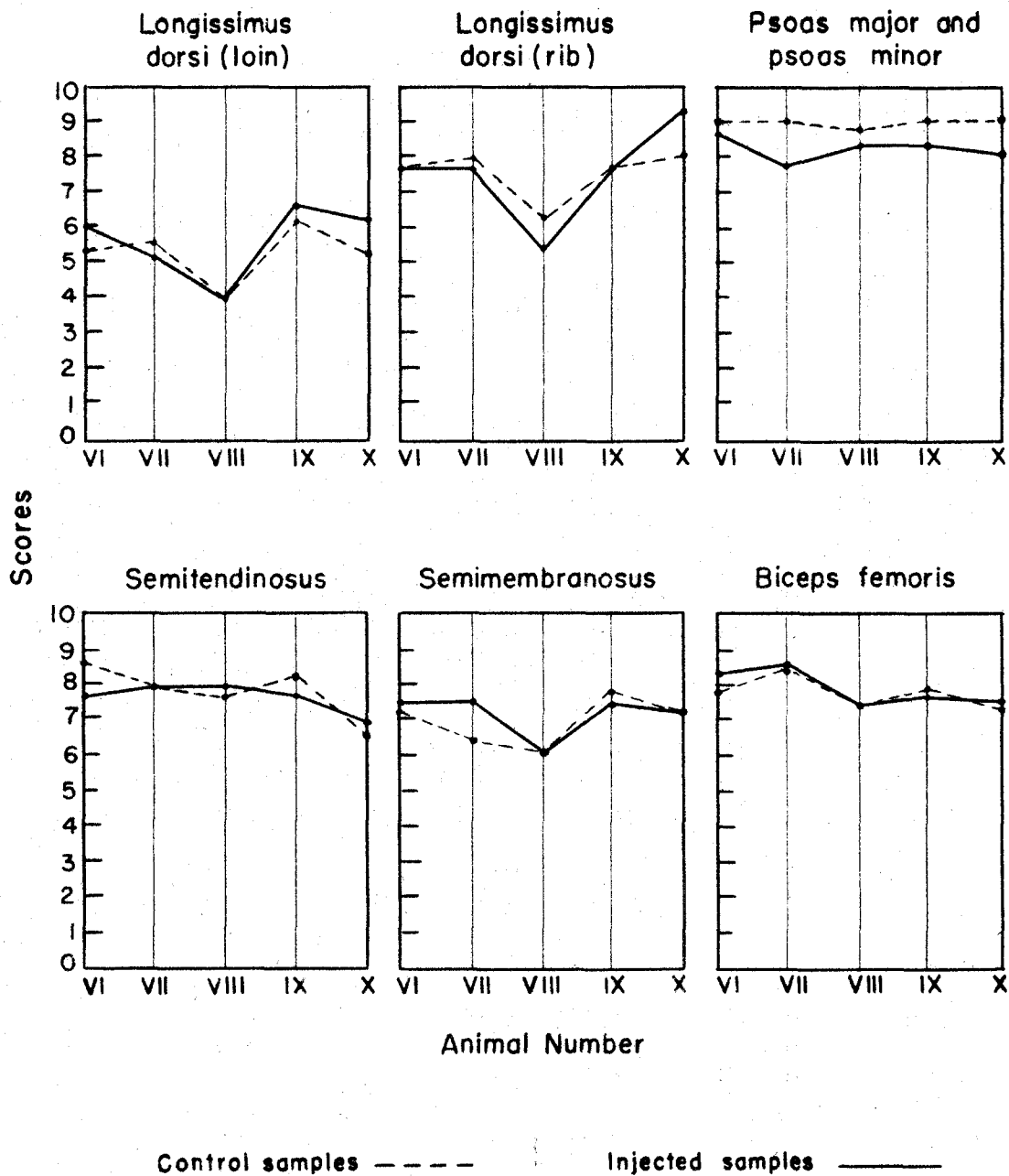


Figure 14. Scores for Tenderness of Canned Beef.
Lactic acid injection.

group. The muscle variation for Animals VI, VII, and VIII was not significant for samples included in the tests with sodium chloride but was highly significant for the samples used with the other two kinds of treatment. Muscle differences were not significant for Animals IX and X.

In general, the judges' rating of juiciness of the canned beef tended to be medium or fairly low. Average scores ranging from 6.9 to 5.6 for animals processed 65 minutes and from 5.4 to 4.2 for animals processed 90 minutes are shown in Table 3. These results are shown graphically in Figures 15, 16, and 17.

Injection of the raw meat with sodium chloride solution improved the juiciness of the samples processed for 65 minutes, but had no significant effect on those processed 90 minutes.

The improvement in juiciness brought about by injection of the meat with the combination of sodium chloride and lactic acid solution was significant at the .05 level for both groups of animals. In the meat processed 65 minutes, there was inconsistency of results for the different muscles. The injected samples of biceps femoris and loin portion of longissimus dorsi were similar to or less juicy than the control samples in contrast to improved or similar juiciness for injected samples of other muscles (Figure 16).

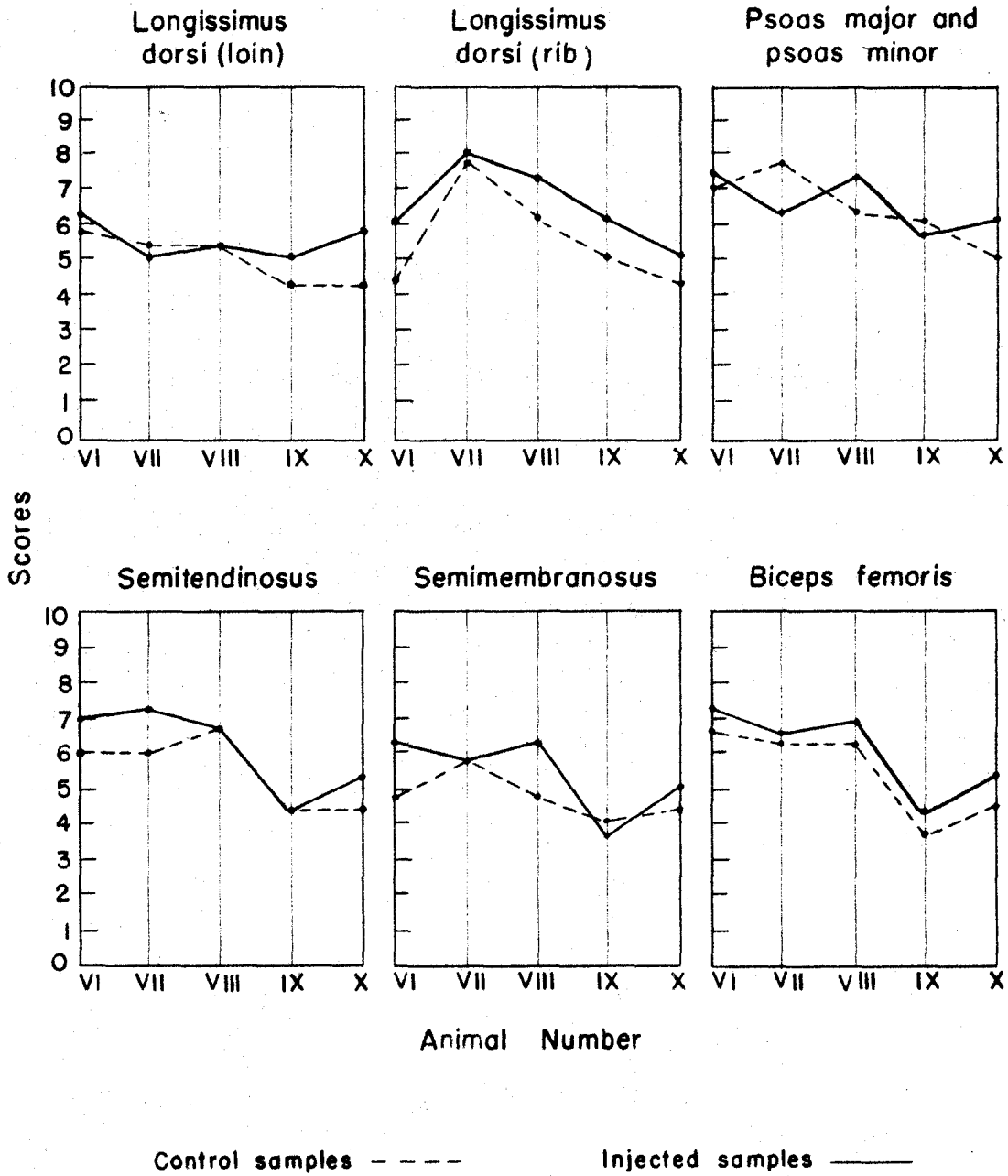


Figure 15. Scores for Juiciness of Canned Beef. Sodium chloride injection.

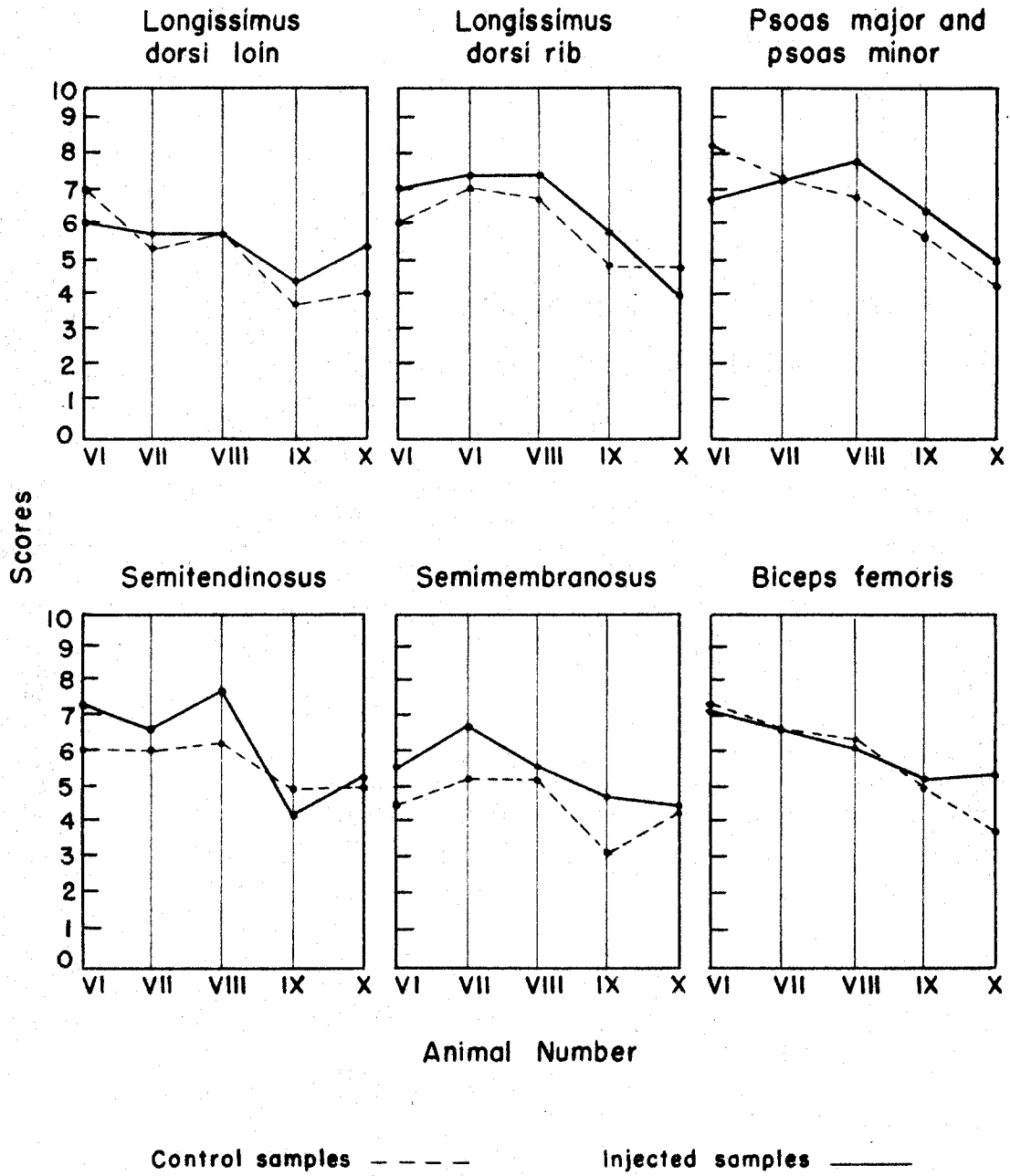


Figure 16. Scores for Juiciness of Canned Beef.
Sodium chloride and lactic acid injection.

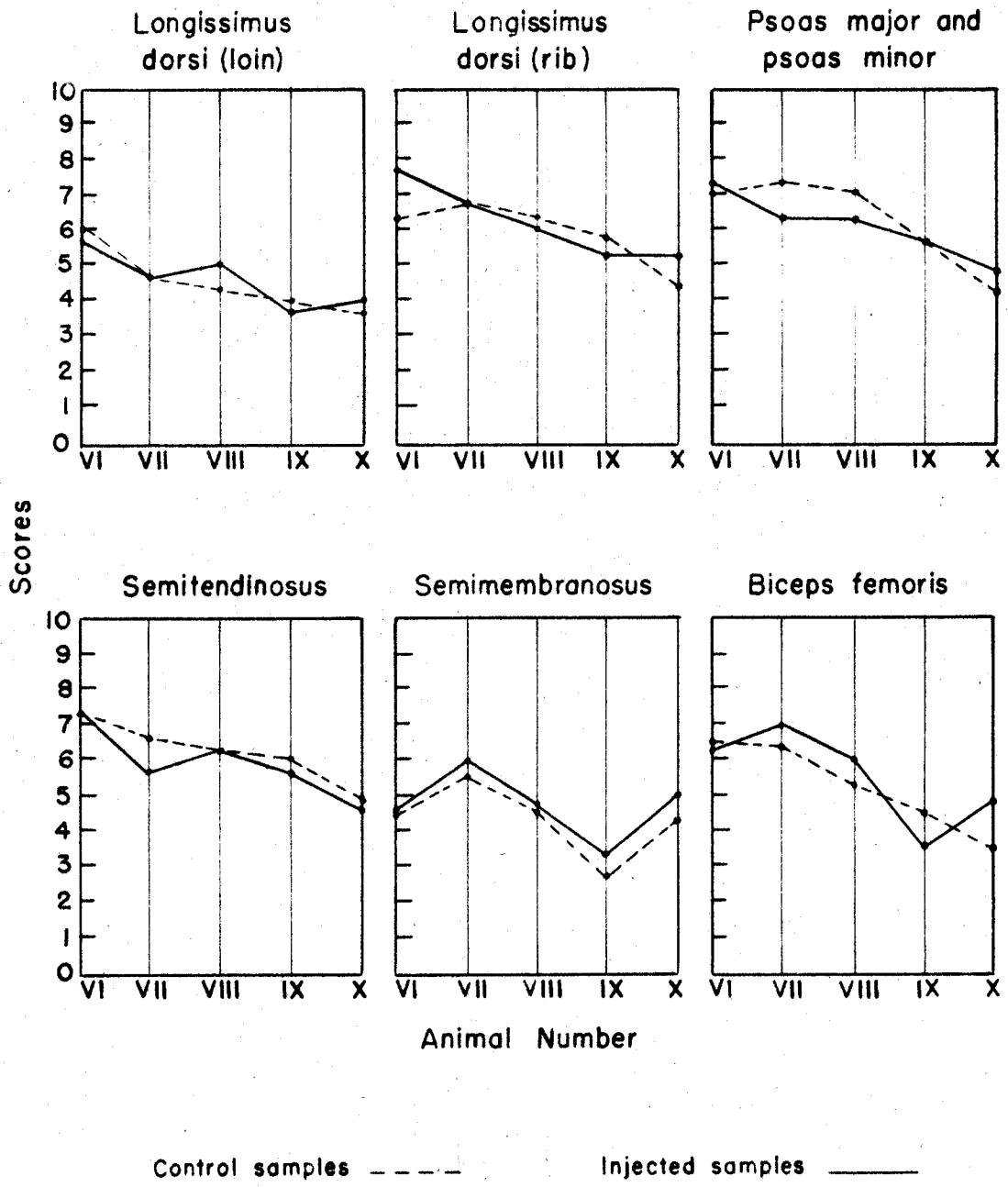


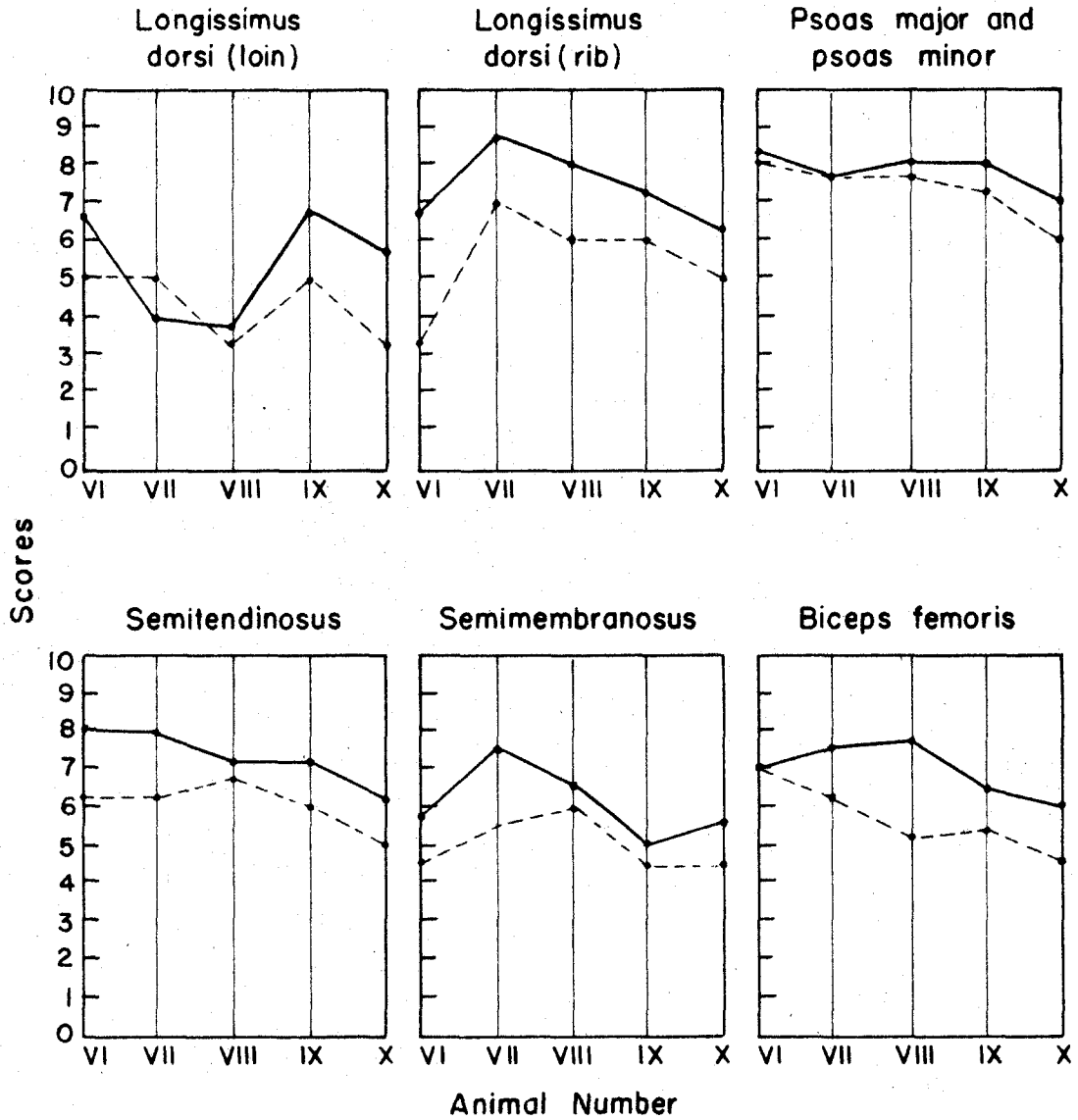
Figure 17. Scores for Juiciness of Canned Beef. Lactic acid injection.

The muscle differences of the samples of meat included in the experiments with lactic acid injection were not significant for the animals of group two, but for group one there was a highly significant difference among muscles. Meat from the loin portion of the longissimus dorsi and from the semimembranosus ranked lower in juiciness than the other muscles (Figure 17).

Improvement in juiciness of canned meat by injection with sodium chloride solution or with a combination of sodium chloride and lactic acid solution was less marked than the improvement in tenderness. Lactic acid injection had little effect on either tenderness or juiciness of the canned meat.

Texture

Texture scores showed considerable variation. The differences among muscles were shown to be significant at the .05 level for the meat from Animals IX and X and also for the samples included in the sodium chloride treatment from Animals VI, VII, and VIII. The texture differences among muscles were highly significant for the samples included in the other two injection treatments of the latter group. In general the psoas muscles were considered by the judges to have the most desirable texture; the longissimus dorsi loin and the semimembranosus, the least desirable texture of the muscles studied (Figures 18, 19, and 20). A highly



Control samples - - - - - Injected samples ———

Figure 18. Scores for Texture of Canned Beef. Sodium chloride injection.

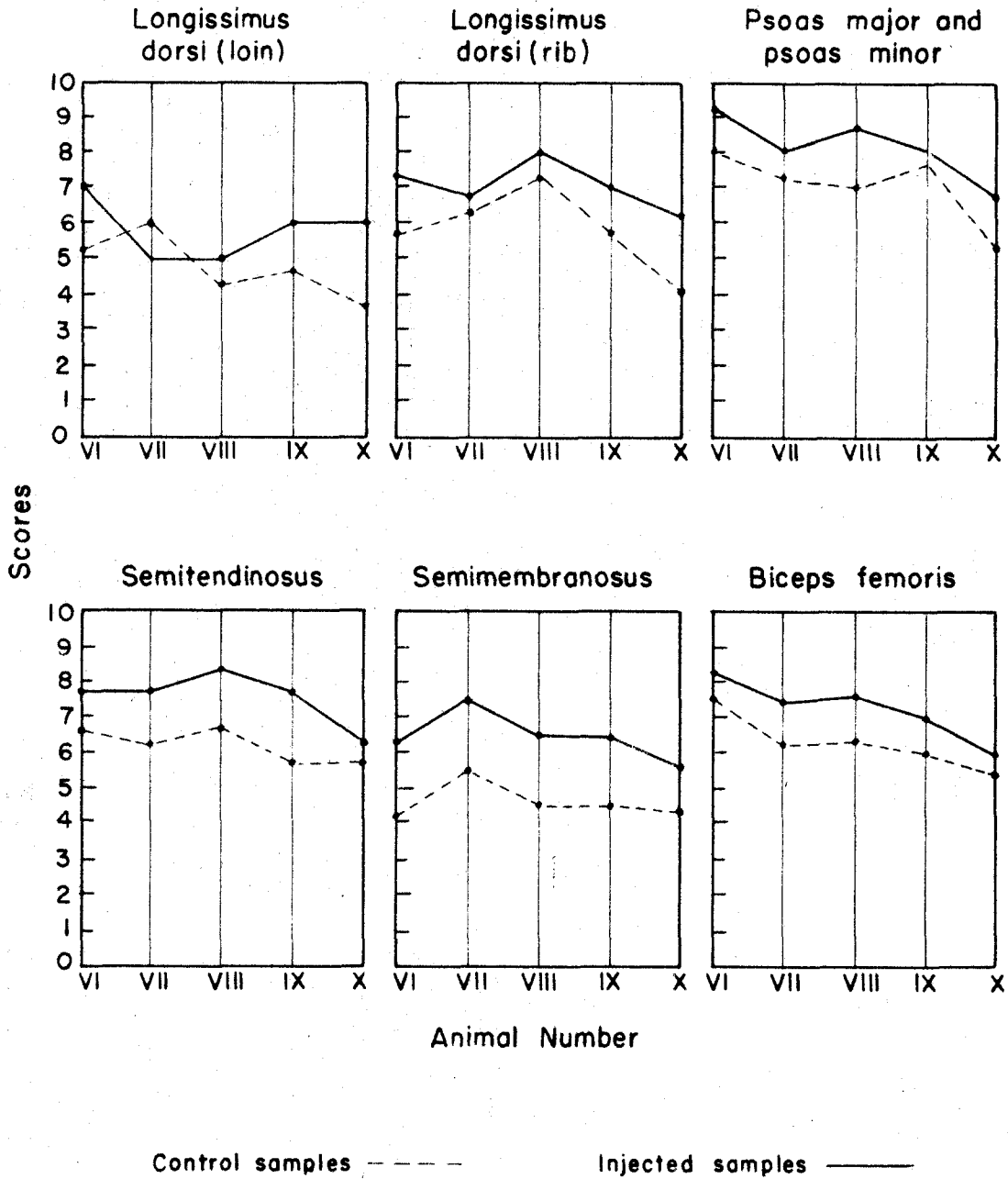


Figure 19. Scores for Texture of Canned Beef.
Sodium chloride and lactic acid injection.

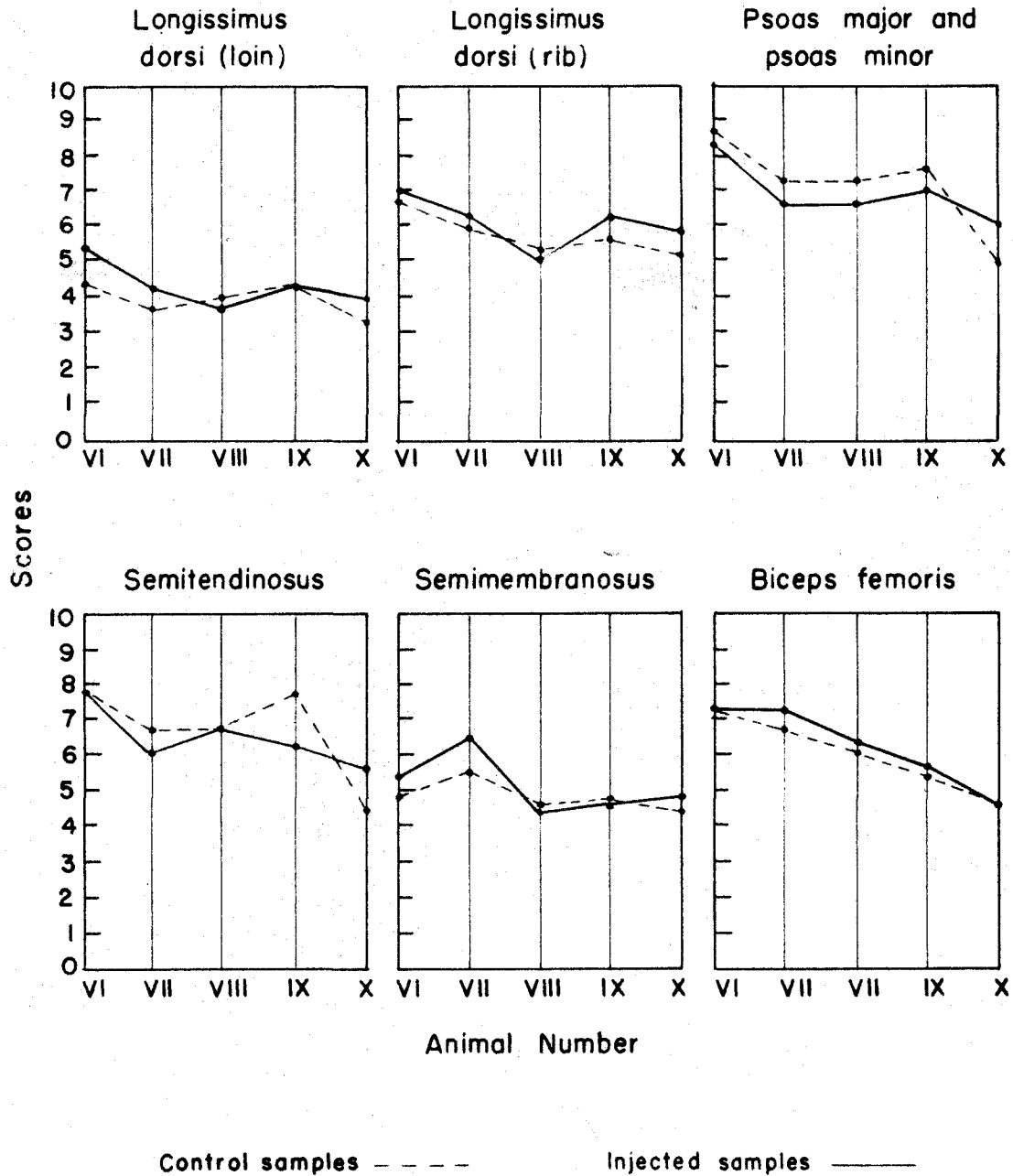


Figure 20. Scores for Texture of Canned Beef. Lactic acid injection.

significant improvement in the texture of the meat was shown by injection with sodium chloride solution or with sodium chloride and lactic acid solution. Average scores (Table 3) for texture of the meat from the five animals were 6.8 and 5.6, respectively, for injected and control samples with sodium chloride treatment; for the combined sodium chloride and lactic acid treatment, 7.1 and 5.8, respectively.

Lactic acid treatment had no significant effect on the texture scores of the canned beef. The average texture scores for the five animals were, respectively, 5.9 and 5.7 for the injected and uninjected meat (Table 3).

Thus, a marked improvement in texture was found in the canned meat as a result of injection of the raw beef with either sodium chloride solution or with the solution in which sodium chloride was combined with lactic acid. No significant change in texture was brought about by injection of the meat with lactic acid solution alone.

The uniformity of penetration of any of the solutions injected into the beef cuts was not determined. An effort was made to insert the needles so the distances between openings would be approximately equal. The pressure pump was operated as evenly as possible, but there was no automatic control to insure the uniform delivery of a specific amount of solution within a given area of the meat. Unequal distribution of the

solution within the meat may account for some of the variations observed in the results.

Histological Appearance of Beef Fibers

The rib portion of the longissimus dorsi muscle was used for histological study. Longitudinal microscopic sections were made of the fresh muscle aged 1 day, of the beef cuts aged 8 days, and of the canned beef. The effects of the three kinds of injection on the microscopic structure of the muscle fibers and the connective tissue were noted. Photomicrographs were taken of certain representative sections. Most of the pictures were of samples from Animal X. Sections were used which show the appearance of the meat from that muscle at the different aging periods, with different kinds of injection, and after canning. A few pictures of sections from other animals were included for comparison.

Sections for microscopic examination were small and represent only a limited area of the muscle. And, as has been previously mentioned, the injected solutions may not have been distributed uniformly throughout the muscle. An effort was made to obtain areas typical of the major portion of the section.

The samples of beef which had received certain of the treatments were especially difficult to section for mounting

on slides. Sodium chloride-treated samples were very troublesome. The uncooked samples were much more difficult to handle in sectioning and in mounting than the cooked ones. Hence, the appearance of the samples from the five animals was taken into consideration for the following descriptions. Disintegration appeared more slowly in the fibers of Animal VIII than in the muscle fibers of the other animals. This is reflected in the low tenderness scores of the canned beef from Animal VIII, particularly for control samples.

Harrison (18) has described the appearance of longitudinal sections of beef from various muscles after aging from one to 30 days. Each muscle had certain characteristics and the muscle fibers changed with aging. Very wavy, kinked fibers were characteristic of fresh muscle tissue, although waves did not always form in the fresh samples. Another characteristic of lack of aging was the longitudinal striations. The longitudinal striations predominated in the wavy fibers and often in the straight fibers of freshly cut beef. Harrison observed no longitudinal striae in the psoas muscle at any aging period. She did observe very heavy longitudinal striations in the longissimus dorsi muscle fibers. Sometimes both longitudinal and cross striae were distinct in some areas of a fiber, giving a checked appearance.

As the muscle aged the contractions were not so great

and the fibers were less wavy. The longitudinal striae gradually became less distinct in most areas of the fibers, the cross ones more distinct, although the longitudinal ones sometimes remained in the waved fibers. With further aging Harrison (18) and Paul (23) both observed disintegration or disappearance of the striations. This first occurred in narrow areas or cracks at wide intervals. With longer aging these disintegrated areas increased in size and numbers. The appearance of disintegration of the striae and its increase with aging were correlated with the increasing tenderness in a majority of the muscles.

The rate at which disintegration appeared varied in different muscles and was more rapid in the same muscle in some animals than in others.

Fresh muscle

The appearance of the muscle fibers of the rib portion of the longissimus dorsi muscle after 1 day of aging is shown in Figure 21. The wavy fibers and the contracture nodes are characteristic of fresh muscle. Longitudinal striae are visible in parts of the fibers, but the magnification for the photomicrograph was not great enough to show them clearly.

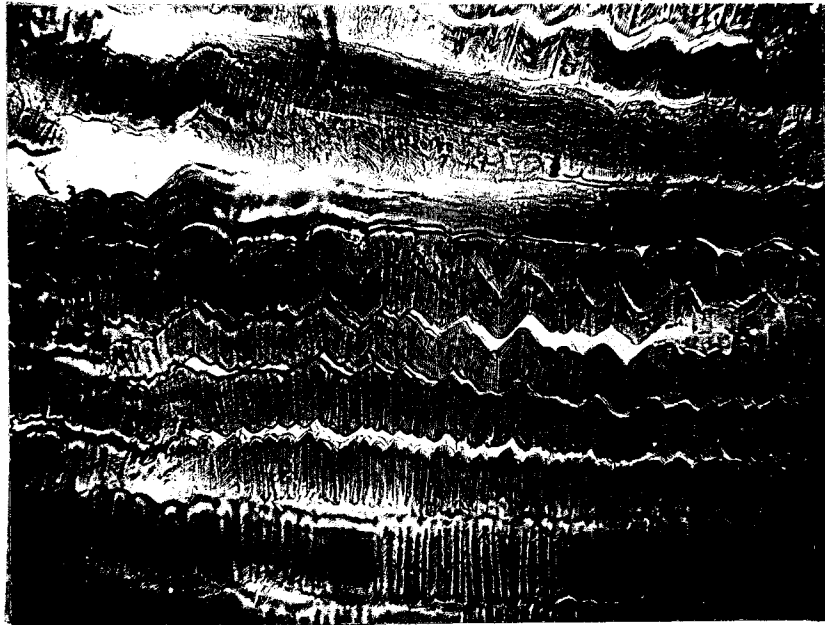


Figure 21. Fresh, Longissimus Dorsi, Rib Portion,
Middle Position. Aged One Day.
Animal X. (Magnification 150x)
This section is typical of fresh
muscle. The fibers are wavy and
contracture nodes are present.

Sodium chloride injection

Since the control samples aged 8 days had not been injected, they represent raw beef cuts which had aged a longer time than the samples from the fresh muscle. The upper picture in Figure 22 is a photomicrograph of a control sample from the sodium chloride-treated series of cuts. This section has the typical appearance of uncooked meat in which some resolution of rigor has taken place. The fibers are nearly straight, shallow waves are present, and the accordion-pleated effect of the fresh muscle has disappeared. The longitudinal striae are still distinct, particularly in waved portions of the fibers, and the sarcolemma is smooth except where a few cracks in the fibers have occurred.

The typical appearance of the raw beef injected with sodium chloride and aged 8 days is illustrated in the lower picture of Figure 22. The section was cut partially transverse of the fibers. In addition the connective tissue did not hold the muscle fibers together in the sodium chloride-treated meat. The segments of the fibers, however, show many cracks and broken places and much disintegration. The cross striae predominated over the longitudinal ones. These are characteristics which would be expected to contribute to tenderization in meat. It is interesting that for this particular sample of canned beef (number 261) the judges' score



Figure 22. Sodium Chloride Injection. Longissimus
Dorsi, Rib Portion. Animal X.
Upper: Aged, Control Sample No. 261.
(Magnification 85x)
Lower: Aged, Injected Sample No. 261.
(Magnification 150x)

for tenderness of the control sample was 7.0 and of the injected sample, 9.3 (Table 1, Appendix), indicating that the injected sample was rated more tender than the control.

A cut of meat from another animal (Animal IX) receiving the sodium chloride injection and aged for 8 days is illustrated in the upper picture of Figure 23. The fibers are fairly straight, and the disintegration fissures are not as numerous as in Animal X, but the parallel course of the fibers has been maintained better than in Animal X. It appears that the tenderizing effect of sodium chloride was not as extensive in this cut of meat as in that of Animal X. The tenderness score received by this cut of meat after canning (Table 1, Appendix) is higher than for its control (8.7 for the injected; 8.3 for the control), but not as high as the score of 9.3 for the sodium chloride-injected cut of the same muscle in Animal X.

The histological appearance of uninjected canned beef is shown in Figure 23, lower picture. The section is from the sodium chloride-treated series of Animal X, but was a control sample. The fibers have a dense, compact appearance and follow a fairly straight course. The whole section has a rather foggy appearance. An examination under high power shows the edges of the fibers to be uneven. There is much granular material, probably disintegration products of



Figure 23. Sodium Chloride Injection. Longissimus Dorsi, Rib Portion.
Upper: Aged, Injected Sample No. 239.
Animal IX. (Magnification 150x)
Lower: Canned, Control Sample No. 261.
Animal X. (Magnification 115x)

collagen since this granular material was found only in the canned beef sections, near the edges of the fibers and between adjacent ones, or within extensive areas of whole fibers.

Microscopic sections of the canned beef injected with sodium chloride solution have the same dense, compact appearance as the canned control samples. The fibers are generally straight and parallel, but have numerous disintegrated areas. Granular material is located within disintegrated areas of the fibers as well as along the edges of fibers and between adjacent ones. Little connective tissue is visible between the fibers.

Harrison (18) noted the opaque appearance of the sections from cooked beef roasts, and Paul (23) reported a decrease in the diameter of the muscle fibers with cooking, as observed from microscopic sections of raw and cooked roasts.

Sodium chloride and lactic acid injection

A control sample is shown in Figure 24, upper picture. The beef had been aged 8 days and was a cut from the same muscle of Animal X as the control sample of the sodium chloride series.

The fibers in the section are typical of those found in



Figure 24. Sodium Chloride and Lactic Acid Injection.
Longissimus Dorsi, Rib Portion. Animal X.
Upper: Aged, Control Sample No. 263.
(Magnification 150x)
Lower: Aged, Injected Sample No. 263.
(Magnification 150x)

other cuts aged 8 days. There are some waved fibers, but not so many as in the fresh sections. Shallow waves with fairly distinct longitudinal striations, some nodes, and a few disintegration cracks in the fibers can be seen. The cross striae predominate in the straight fibers.

A section of the matching injected sample aged 8 days is shown in Figure 24, lower picture. The disintegration of the fibers is more pronounced than in the control sample.

The histological sections of the control and injected samples of canned meat from the rib portion of the longissimus dorsi muscle are pictured in Figure 25. The disintegration cracks are more numerous and the areas more extensive in the injected than in the control sample. This is in keeping with the higher tenderness scores given by the judges to the sodium chloride-lactic acid-injected meat than to control samples.

Examples of canned beef injected with combined sodium chloride and lactic acid from two other animals are shown in Figure 26. The upper picture represents meat from the longissimus dorsi, rib portion, of Animal VII, which was processed 65 minutes; the lower picture, meat of Animal IX, which was processed 90 minutes. Both sections show straight fibers that are dense and compact and that have considerable disintegration. Granular material is present within and between the fibers. The meat processed the longer time has a

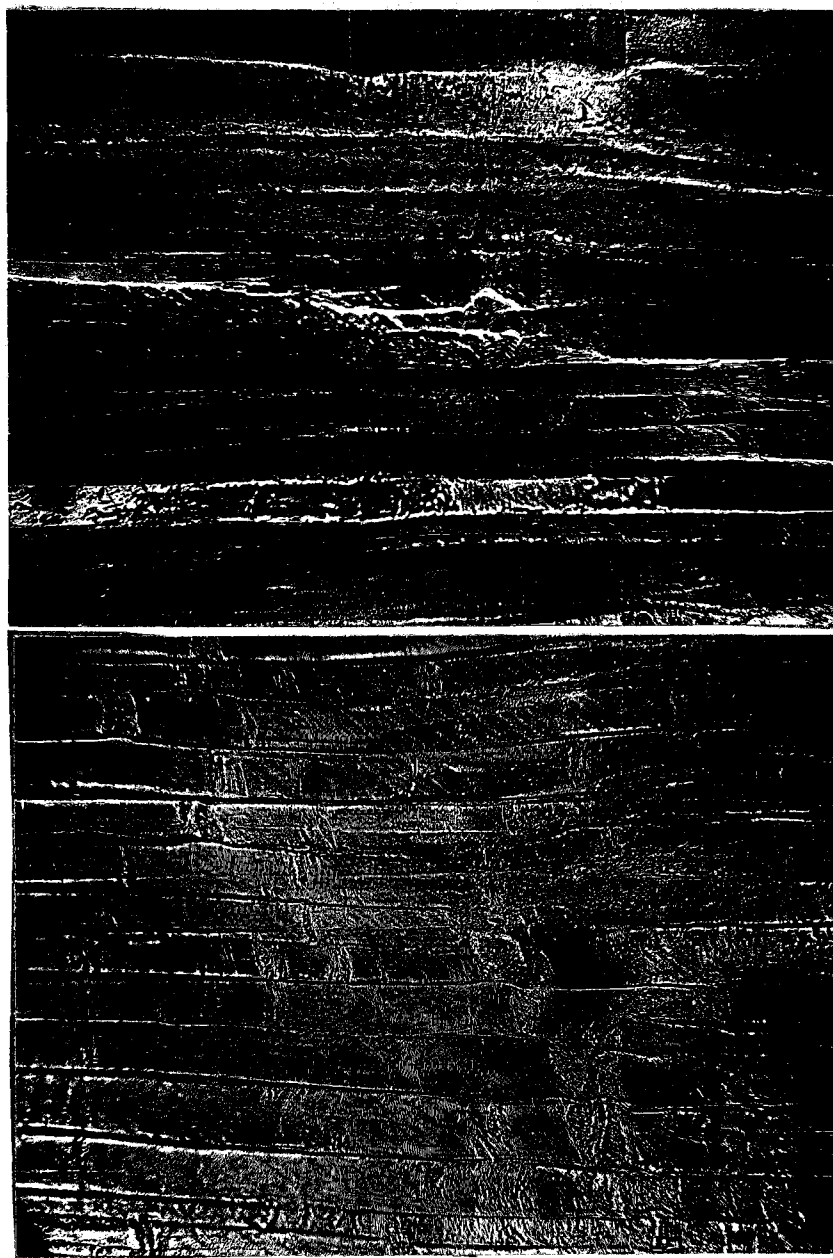


Figure 25. Sodium Chloride and Lactic Acid Injection.
Longissimus Dorsi, Rib Portion. Animal X.
Upper: Canned, Control Sample No. 263.
(Magnification 150x)
Lower: Canned, Injected Sample No. 263.
(Magnification 150x)

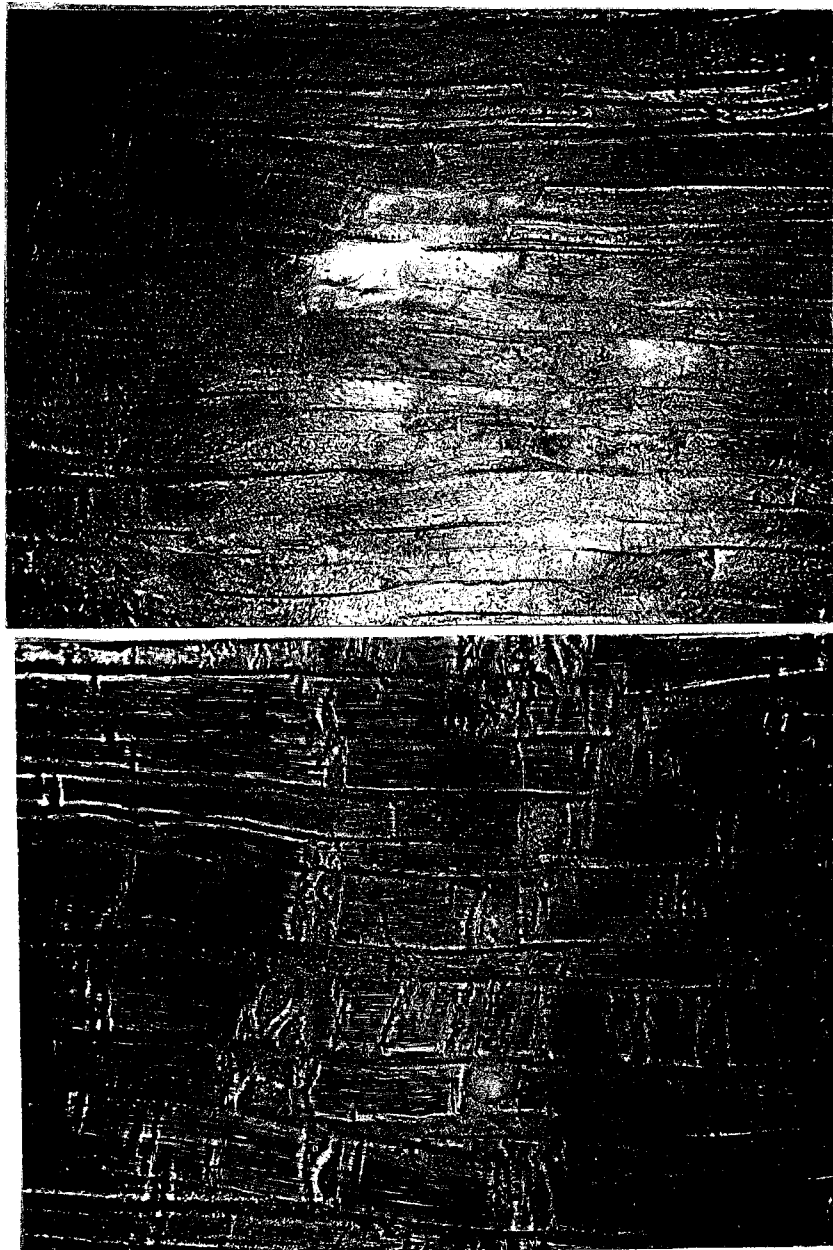


Figure 26. Sodium Chloride and Lactic Acid Injection.
Longissimus Dorsi, Rib Portion.
Upper: Canned, Injected Sample No. 191.
Animal VII. (Magnification 115x)
Lower: Canned, Injected Sample No. 238.
Animal IX. (Magnification 165x)

larger amount of disintegration (probably animal variation) in the fibers and looks more fragile than the meat processed the shorter time.

Lactic acid injection

The aged samples of the series treated with lactic acid were found to have histological features very nearly like those of the control samples. The disintegration is a little more pronounced and the fibers slightly less wavy in the cuts receiving the lactic acid injection than in the controls.

(See Figure 27.)

The canned samples of the lactic acid series are illustrated in Figure 28. The areas of disintegration are a little more extensive in the injected than in the uninjected cut of beef but, otherwise, the two cuts are similar in microscopic structure. This agrees with the uniformity of the scores given by the judges to the palatability factors of control and lactic acid-treated samples of canned beef.

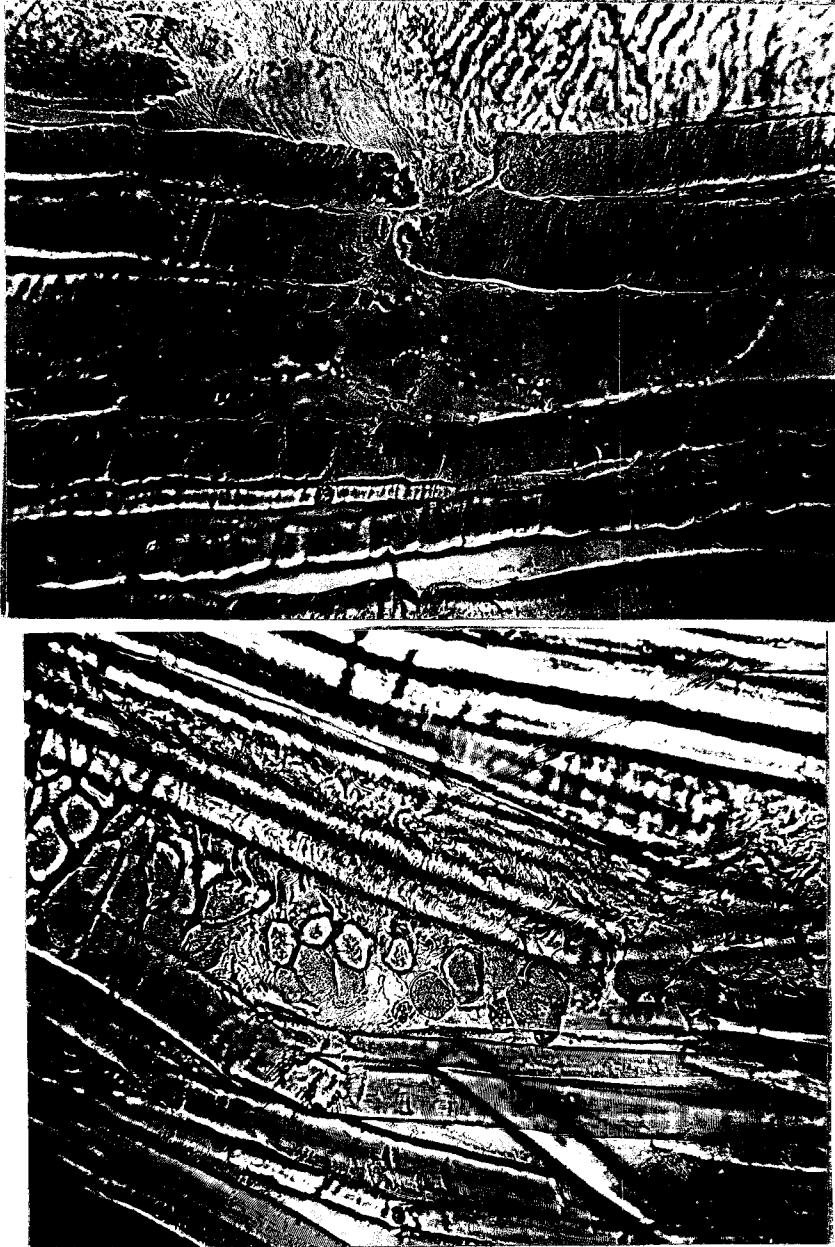


Figure 27. Lactic Acid Injection. Longissimus Dorsi,
Rib Portion. Animal X.
Upper: Aged, Control Sample No. 262.
(Magnification 150x)
Lower: Aged, Injected Sample No. 262.
(Magnification 150x)

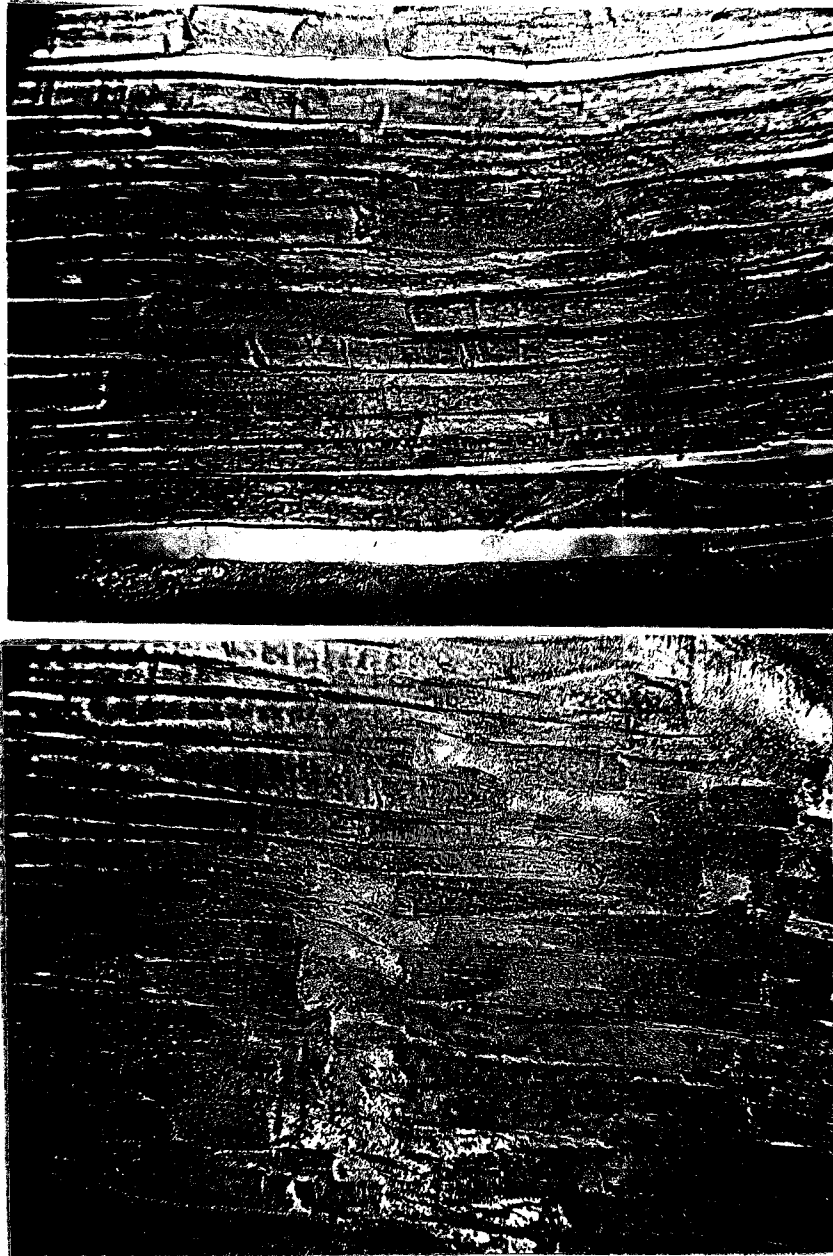


Figure 28. Lactic Acid Injection. Longissimus Dorsi,
Rib Portion. Animal X.

Upper: Canned, Control Sample No. 262.
(Magnification 150x)

Lower: Canned, Injected Sample No. 262.
(Magnification 150x)

Slicing Quality of Canned Beef

The slicing quality of the canned beef was evaluated in three ways: (1) by count of the number of slices obtained from each can, (2) by weight, and (3) by judges' scores. Slices were classified as firm or crumbly. If practically the whole section held together after cutting on the machine it was called a firm slice. If between three-fourths and the whole slice held together it was classified as crumbly. If less than three-fourths of the slice held together it was grouped with the unsliceable portion. Slices were rated by the scoring panel on the proportion of the slice holding together. The scores were based on a value of 10 for extremely good slicing quality and 1 for poor quality.

The averages of the numbers of slices obtained for firm, crumbly, and total slices are shown in Table 4. Variability in the sliceability of the canned beef occurred among muscles and among animals. Furthermore, cuts from the same muscle sometimes gave variable results.

None of the three types of injection improved the slicing quality appreciably. The average figures (Table 4) for the firm slices from samples of all animals in the sodium chloride series were 39.8 and 39.3 per cent for the control and injected cuts, respectively. The results are presented

Table 4. Averages of Numbers of Slices Obtained.

Animal no.	No. of muscles	No. slices obtained		Possible no. slices	Slices obtained		
		Firm	Crumbly		Firm %	Crumbly %	Total %
SODIUM CHLORIDE INJECTION							
Control samples							
VI	6	5.0	3.9	10.5	48.1	37.1	85.2
VII	6	3.9	3.4	11.2	34.0	31.4	65.4
VIII	6	4.8	3.7	11.2	44.1	32.2	76.3
Av. (3 animals)		4.6	3.7	11.0	42.1	33.6	75.6
IX	6	3.3	3.1	11.1	32.7	26.8	59.5
X	6	4.7	4.5	11.4	40.1	39.6	79.8
Av. (2 animals)		4.0	3.8	11.2	36.4	33.2	69.6
Av. (5 animals)		4.3	3.7	11.1	39.8	33.4	73.2
Injected samples							
VI	6	6.0	2.8	10.4	58.2	26.0	84.2
VII	6	4.0	3.3	11.3	34.4	29.4	63.9
VIII	6	2.8	3.6	11.1	27.4	32.7	60.1
Av. (3 animals)		4.3	3.2	10.9	40.0	29.4	69.4
IX	6	2.3	1.6	10.6	24.0	14.8	38.8
X	6	6.3	3.4	11.6	52.6	31.1	83.7
Av. (2 animals)		4.3	2.5	11.1	38.3	23.0	61.2
Av. (5 animals)		4.3	2.9	11.0	39.3	26.8	66.1

(continued)

Table 4 (continued)

Animal no.	No. of muscles	No. slices obtained		Possible no. slices	Slices obtained		
		Firm	Crumbly		Firm %	Crumbly %	Total %
SODIUM CHLORIDE AND LACTIC ACID INJECTION							
Control samples							
VI	6	3.7	4.7	10.6	33.2	44.9	78.1
VII	6	5.1	3.4	11.3	45.2	29.7	74.9
VIII	6	3.2	4.9	11.3	31.0	42.9	73.9
Av. (3 animals)		4.0	4.3	11.1	36.5	39.2	75.6
IX	6	2.0	2.1	10.2	20.1	20.3	40.4
X	6	5.5	1.9	11.8	47.3	16.2	63.4
Av. (2 animals)		3.8	2.0	11.0	33.7	18.2	51.9
Av. (5 animals)		3.9	3.4	11.0	35.4	30.8	66.1
Injected samples							
VI	6	4.3	4.3	10.6	39.8	42.2	82.0
VII	6	3.4	3.8	10.8	31.7	35.2	66.8
VIII	6	3.8	5.2	10.9	35.0	46.6	81.6
Av. (3 animals)		3.8	4.4	10.8	35.5	41.3	76.8
IX	6	1.2	2.8	10.4	13.4	26.6	40.0
X	6	5.1	3.5	11.2	44.6	31.2	75.8
Av. (2 animals)		3.2	3.2	10.8	29.0	28.9	57.9
Av. (5 animals)		3.6	3.9	10.8	32.9	36.4	69.2

(continued)

Table 4 (continued)

Animal no.	No. of muscles	No. slices obtained		Possible no. slices	Slices obtained		
		Firm	Crumbly		Firm %	Crumbly %	Total %
LACTIC ACID INJECTION							
Control samples							
VI	6	5.2	3.7	10.8	47.3	34.3	81.6
VII	6	4.2	2.8	11.4	37.5	26.2	63.7
VIII	6	3.6	2.7	11.3	33.4	23.5	56.9
Av. (3 animals)		4.3	3.1	11.2	39.4	28.0	67.4
IX	6	2.2	2.4	10.8	20.9	22.2	43.1
X	6	5.8	1.9	11.2	51.4	17.2	68.6
Av. (2 animals)		4.0	2.2	11.0	36.2	19.7	55.8
Av. (5 animals)		4.2	2.7	11.1	38.1	24.7	62.8
Injected samples							
VI	6	3.9	4.0	10.9	34.4	37.4	71.8
VII	6	4.8	2.2	10.9	42.3	19.9	62.2
VIII	6	6.5	1.9	11.2	58.6	16.6	75.2
Av. (3 animals)		5.1	2.7	11.0	45.1	24.6	69.7
IX	6	2.2	2.2	10.6	20.8	21.5	42.2
X	6	6.2	2.9	11.0	56.6	26.0	82.6
Av. (2 animals)		4.2	2.6	10.8	38.7	23.8	62.4
Av. (5 animals)		4.7	2.6	10.9	42.5	24.3	66.8

graphically for each animal in Figure 29.

Sodium chloride and lactic acid together were no more effective than sodium chloride alone in improving the slicing quality of the beef. The figures for the average percentages of firm slices from the five animals were 35.4 and 32.9, respectively for control and injected samples in the sodium chloride-lactic acid series. The results for individual animals are shown in graphs, Figure 30.

The results with lactic acid injection were only slightly more favorable than for the other two kinds of injection. The averages for the firm slices from the five animals were 42.5 per cent for the injected meat and 38.1 per cent for the uninjected samples. However, the variability of results for the different animals (Figure 31) was so large that no real improvement in slicing quality was secured.

The averages of the judges' scores for slicing quality of the canned beef are given in Table 5. The differences in scores between the control and injected samples were not large enough to be significant with the sodium chloride series. The differences between muscles as to slicing quality were highly significant. The semitendinosus was rated the highest of the muscles in sliceability; the semimembranosus, the lowest. Within the group composed of Animals VI, VII, and VIII, the differences among animals were not significant,

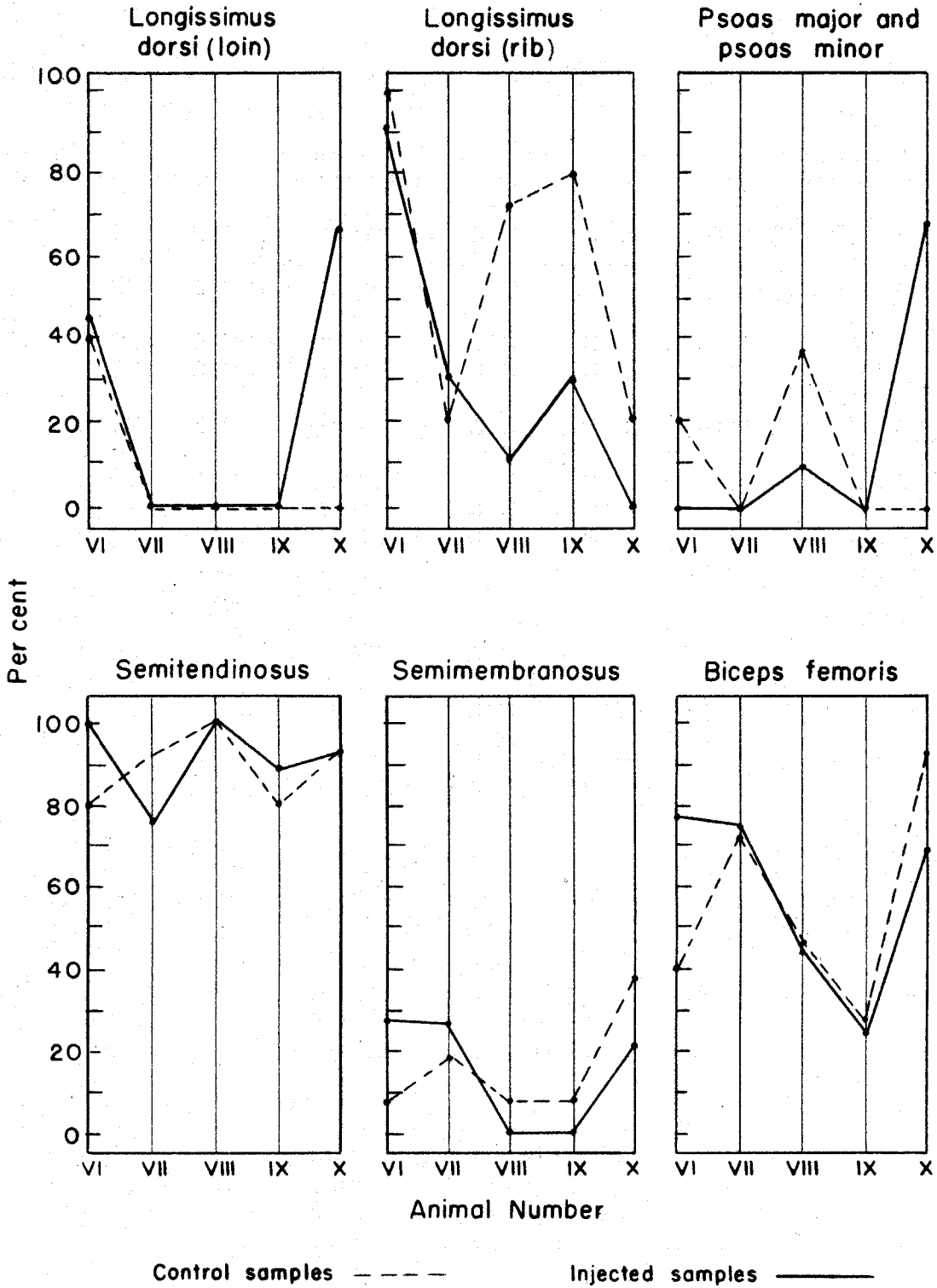


Figure 29. Number of Firm Slices.
Sodium chloride injection.

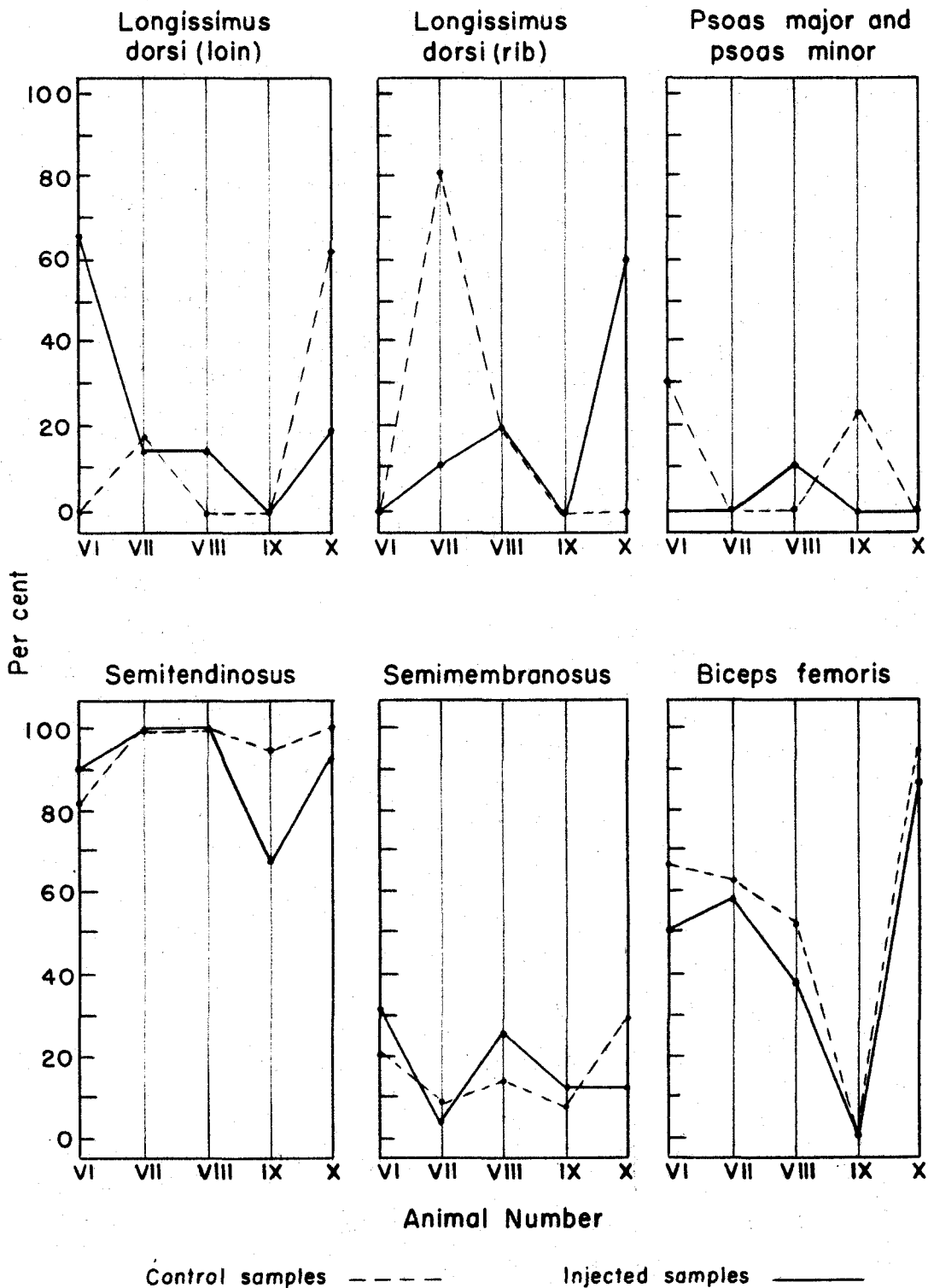


Figure 30. Number of Firm Slices.
Sodium chloride and lactic acid
injection.

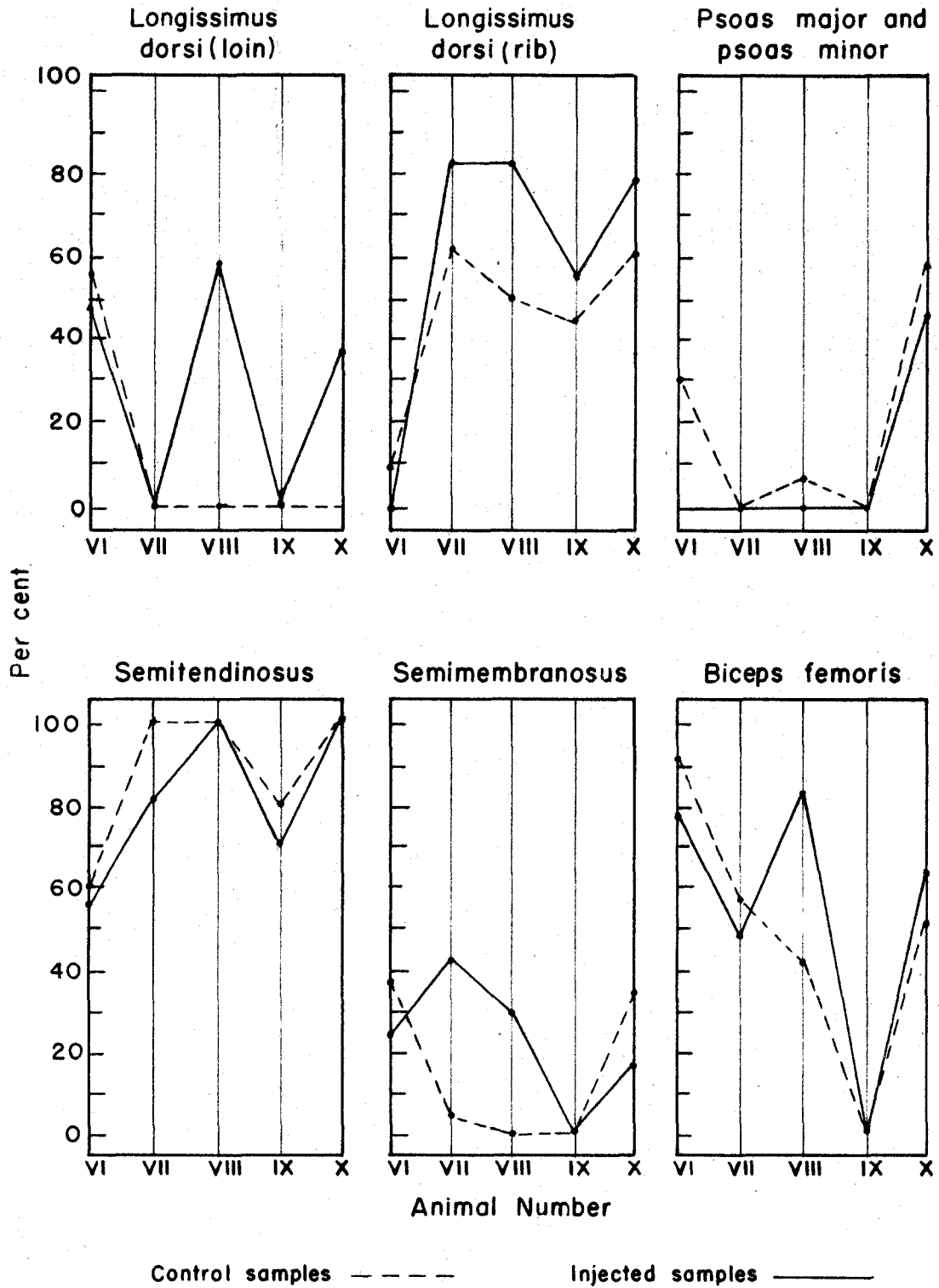


Figure 31. Number of Firm Slices.
Lactic acid injection.

Table 5. Averages of Judges' Ratings of Sliceability and Averages of Sliceability on Basis of Weight.

Animal no.	No. of muscles	Slice-ability score	Sliceability on basis of wt.			
			Canned meat gm.	Unslice-able meat gm.	Sliceable meat gm.	(by difference) %

SODIUM CHLORIDE INJECTION

Control samples

VI	6	7.6	383	46	338	87.8
VII	6	7.6	392	111	281	71.4
VIII	6	7.9	370	73	296	80.2
Av. (3 animals)		7.7	382	77	305	79.8
IX	6	7.0	377	132	245	65.2
X	6	7.2	357	69	288	81.0
Av. (2 animals)		7.1	367	100	266	73.1
Av. (5 animals)		7.5	376	86	290	77.1

Injected samples

VI	6	7.6	370	45	325	87.9
VII	6	6.8	384	125	259	68.9
VIII	6	6.7	356	126	231	64.6
Av. (3 animals)		7.0	370	99	272	73.8
IX	6	5.0	364	198	166	45.7
X	6	8.3	364	47	317	87.1
Av. (2 animals)		6.6	364	122	242	66.4
Av. (5 animals)		6.9	368	108	260	70.8

(continued)

Table 5 (continued)

Animal no.	No. of muscles	Slice-ability score	Sliceability on basis of wt.			
			Canned meat gm.	Unslice-able meat gm.	Sliceable meat gm.	(by difference) %
SODIUM CHLORIDE AND LACTIC ACID INJECTION						
Control samples						
VI	6	7.8	382	69	312	82.0
VII	6	7.5	387	82	305	79.3
VIII	6	7.2	372	80	292	78.6
Av. (3 animals)		7.5	380	77	303	80.0
IX	6	5.4	369	194	175	47.5
X	6	7.5	367	124	243	67.0
Av. (2 animals)		6.4	368	159	209	57.2
Av. (5 animals)		7.1	375	110	265	70.9
Injected samples						
VI	6	8.4	379	56	323	85.3
VII	6	6.2	384	113	270	71.4
VIII	6	7.6	356	56	300	84.3
Av. (3 animals)		7.4	373	75	298	80.3
IX	6	6.1	361	195	166	46.0
X	6	7.9	366	68	299	81.5
Av. (2 animals)		7.0	364	132	232	63.8
Av. (5 animals)		7.2	369	98	272	73.7

(continued)

Table 5 (continued)

Animal no.	No. of muscles	Slice-ability score	Sliceability on basis of wt.			
			Canned meat gm.	Unslice-able meat gm.	Sliceable meat gm.	(by difference) %
LACTIC ACID INJECTION						
Control samples						
VI	6	8.2	379	57	322	84.5
VII	6	6.2	396	124	272	68.6
VIII	6	6.6	371	146	225	60.7
Av. (3 animals)		7.0	382	109	273	71.3
IX	6	5.8	374	191	183	49.0
X	6	7.3	365	101	264	73.0
Av. (2 animals)		6.6	370	146	224	61.0
Av. (5 animals)		6.8	377	124	253	67.2
Injected samples						
VI	6	7.6	371	84	288	77.8
VII	6	7.0	371	111	260	69.8
VIII	6	7.3	357	69	289	81.2
Av. (3 animals)		7.3	366	88	279	76.3
IX	6	5.3	357	182	175	49.2
X	6	7.6	345	56	290	83.9
Av. (2 animals)		6.4	351	119	232	66.6
Av. (5 animals)		7.0	360	100	260	72.4

but the differences between Animals IX and X were highly significant.

The scores for slicing quality of the injected samples in the sodium chloride-lactic acid series were not significantly different from the control samples. Muscle differences were significant at the .01 level for Animals VI, VII, and VIII, but the differences were not quite large enough to be significant for Animals IX and X. The meat of Animal X sliced significantly better than that of Animal IX.

Lactic acid injection made no significant difference in judges' scores for slicing quality. Muscle differences were significant at the .05 level. The judges rated the meat of Animal X significantly higher in slicing quality than the meat of Animal IX. A possible explanation of the higher rating of Animal X in sliceability than for Animal IX is that the connective tissue of the aged dairy cow (Animal X) was firm enough, even after processing for 90 minutes, to hold the muscle fibers together.

The percentage values for sliceability calculated by deduction of the weight of the unsliceable meat from the weight of the meat in the can (See Table 5) are in good agreement with the values obtained by count of slices and by judges' ratings of sliceability.

pH Values

The average pH values for the raw meat are approximately 5.45, regardless of aging 1 or 8 days or the kind of injection (Table 6). The range among the average values for the five animals is 5.61 to 5.37. Canned beef had higher pH values than the raw meat and the meat of Animal X, either cooked or uncooked, was higher in pH value than the meat of the other animals. Animal X was the aged dairy cow and the meat of this animal was a dark red color. The raw meat was sticky and rubbery and had a rather strong odor. Hall (17) reported a relationship between dark color in beef and high pH values. Bate-Smith (7) enumerated qualities found in beef having high pH values, among which were dark color, slimy or sticky feel, and flabbiness. The kind of injection had little apparent effect on the pH values of the raw or canned beef.

Weight Changes

The animals used in this series of experiments were fairly uniform in weight. The live weights of the four steers were 816, 800, 830, and 930 pounds, respectively; the weight of the dairy cow was 965 pounds. The warm dressed weights of the half carcasses were, for left and right halves of each animal, respectively: 231, 232; 222, 216; 230, 233; 280, 275; and 195, 190 pounds. The cow had the highest

Table 6. Averages of pH Values.

Animal no.	No. of muscles	Beef muscles (uninjected) aged one day pH	Beef cuts aged eight days pH	Canned beef pH
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SODIUM CHLORIDE INJECTION

Left muscle		Control samples		
VI	6	5.38	5.44	5.80
VII	6	5.42	5.44	5.68
VIII	6	5.40	5.38	5.75
Av. (3 animals)		5.40	5.42	5.74
IX	6	5.38	5.37	5.64
X	6	5.61	5.55	5.93
Av. (2 animals)		5.50	5.46	5.78
Av. (5 animals)		5.44	5.44	5.76
Right muscle		Injected samples		
VI	6	5.39	5.43	5.77
VII	6	5.44	5.49	5.65
VIII	6	5.42	5.41	5.73
Av. (3 animals)		5.42	5.44	5.72
IX	6	5.38	5.42	5.62
X	6	5.60	5.54	5.86
Av. (2 animals)		5.49	5.48	5.74
Av. (5 animals)		5.45	5.46	5.73

(continued)

Table 6 (continued)

Animal no.	No. of muscles	Beef muscles (uninjected) aged one day pH	Beef cuts aged eight days pH	Canned beef pH
SODIUM CHLORIDE AND LACTIC ACID INJECTION				
Left muscle			Control samples	
VI	6	(same as for sodium chloride injection)	5.44	5.80
VII	6		5.42	5.64
VIII	6		5.39	5.73
Av. (3 animals)			5.42	5.72
IX	6		5.39	5.65
X	6		5.53	5.93
Av. (2 animals)			5.46	5.79
Av. (5 animals)			5.43	5.75
Right muscle			Injected samples	
VI	6	(same as for sodium chloride injection)	5.47	5.78
VII	6		5.48	5.63
VIII	6		5.41	5.70
Av. (3 animals)			5.45	5.70
IX	6		5.42	5.64
X	6		5.60	5.87
Av. (2 animals)			5.51	5.76
Av. (5 animals)			5.48	5.72

(continued)

Table 6 (continued)

Animal no.	No. of muscles	Beef muscles (uninjected) aged one day	Beef cuts aged eight days	Canned beef
		pH	pH	pH
LACTIC ACID INJECTION				
Left muscle			Control samples	
VI	6	(same as for sodium chloride injection)	5.42	5.80
VII	6		5.48	5.67
VIII	6		5.40	5.73
Av. (3 animals)			5.43	5.73
IX	6		5.38	5.65
X	6		5.54	5.92
Av. (2 animals)			5.46	5.78
Av. (5 animals)			5.44	5.75
Right muscle			Injected samples	
VI	6	(same as for sodium chloride injection)	5.47	5.82
VII	6		5.48	5.68
VIII	6		5.41	5.73
Av. (3 animals)			5.45	5.74
IX	6		5.38	5.65
X	6		5.56	5.94
Av. (2 animals)			5.47	5.80
Av. (5 animals)			5.46	5.76

live weight but the lowest dressed weights for each half of the carcass of any of the animals. The weights of the muscles (untrimmed) after separation from the carcass are shown in Table 7.

The cuts of beef from each animal were trimmed to approximately a certain weight prior to aging. The averages of the weights of the beef cuts, of the beef plus injecting solution, and of the cuts after aging are shown in Table 8. The control samples lost a small percentage of their initial weight during the aging period; injected samples had higher average weights after aging than their initial weights, i. e., some of the injected solution remained in the meat at the end of 8 days of aging. Weight differences associated with a particular kind of injection were very small.

The average figures for the proportions of meat and of liquid in the can after processing were fairly uniform for the control and injected samples with the three kinds of injection (Table 9). Approximately two-thirds of the weight of meat put into the can was in the form of meat after processing and the other one-third was liquid. The differences in the percentage of liquid to meat for control and injected samples were too small to be of importance. The average values for the percentage of liquid in the cans of meat processed 90 minutes were no higher than for the meat processed 65 minutes.

Table 7. Weight of Muscles (Untrimmed) after Separation from Carcass.

Muscle	Wt. of muscle (untrimmed)	
	Left lb.	Right lb.
<u>Animal VI</u>		
Longissimus dorsi, loin portion	5.9	6.0
Longissimus dorsi, rib portion	7.5	7.2
Psoas major and psoas minor	4.1	3.7
Semitendinosus	3.5	3.4
Semimembranosus	8.5	8.6
Biceps femoris	8.5	9.1
<u>Animal VII</u>		
Longissimus dorsi, loin portion	6.2	6.2
Longissimus dorsi, rib portion	7.8	8.0
Psoas major and psoas minor	3.7	4.3
Semitendinosus	3.1	3.4
Semimembranosus	9.1	8.7
Biceps femoris	9.3	8.9
<u>Animal VIII</u>		
Longissimus dorsi, loin portion	5.9	6.5
Longissimus dorsi, rib portion	10.1	10.5
Psoas major and psoas minor	4.0	4.1
Semitendinosus	3.5	3.6
Semimembranosus	9.9	9.6
Biceps femoris	10.1	9.7
<u>Animal IX</u>		
Longissimus dorsi, loin portion	7.0	6.9
Longissimus dorsi, rib portion	11.6	11.0
Psoas major and psoas minor	5.6	4.9
Semitendinosus	4.7	4.7
Semimembranosus	11.1	11.9
Biceps femoris	12.6	11.8
<u>Animal X</u>		
Longissimus dorsi, loin portion	4.0	3.8
Longissimus dorsi, rib portion	7.0	5.9
Psoas major and psoas minor	3.8	3.2
Semitendinosus	2.8	2.9
Semimembranosus	11.5	11.9
Biceps femoris	8.4	8.1

Table 8. Averages of Initial Weights of Beef Cuts, of the Cuts Plus Injecting Solution, and of the Cuts after Aging.

Animal no.	No. of muscles	Initial wt. gm.	Beef plus injecting solution gm.	Wt. beef after aging gm.	%
SODIUM CHLORIDE INJECTION					
Control samples					
VI	6	606		600	99.0
VII	6	602		593	98.5
VIII	6	606		596	98.4
Av. (3 animals)		605		596	98.6
IX	6	605		590	97.6
X	6	605		599	99.0
Av. (2 animals)		605		594	98.3
Av. (5 animals)		605		596	98.5
Injected samples					
VI	6	605	665	638	105.5
VII	6	604	664	644	106.5
VIII	6	606	666	631	104.1
Av. (3 animals)		605	665	638	105.4
IX	6	605	665	638	105.3
X	6	606	666	642	106.0
Av. (2 animals)		606	666	640	105.6
Av. (5 animals)		605	665	639	105.5

(continued)

Table 8 (continued)

Animal no.	No. of muscles	Initial wt. gm.	Beef plus injecting solution gm.	Wt. beef after aging	
				gm.	%
SODIUM CHLORIDE AND LACTIC ACID INJECTION					
Control samples					
VI	6	606		597	98.6
VII	6	603		594	98.5
VIII	6	606		597	98.5
Av. (3 animals)		605		596	98.5
IX	6	604		591	97.9
X	6	605		597	98.6
Av. (2 animals)		604		594	98.2
Av. (5 animals)		605		595	98.4
Injected samples					
VI	6	606	666	636	105.1
VII	6	604	664	636	105.3
VIII	6	606	666	636	104.9
Av. (3 animals)		605	665	636	105.1
IX	6	604	664	628	103.8
X	6	605	665	640	105.8
Av. (2 animals)		604	664	634	104.8
Av. (5 animals)		605	665	635	105.0

(continued)

Table 8 (continued)

Animal no.	No. of muscles	Initial wt.	Beef plus injecting solution	Wt. beef after aging	
		gm.	gm.	gm.	%
LACTIC ACID INJECTION					
Control samples					
VI	6	605		600	99.3
VII	6	604		594	98.4
VIII	6	606		597	98.4
Av. (3 animals)		605		597	98.7
IX	6	606		596	98.5
X	6	605		598	98.9
Av. (2 animals)		606		597	98.7
Av. (5 animals)		605		597	98.7
Injected samples					
VI	6	604	664	624	103.3
VII	6	603	663	622	103.1
VIII	6	606	666	615	101.4
Av. (3 animals)		604	664	620	102.6
IX	6	604	664	615	101.8
X	6	606	666	634	104.5
Av. (2 animals)		605	665	624	103.2
Av. (5 animals)		605	665	622	102.8

Table 9. Averages of Weights of Beef before Processing and Weights of Meat and of Liquid after Processing.

Animal no.	No. of muscles	Wt. before processing gm.	Wt. after processing			
			Meat		Liquid	
			gm.	%	gm.	%
SODIUM CHLORIDE INJECTION						
Control samples						
VI	6	569	383	67.4	185	32.5
VII	6	570	392	68.8	173	30.3
VIII	6	569	370	64.9	195	34.3
Av. (3 animals)		569	382	67.0	184	32.4
IX	6	569	377	66.3	184	32.4
X	6	569	357	62.8	188	33.0
Av. (2 animals)		569	367	64.6	186	32.7
Av. (5 animals)		569	376	66.0	185	32.5
Injected samples						
VI	6	570	370	64.9	197	34.5
VII	6	570	384	67.4	180	31.6
VIII	6	570	356	62.5	206	36.1
Av. (3 animals)		570	370	64.9	194	34.1
IX	6	568	364	63.9	180	31.6
X	6	569	364	64.0	185	32.4
Av. (2 animals)		568	364	64.0	182	32.0
Av. (5 animals)		569	368	64.5	190	33.2

(continued)

Table 9 (continued)

Animal no.	No. of muscles	Wt. before processing gm.	Wt. after processing			
			Meat		Liquid	
			gm.	%	gm.	%
SODIUM CHLORIDE AND LACTIC ACID INJECTION						
Control samples						
VI	6	569	382	67.0	183	32.2
VII	6	569	387	68.0	179	31.6
VIII	6	570	372	65.3	192	33.7
Av. (3 animals)		569	380	66.8	185	32.5
IX	6	569	369	64.8	187	32.8
X	6	569	367	64.5	186	32.8
Av. (2 animals)		569	368	64.6	186	32.8
Av. (5 animals)		569	375	65.9	185	32.6
Injected samples						
VI	6	570	379	66.5	178	31.3
VII	6	569	384	67.3	182	32.0
VIII	6	570	356	62.6	207	36.3
Av. (3 animals)		570	373	65.5	189	33.2
IX	6	569	361	63.5	191	33.6
X	6	569	366	64.4	178	31.3
Av. (2 animals)		569	364	64.0	184	32.4
Av. (5 animals)		569	369	64.9	187	32.9

(continued)

Table 9 (continued)

Animal no.	No. of muscles	Wt. before processing gm.	Wt. after processing			
			Meat		Liquid	
			gm.	%	gm.	%
LACTIC ACID INJECTION						
Control samples						
VI	6	569	379	66.6	187	32.8
VII	6	569	396	69.6	170	29.8
VIII	6	569	371	65.2	194	34.0
Av. (3 animals)		569	382	67.1	184	32.2
IX	6	569	374	65.8	175	30.7
X	6	569	365	64.1	208	36.5
Av. (2 animals)		569	370	65.0	192	33.6
Av. (5 animals)		569	377	66.3	187	32.8
Injected samples						
VI	6	569	371	65.2	191	33.6
VII	6	569	371	65.2	192	33.7
VIII	6	568	357	62.9	205	36.1
Av. (3 animals)		569	366	64.4	196	34.5
IX	6	568	357	62.8	192	33.8
X	6	569	345	60.7	176	31.0
Av. (2 animals)		568	351	61.8	184	32.4
Av. (5 animals)		569	360	63.4	191	33.6

General Characteristics of Beef Samples

Some characteristics noted in the raw meat have already been mentioned, i.e., the color, rubber-like quality, and stickiness of the meat of Animal X. Muscles of one animal differed in color; for example, the semitendinosus was noticeably lighter in color than the other muscles; one portion of the biceps femoris was a lighter pink than the rest of the muscle. During aging the amount of "drip" from the beef cuts of the dairy cow was small. The raw meat of this animal was very difficult to cut into pieces.

The slices of canned meat were observed for their general appearance. Much separation at the primary bundles of muscle fibers was noted. The connective tissue meshwork was either fragile or small in amount. A stringy texture in the meat was associated with the tendency to separate into small bundles of muscle fibers. Many of the samples injected with either of the solutions containing sodium chloride, and a few other samples, had an iridescent appearance at the surface of the slices.

The cans of meat placed in the incubator at 100°F. immediately after removal from the processing retort showed no signs of spoilage after 6 months or more of storage.

The fibrous nature of the cooked meat samples after maceration in the Waring blender during the preparation of pH samples was noted. The semitendinosus, semimembranosus, and biceps femoris were particularly fibrous. The pull on the electric motor of the blender was noticeable when the samples from the aged dairy cow were being macerated.

The liquid from the canned meat was brown, some samples having a yellow cast, others shading toward red. Some variations in the brightness or dullness were observed, but no definite pattern could be distinguished.

Gelation of Liquid

Observations of the liquid from the canned beef after 24 hours of refrigeration showed wide variations in gelation. Some samples were stiff, quivering gels; others were thin, watery liquids. Varying degrees of gelation between these extremes were noted. In general, firmer gels were found among the samples from Animals VI, VII, and VIII, processed 65 minutes, than in those from Animals IX and X, processed 90 minutes. Gelation was less extensive for most of the samples from Animal IX than for those from the other animals.

It appeared that the processing time of 65 minutes was sufficient to bring about degradation of collagen to gelatin, but that, by the end of 90 minutes, the gelatin was also

partially degraded. The differences in results between Animals IX and X, both of which were processed 90 minutes, may be accounted for in the higher initial content of tough connective tissue in the aged dairy cow (Animal X) than in the younger animal. The injection treatment with sodium chloride, lactic acid, or the mixture of two in solution had no consistent effect on gelation of the liquid from the canned meat of any of the animals.

SUMMARY

A study was made of the effect on canned beef of injecting the raw meat with one of the following solutions: sodium chloride, lactic acid, or a mixture of sodium chloride and lactic acid. The concentration of sodium chloride in each of the two salt-containing solutions was 15 per cent, making the proportion of approximately 1.5 grams of salt to 100 grams of meat after injection. The lactic acid solution, as well as the solution with both sodium chloride and lactic acid, contained sufficient acid to give a pH value of 3.4 for the solution.

Five animals were used in the study; four were steers (carcass grade Commercial) and one was an aged dairy cow (carcass grade Cutter). Matching pairs of the rib and loin portions of the longissimus dorsi, psoas major and psoas minor, semitendinosus, semimembranosus, and biceps femoris muscles were separated from the carcass 1 day after slaughter and divided into three or six cuts for aging and canning.

Cuts from the right side of the animal were injected; those from the left side were used as controls. All the cuts were aged 8 days at 34° to 36°F. prior to canning. Samples

for histological study and pH determinations were taken from the fresh muscle, the aged cuts, and from the canned meat. The meat from three steers was processed at 240°F. (10 pounds) for 65 minutes; from the fourth steer and the aged dairy cow, 90 minutes.

The canned meat was sliced on a mechanical slicer and representative slices were rated by the judges for six palatability factors. Slicing quality was evaluated by: (1) number and characteristics of slices obtained, (2) by weight of unsliceable meat, and (3) by judges' scores. Statistical analyses were made of the scores for flavor of the meat, tenderness, juiciness, texture, and slicing quality to determine the significance of the differences. Observations were made of the general appearance of the canned beef and the liquid.

Aroma scores for the canned beef were not affected by the injecting treatments, but were lower for the aged dairy cow than for the other animals.

Flavor, tenderness, and texture scores for the canned beef from the five animals were markedly improved (significant at the .01 level) by injection of the meat with either sodium chloride solution or the combination of sodium chloride and lactic acid. The lactic acid solution alone had no significant effect on these three palatability factors.

The scores for the flavor of the canned beef were not affected by the muscles used, but the beef of the aged dairy cow was undesirably strong flavored.

Tenderness was influenced by the injecting treatments and variations occurred among muscles and among animals.

Differences in texture were found in muscles and in animals. The psoas muscles had the highest rating; the loin portion of the longissimus dorsi and the semimembranosus, the lowest. Beef from the dairy cow had a low texture rating.

The average juiciness scores for the sodium chloride-injected samples of canned beef processed 65 minutes were higher than for the controls, but the differences between the juiciness scores for the injected samples processed 90 minutes and the control samples were not significant. Combined salt and lactic acid solution improved the juiciness of the injected samples compared to control samples, but the differences were significant only at the .05 level. Differences in the lactic acid-treated samples versus controls were within experimental error.

The flavor of liquid from the can was improved by treatment of beef with either of the salt-containing solutions.

Slicing quality of the canned beef was little affected by injection of the raw meat with any of the three solutions

tested. Little variation in sliceability occurred among the cuts processed 65 minutes. Variations did occur among those processed 90 minutes; the beef from the cow sliced better than that from the steer. Muscles were markedly different in sliceability in all but one series of tests. The semiten-dinosus consistently sliced well.

The histological characteristics observed in the longitudinal sections of the rib portion of the longissimus dorsi helped to explain the results secured in the palatability scores. Some disintegration of the fiber striations had occurred after aging 8 days. The extent of this disintegration varied with the muscle and the animal.

The effect of injection with either sodium chloride solution or a mixture of sodium chloride and lactic acid was an increase in the number and extent of the disintegration fissures in the fibers. This disintegration would be expected to contribute to tenderness of the meat, and the judges rated the injected samples receiving either of these treatments as more tender than the controls. Samples injected with only lactic acid appeared much like the control samples in histological features.

The average pH values for the five animals were similar for fresh muscle and for the cuts of beef aged 8 days, regardless of injection treatment. These values were close

to pH 5.45. The meat of the aged dairy cow was higher in pH than the meat of the steers and had the dark red color and the sticky, rubbery feel that are characteristic of meat of high pH. The canned beef samples of all animals were higher in pH than the raw meat. Injection with any of the three solutions had little effect on pH values of the meat after canning.

Small losses in weight were found in the control cuts of beef, but the average weight of the injected samples was greater after 8 days of aging than the initial weight. Thus some of the injected solution remained in the meat. The canned beef was approximately two-thirds meat and one-third liquid. Injection of the raw meat made little difference in the proportion of liquid to meat in the canned product.

CONCLUSIONS

Under the conditions of this study the following conclusions are made:

1. As indicated by palatability scores, the injection of beef cuts with either sodium chloride solution or a combination of sodium chloride and lactic acid solution markedly improved the flavor, tenderness, and texture of the canned beef compared to uninjected control cuts. Juiciness was improved for some of the cuts but not for all. Injection of beef cuts with lactic acid solution had no significant effect on any of the six palatability factors evaluated. Aroma scores were little affected by the injection treatment of the meat. Flavor of the liquid from the can was slightly improved by injection with the salt-containing solutions but not by the lactic acid solution.

2. The slicing quality of the canned meat was fairly low, and no improvement was secured by injection of the raw meat with any of the three solutions tested. Sliceability of the canned beef, however, varied among the muscles and the animals.

3. The tenderness of the canned beef was related to the microscopic changes which occurred in the beef.

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ACKNOWLEDGMENTS

The writer wishes to express sincere appreciation to Professor Belle Lowe for her constant guidance and help throughout this study. The cooperation of Professor R. G. Tischer and Mr. E. A. Kline is deeply appreciated. The writer wishes to thank Professor P. G. Homeyer for assistance in the statistical aspects of the study. Grateful acknowledgment is made of the help of Mrs. Alma R. Plagge in the laboratory work and in the evaluation of the canned meat.

APPENDIX

Table 1. Palatability of Canned Beef. Average palatability scores grouped according to muscle used and processing time. Scoring range is 10 for extremely good quality to 1 for extremely poor quality.

Animal* no.	Sample no.	Palatability scores					
		Aroma	Flavor		Tender- ness	Juici- ness	Tex- ture
			Meat	Liquid			
LONGISSIMUS DORSI MUSCLE, LOIN PORTION							
<u>Sodium chloride injection</u>							
Control samples							
VI	162	7.7	7.0	6.3	6.3	5.7	5.0
VII	188	7.7	7.0	6.3	5.7	5.3	5.0
VIII	211	8.0	7.0	5.7	4.0	5.3	3.3
Av. (3 animals)		7.8	7.0	6.1	5.3	5.4	4.4
IX	234	7.7	6.7	5.0	6.7	4.3	5.0
X	260	5.7	5.0	5.3	5.7	4.3	3.3
Av. (5 animals)		7.4	6.5	5.7	5.7	5.0	4.3
Injected samples							
VI	162	7.7	8.3	7.7	8.0	6.3	6.7
VII	188	7.3	6.7	6.3	5.3	5.0	4.0
VIII	211	7.3	7.3	6.0	4.7	5.3	3.7
Av. (3 animals)		7.4	7.4	6.7	6.0	5.5	4.8
IX	234	7.3	7.7	7.0	8.0	5.0	6.7
X	260	6.7	5.7	5.3	7.7	5.7	5.7
Av. (5 animals)		7.3	7.1	6.5	6.7	5.5	5.4

(continued)

*Animals VI, VII, and VIII were steers, carcass grade Commercial, processed 65 minutes; Animal IX was a steer, carcass grade Commercial, processed 90 minutes; Animal X was a cow, carcass grade Cutter, processed 90 minutes.

Table 1 (continued)

Animal no.	Sample no.	Palatability scores					
		Aroma	Flavor		Tender-ness	Juici-ness	Tex-ture
Meat	Liquid						
LONGISSIMUS DORSI MUSCLE, LOIN PORTION (continued)							
<u>Lactic acid injection</u>							
Control samples							
VI	164	8.3	7.3	6.7	5.3	6.0	4.3
VII	187	7.7	6.7	7.0	5.7	4.7	3.7
VIII	210	8.0	6.7	5.7	4.0	4.3	4.0
Av. (3 animals)		8.0	6.9	6.5	5.0	5.0	4.0
IX	236	8.0	7.0	5.0	6.3	4.0	4.3
X	259	6.3	5.3	5.0	5.3	3.7	3.3
Av. (5 animals)		7.7	6.6	5.9	5.3	4.5	3.9
Injected samples							
VI	164	8.0	8.0	7.3	6.0	5.7	5.3
VII	187	7.7	7.0	6.7	5.3	4.7	4.3
VIII	210	7.7	7.0	6.0	4.0	5.0	3.7
Av. (3 animals)		7.8	7.3	6.7	5.1	5.1	4.4
IX	236	7.3	7.3	5.7	6.7	3.7	4.3
X	259	6.3	5.3	5.7	6.3	4.0	4.0
Av. (5 animals)		7.4	6.9	6.3	5.7	4.6	4.3

(continued)

Table 1 (continued)

Animal no.	Sample no.	Palatability scores					
		Aroma	Flavor		Tender-ness	Juici-ness	Tex-ture
			Meat	Liquid			

LONGISSIMUS DORSI MUSCLE, LOIN PORTION (continued)

Sodium chloride and lactic acid injection

Control samples

VI	163	7.7	8.0	6.7	6.3	7.0	5.3
VII	186	7.7	7.3	8.0	7.7	5.3	6.0
VIII	212	7.7	7.0	5.7	4.7	5.7	4.3
Av. (3 animals)		7.7	7.4	6.8	6.2	6.0	5.2
IX	235	7.3	7.0	5.3	6.0	3.7	4.7
X	258	6.0	6.0	5.3	6.3	4.0	3.7
Av. (5 animals)		7.3	7.1	6.2	6.2	5.1	4.8

Injected samples

VI	163	8.0	8.3	7.0	8.0	6.0	7.0
VII	186	7.7	7.0	7.7	7.7	5.7	5.0
VIII	212	8.0	7.0	7.3	6.3	5.7	5.0
Av. (3 animals)		7.9	7.4	7.3	7.3	5.8	5.7
IX	235	7.7	7.0	7.0	7.7	4.3	6.0
X	258	6.0	6.3	5.7	8.3	5.3	6.0
Av. (5 animals)		7.5	7.1	6.9	7.6	5.4	5.8

(continued)

Table 1 (continued)

Animal no.	Sample no.	Palatability scores					
		Aroma	Flavor		Tender-ness	Juici-ness	Tex-ture
			Meat	Liquid			
LONGISSIMUS DORSI MUSCLE, RIB PORTION							
<u>Sodium chloride injection</u>							
Control samples							
VI	167	7.3	7.3	6.0	4.3	4.3	3.3
VII	189	7.7	7.7	7.0	8.3	7.7	7.0
VIII	214	7.3	8.0	6.3	7.3	6.0	6.0
Av. (3 animals)		7.4	7.3	6.4	6.6	6.0	5.4
IX	239	7.3	8.3	6.3	8.3	5.0	6.0
X	261	6.3	6.0	3.3	7.0	4.3	5.0
Av. (5 animals)		7.2	7.3	5.8	7.0	5.5	5.5
Injected samples							
VI	167	7.7	7.7	7.3	8.0	6.0	6.7
VII	189	8.0	8.7	8.0	9.7	8.0	8.7
VIII	214	8.0	8.7	8.0	9.0	7.3	8.0
Av. (3 animals)		7.9	8.4	7.8	8.9	7.1	7.8
IX	239	7.7	8.3	7.0	8.7	6.0	7.3
X	261	7.0	6.3	3.3	9.3	5.0	6.3
Av. (5 animals)		7.7	7.9	6.7	8.9	6.5	7.4

(continued)

Table 1 (continued)

Animal no.	Sample no.	Palatability scores					Texture
		Aroma	Flavor		Tender-ness	Juici-ness	
			Meat	Liquid			
LONGISSIMUS DORSI MUSCLE, RIB PORTION (continued)							
<u>Lactic acid injection</u>							
Control samples							
VI	165	8.3	7.7	7.0	7.7	6.3	6.7
VII	190	8.0	8.0	7.0	8.0	6.7	6.0
VIII	215	7.7	7.7	5.7	6.3	6.3	5.3
Av. (3 animals)		8.0	7.8	6.6	7.3	6.4	6.0
IX	237	7.3	7.7	5.7	7.7	5.7	5.7
X	262	6.7	6.0	3.0	8.0	4.3	5.3
Av. (5 animals)		7.6	7.4	5.7	7.5	5.9	5.8
Injected samples							
VI	165	8.0	8.0	6.3	7.7	7.7	7.0
VII	190	8.0	7.7	7.7	7.7	6.7	6.3
VIII	215	7.7	7.7	5.7	5.3	6.0	5.0
Av. (3 animals)		7.9	7.8	6.6	6.9	6.8	6.1
IX	237	7.7	7.3	5.7	7.7	5.3	6.3
X	262	6.7	6.3	4.0	9.3	5.3	6.0
Av. (5 animals)		7.6	7.4	5.9	7.5	6.2	6.1

(continued)

Table 1 (continued)

Animal no.	Sample no.	Palatability scores					
		Aroma	Flavor		Tender-ness	Juici-ness	Tex-ture
			Meat	Liquid			

LONGISSIMUS DORSI MUSCLE, RIB PORTION (continued)

Sodium chloride and lactic acid injection

Control samples

VI	166	8.0	7.0	5.7	7.7	6.0	5.7
VII	191	8.7	7.3	6.7	7.7	7.0	6.3
VIII	213	8.0	8.0	6.0	8.0	6.7	7.3
Av. (3 animals)		8.2	7.4	6.1	7.8	6.6	6.4
IX	238	7.3	7.3	5.7	7.3	4.7	5.7
X	263	6.3	6.0	3.0	6.3	4.7	4.0
Av. (5 animals)		7.7	7.1	5.4	7.4	5.8	5.8

Injected samples

VI	166	8.0	7.7	7.3	8.3	7.0	7.3
VII	191	7.7	8.7	8.0	8.0	7.3	6.7
VIII	213	7.7	8.7	7.7	9.3	7.3	8.0
Av. (3 animals)		7.8	8.4	7.7	8.5	7.2	7.3
IX	238	7.3	8.7	8.0	9.0	5.7	7.0
X	263	6.7	6.0	4.0	9.3	4.0	6.3
Av. (5 animals)		7.5	8.0	7.0	8.8	6.3	7.1

(continued)

Table 1 (continued)

Animal no.	Sample no.	Palatability scores					
		Aroma	Flavor		Tender-ness	Juici-ness	Tex-ture
			Meat	Liquid			
PSOAS MAJOR AND PSOAS MINOR MUSCLES							
<u>Sodium chloride injection</u>							
Control samples							
VI	168	8.7	8.0	7.0	9.0	7.0	8.0
VII	193	8.3	7.7	7.0	8.3	7.7	7.7
VIII	218	8.3	8.0	6.0	9.0	6.3	7.7
Av. (3 animals)		8.4	7.9	6.7	8.8	7.0	7.8
IX	240	7.3	7.3	5.7	9.3	6.0	7.3
X	265	6.3	4.3	4.0	9.3	5.0	6.0
Av. (5 animals)		7.8	7.1	5.9	9.0	6.4	7.3
Injected samples							
VI	168	8.0	8.3	7.3	9.3	7.3	8.3
VII	193	7.7	8.7	8.0	9.7	6.7	7.7
VIII	218	8.3	8.7	7.7	9.3	7.3	8.0
Av. (3 animals)		8.0	8.6	7.7	9.4	7.1	8.0
IX	240	6.7	8.0	7.3	10.0	5.7	8.0
X	265	6.3	5.3	4.7	9.7	6.0	6.7
Av. (5 animals)		7.4	7.8	7.0	9.6	6.6	7.7

(continued)

Table 1 (continued)

Animal no.	Sample no.	Palatability scores					
		Aroma	Flavor		Tender-ness	Juici-ness	Tex-ture
			Meat	Liquid			
PSOAS MAJOR AND PSOAS MINOR MUSCLES (continued)							
<u>Lactic acid injection</u>							
Control samples							
VI	169	8.7	7.3	6.3	9.0	7.0	8.7
VII	194	8.3	8.3	7.3	9.0	7.3	7.3
VIII	216	7.7	8.3	6.7	8.7	7.0	7.3
Av. (3 animals)		8.2	8.0	6.8	8.9	7.1	7.8
IX	241	7.7	7.7	5.3	9.0	5.7	7.7
X	266	5.0	3.3	2.7	9.0	4.3	5.0
Av. (5 animals)		7.5	7.0	5.7	8.9	6.3	7.2
Injected samples							
VI	169	8.3	7.7	7.0	8.7	7.3	8.3
VII	194	8.0	7.0	6.3	7.7	6.3	6.7
VIII	216	8.0	8.0	6.7	8.3	6.3	6.7
Av. (3 animals)		8.1	7.6	6.7	8.2	6.6	7.2
IX	241	7.3	7.3	5.3	8.3	5.7	7.0
X	266	6.3	4.7	3.0	9.0	4.7	6.0
Av. (5 animals)		7.6	6.9	5.7	8.4	6.1	6.9

(continued)

Table 1 (continued)

Animal no.	Sample no.	Palatability scores					
		Aroma	Flavor		Tender-ness	Juici-ness	Tex-ture
			Meat	Liquid			

PSOAS MAJOR AND PSOAS MINOR MUSCLES (continued)

Sodium chloride and lactic acid injection

Control samples

VI	170	8.3	7.7	7.3	8.7	6.7	8.0
VII	192	8.0	7.7	7.0	8.0	7.3	7.3
VIII	217	7.7	8.0	6.3	8.7	6.7	7.0
Av. (3 animals)		8.0	7.8	6.9	8.5	6.9	7.4
IX	242	7.3	7.3	5.7	9.3	5.7	7.7
X	264	6.3	4.0	3.0	8.7	4.3	5.3
Av. (5 animals)		7.5	6.9	5.9	8.7	6.1	7.1

Injected samples

VI	170	8.3	9.0	7.7	9.7	8.3	9.3
VII	192	8.0	8.7	8.0	9.7	7.3	8.0
VIII	217	8.7	8.7	8.3	9.7	7.7	8.7
Av. (3 animals)		8.3	8.8	8.0	9.7	7.8	8.7
IX	242	7.7	8.3	7.0	9.7	6.3	8.0
X	264	6.7	5.3	4.7	9.3	5.0	6.7
Av. (5 animals)		7.9	8.0	7.1	9.6	6.9	8.1

(continued)

Table 1 (continued)

Animal no.	Sample no.	Palatability scores					
		Aroma	Flavor		Tender-ness	Juici-ness	Tex-ture
			Meat	Liquid			
SEMITENDINOSUS MUSCLE							
<u>Sodium chloride injection</u>							
Control samples							
VI	172	8.3	7.7	6.7	8.3	6.0	6.3
VII	197	8.3	7.7	6.0	8.0	6.0	6.3
VIII	219	7.7	7.7	5.7	7.7	6.7	6.7
Av. (3 animals)		8.1	7.7	6.1	8.0	6.2	6.4
IX	244	8.3	7.7	5.7	7.7	4.3	6.0
X	269	6.0	5.3	4.3	7.7	4.3	5.0
Av. (5 animals)		7.7	7.2	5.7	7.9	5.5	6.1
Injected samples							
VI	172	8.3	8.3	7.0	8.7	7.0	8.0
VII	197	8.3	8.7	7.7	9.3	7.3	8.0
VIII	219	8.0	8.0	7.0	7.7	6.7	7.3
Av. (3 animals)		8.2	8.3	7.2	8.6	7.0	7.8
IX	244	7.7	8.0	7.3	9.3	4.3	7.3
X	269	6.3	6.0	5.7	8.7	5.3	6.3
Av. (5 animals)		7.7	7.8	6.9	8.7	6.1	7.4

(continued)

Table 1 (continued)

Animal no.	Sample no.	Palatability scores					
		Aroma	Flavor		Tender-ness	Juici-ness	Tex-ture
			Meat	Liquid			

SEMITENDINOSUS MUSCLE (continued)

Lactic acid injection

Control samples

VI	173	8.0	8.0	6.0	8.7	7.3	7.7
VII	195	7.7	8.0	7.0	8.0	6.7	6.7
VIII	220	7.7	7.7	6.3	7.7	6.3	6.7
Av. (3 animals)		7.8	7.9	6.4	8.1	6.8	7.0
IX	245	7.7	7.7	6.0	8.3	6.0	7.7
X	267	6.3	5.3	4.3	6.7	5.0	4.3
Av. (5 animals)		7.5	7.3	5.9	7.9	6.3	6.6

Injected samples

VI	173	8.3	8.0	6.3	7.7	7.3	7.7
VII	195	8.0	7.3	5.7	8.0	5.7	6.0
VIII	220	8.0	7.7	6.3	8.0	6.3	6.7
Av. (3 animals)		8.1	7.7	6.1	7.9	6.4	6.8
IX	245	7.7	7.7	5.7	7.7	5.7	6.3
X	267	6.0	5.7	4.7	7.0	4.7	5.7
Av. (5 animals)		7.6	7.3	5.7	7.7	5.9	6.5

(continued)

Table 1 (continued)

Animal no.	Sample no.	Palatability scores					
		Aroma	Flavor		Tender-ness	Juici-ness	Tex-ture
			Meat	Liquid			
SEMITENDINOSUS MUSCLE (continued)							
<u>Sodium chloride and lactic acid injection</u>							
Control samples							
VI	171	7.7	7.3	6.0	7.7	6.0	6.7
VII	196	8.0	8.0	6.3	7.7	6.0	6.3
VIII	221	7.7	8.0	6.0	7.3	6.3	6.7
Av. (3 animals)		7.8	7.8	6.1	7.6	6.1	6.6
IX	243	7.7	8.0	6.0	7.3	5.0	5.7
X	268	6.3	5.3	4.7	7.7	5.0	5.7
Av. (5 animals)		7.5	7.3	5.8	7.5	5.7	6.2
Injected samples							
VI	171	8.3	7.7	7.3	8.3	7.3	7.7
VII	196	8.0	8.7	7.7	9.3	6.7	7.7
VIII	221	8.0	8.3	7.3	8.0	7.7	8.3
Av. (3 animals)		8.1	8.2	7.4	8.5	7.2	7.9
IX	243	7.7	8.3	7.0	9.0	4.3	7.7
X	268	6.0	6.0	6.0	9.3	5.3	6.3
Av. (5 animals)		7.6	7.8	7.1	8.8	6.3	7.5

(continued)

Table 1 (continued)

Animal no.	Sample no.	Palatability scores					
		Aroma	Flavor		Tender-ness	Juici-ness	Tex-ture
			Meat	Liquid			
SEMIMEMBRANOSUS MUSCLE							
<u>Sodium chloride injection</u>							
Control samples							
VI	177	7.3	7.3	5.3	5.7	5.0	4.0
	179	7.3	7.0	7.3	7.3	4.7	5.0
Av.		7.3	7.2	6.3	6.5	4.8	4.5
VII	199	7.7	7.0	6.7	6.3	5.0	5.0
	200	7.7	7.3	6.3	7.0	6.7	6.0
Av.		7.7	7.2	6.5	6.6	5.8	5.5
VIII	222	7.3	7.3	6.3	7.3	4.7	5.7
	226	7.3	7.3	6.7	8.3	5.0	6.3
Av.		7.3	7.3	6.5	7.8	4.8	6.0
Av. (3 animals)		7.4	7.2	6.4	7.0	5.1	5.3
IX	249	6.7	7.3	5.7	6.7	4.7	4.0
	251	7.3	6.7	6.0	7.0	3.3	4.7
Av.		7.0	7.0	5.8	6.8	4.0	4.4
X	271	7.0	5.7	4.7	6.0	4.3	3.7
	272	6.7	5.3	4.7	6.7	4.3	5.0
Av.		6.8	5.5	4.7	6.4	4.3	4.4
Av. (5 animals)		7.2	6.8	6.0	6.8	4.7	5.0

(continued)

Table 1 (continued)

Animal no.	Sample no.	Palatability scores					
		Aroma	Flavor		Tender-ness	Juici-ness	Tex- ture
			Meat	Liquid			
SEMIMEMBRANOSUS MUSCLE (continued)							
<u>Sodium chloride injection (continued)</u>							
Injected samples							
VI	177	7.3	8.0	6.7	7.3	6.3	6.7
	179	7.7	7.3	6.3	7.0	6.0	5.0
Av.		7.5	7.6	6.5	7.2	6.2	5.8
VII	199	7.7	8.0	7.3	8.3	5.3	7.0
	200	8.0	8.7	7.3	9.0	6.3	8.0
Av.		7.8	8.4	7.3	8.6	5.8	7.5
VIII	222	8.0	7.7	7.0	8.7	6.3	6.7
	226	7.7	8.3	7.3	8.0	6.0	6.7
Av.		7.8	8.0	7.2	8.4	6.2	6.7
Av. (3 animals)		7.7	8.0	7.0	8.1	6.1	6.7
IX	249	7.0	7.7	7.0	7.3	4.0	4.7
	251	8.0	8.0	7.3	8.3	3.3	5.3
Av.		7.5	7.8	7.2	7.8	3.6	5.0
X	271	7.0	6.3	6.3	8.0	5.7	5.3
	272	6.7	6.7	6.0	8.7	4.3	5.7
Av.		6.8	6.5	6.2	8.4	5.0	5.5
Av. (5 animals)		7.5	7.7	6.9	8.1	5.4	6.1

(continued)

Table 1 (continued)

Animal no.	Sample no.	Palatability scores					
		Aroma	Flavor		Tender-ness	Juici-ness	Tex-ture
			Meat	Liquid			
SEMIMEMBRANOSUS MUSCLE (continued)							
<u>Lactic acid injection</u>							
Control samples							
VI	175	7.3	7.3	6.0	6.7	5.3	4.0
	176	7.7	6.7	6.3	7.7	3.7	5.7
Av.		7.5	7.0	6.2	7.2	4.5	4.8
VII	198	7.7	6.7	6.3	6.7	5.0	5.7
	202	7.3	7.7	7.0	6.3	6.0	5.0
Av.		7.5	7.2	6.6	6.5	5.5	5.4
VIII	225	7.7	7.0	6.3	6.3	5.0	4.0
	227	7.3	7.3	5.7	5.7	4.3	4.3
Av.		7.5	7.2	6.0	6.0	4.6	4.2
Av. (3 animals)		7.5	7.1	6.3	6.6	4.9	4.8
IX	247	7.0	7.0	5.3	8.0	2.7	4.7
	248	7.3	6.3	5.7	7.7	3.0	4.7
Av.		7.2	6.5	5.5	7.8	2.8	4.7
X	270	6.7	6.0	5.0	7.3	4.0	4.0
	274	7.0	6.0	4.3	7.3	4.3	4.7
Av.		6.8	6.0	4.6	7.3	4.2	4.4
Av. (5 animals)		7.3	6.8	5.8	7.0	4.3	4.7

(continued)

Table 1 (continued)

Animal no.	Sample no.	Palatability scores					
		Aroma	Flavor		Tender-ness	Juici-ness	Tex-ture
			Meat	Liquid			
SEMIMEMBRANOSUS MUSCLE (continued)							
<u>Lactic acid injection (continued)</u>							
Injected samples							
VI	175	7.7	7.3	7.0	7.7	4.0	5.0
	176	7.7	7.7	6.0	7.3	5.3	5.7
Av.		7.7	7.5	6.5	7.5	4.6	5.4
VII	198	8.0	7.3	7.0	7.3	5.0	6.0
	202	7.7	8.0	6.7	7.7	7.0	7.0
Av.		7.8	7.6	6.8	7.5	6.0	6.5
VIII	225	7.3	7.0	6.0	5.3	6.3	4.0
	227	7.7	7.3	6.0	6.7	3.3	4.7
Av.		7.5	7.2	6.0	6.0	4.8	4.4
Av. (3 animals)		7.7	7.4	6.4	7.0	5.1	5.4
IX	247	7.0	6.7	5.0	8.0	3.0	4.3
	248	7.3	7.0	5.7	7.0	3.3	5.0
Av.		7.2	6.8	5.4	7.5	3.2	4.6
X	270	6.7	5.7	5.0	7.7	5.0	5.0
	274	6.3	5.7	5.0	6.7	5.0	4.7
Av.		6.5	5.7	5.0	7.2	5.0	4.8
Av. (5 animals)		7.3	7.0	5.9	7.1	4.7	5.1

(continued)

Table 1 (continued)

Animal no.	Sample no.	Palatability scores					
		Aroma	Flavor		Tender-ness	Juici-ness	Tex- ture
			Meat	Liquid			
SEMIMEMBRANOSUS MUSCLE (continued)							
<u>Sodium chloride and lactic acid injection</u>							
Control samples							
VI	174	7.7	7.7	6.3	7.3	5.3	4.7
	178	7.7	7.0	6.7	6.0	3.7	3.7
Av.		7.7	7.4	6.5	6.6	4.5	4.2
VII	201	7.7	6.7	6.7	6.0	4.7	5.0
	203	8.0	8.0	6.3	8.3	5.7	6.3
Av.		7.8	7.4	6.5	7.2	5.2	5.6
VIII	223	7.3	7.0	7.7	6.0	5.0	4.7
	224	7.3	7.3	6.3	5.3	5.3	4.3
Av.		7.3	7.2	7.0	5.6	5.2	4.5
Av. (3 animals)		7.6	7.3	6.7	6.5	5.0	4.8
IX	246	7.3	6.7	5.7	6.7	3.0	4.0
	250	7.7	6.3	5.3	7.7	3.0	5.0
Av.		7.5	6.5	5.5	7.2	3.0	4.5
X	273	7.0	5.7	5.0	6.3	3.7	4.0
	275	6.3	5.7	4.0	7.7	5.0	4.7
Av.		6.6	5.7	4.5	7.0	4.4	4.4
Av. (5 animals)		7.4	6.8	6.0	6.7	4.5	4.6

(continued)

Table 1 (continued)

Animal no.	Sample no.	Palatability scores					
		Aroma	Flavor		Tender-ness	Juici-ness	Tex-ture
			Meat	Liquid			
SEMIMEMBRANOSUS MUSCLE (continued)							
<u>Sodium chloride and lactic acid injection (continued)</u>							
Injected samples							
VI	174	7.7	8.0	7.7	8.3	6.3	7.0
	178	7.3	7.7	7.7	7.0	5.0	5.7
Av.		7.5	7.8	7.7	7.6	5.6	6.4
VII	201	8.0	8.3	8.0	8.3	6.7	7.3
	203	7.7	8.0	7.0	8.7	6.7	7.7
Av.		7.8	8.2	7.5	8.5	6.7	7.5
VIII	223	7.7	8.0	6.0	7.7	5.7	6.3
	224	8.0	8.3	8.0	7.7	5.3	6.7
Av.		7.8	8.2	7.0	7.7	5.5	6.5
Av. (3 animals)		7.7	8.1	7.4	7.9	5.9	6.8
IX	246	7.3	7.7	7.3	8.0	4.7	6.0
	250	7.7	8.0	7.3	9.0	4.7	6.7
Av.		7.5	7.8	7.3	8.5	4.7	6.4
X	273	7.0	6.3	6.0	8.7	5.0	6.0
	275	6.7	6.7	6.0	8.7	4.0	5.3
Av.		6.8	6.5	6.0	8.7	4.5	5.6
Av. (5 animals)		7.5	7.7	7.1	8.2	5.4	6.5

(continued)

Table 1 (continued)

Animal no.	Sample no.	Palatability scores					
		Aroma	Flavor		Tender-ness	Juici-ness	Tex-ture
			Meat	Liquid			
BICEPS FEMORIS MUSCLE							
<u>Sodium chloride injection</u>							
Control samples							
VI	184	8.0	8.0	6.0	8.3	7.0	8.0
	185	8.0	6.7	6.3	7.7	6.3	6.0
Av.		8.0	7.4	6.2	8.0	6.6	7.0
VII	205	8.3	7.3	6.3	7.7	6.3	6.3
	207	7.7	8.0	6.7	7.3	6.0	6.3
Av.		8.0	7.6	6.5	7.5	6.2	6.3
VIII	228	7.7	7.7	6.3	8.0	7.7	5.7
	230	8.0	7.0	6.3	6.7	4.7	4.7
Av.		7.8	7.4	6.3	7.4	6.2	5.2
Av. (3 animals)		7.9	7.5	6.3	7.6	6.3	6.2
IX	256	7.3	6.3	5.3	7.3	4.0	4.7
	257	7.3	6.7	5.7	8.7	3.0	6.0
Av.		7.3	6.5	5.5	8.0	3.5	5.4
X	277	5.7	3.3	3.3	6.7	3.7	3.3
	279	6.7	3.7	3.7	8.3	5.0	6.0
Av.		6.2	3.5	3.5	7.5	4.4	4.6
Av. (5 animals)		7.5	6.5	5.6	7.7	5.4	5.7

(continued)

Table 1 (continued)

Animal no.	Sample no.	Palatability scores					
		Aroma	Flavor		Tender-ness	Juici-ness	Tex-ture
			Meat	Liquid			
BICEPS FEMORIS MUSCLE (continued)							
<u>Sodium chloride injection (continued)</u>							
Injected samples							
VI	184	7.7	7.7	8.0	6.3	7.3	6.3
	185	7.7	7.7	7.3	8.7	7.0	7.7
Av.		7.7	7.7	7.6	7.5	7.2	7.0
VII	205	7.7	8.0	8.0	8.3	6.0	7.3
	207	7.7	8.3	7.3	9.3	7.0	8.0
Av.		7.7	8.2	7.6	8.8	6.5	7.6
VIII	228	8.0	8.0	7.0	9.0	7.3	8.3
	230	7.7	7.7	7.3	8.7	6.3	7.3
Av.		7.8	7.8	7.2	8.8	6.8	7.8
Av. (3 animals)		7.7	7.9	7.5	8.4	6.8	7.5
IX	256	7.0	7.7	7.0	8.0	4.7	6.0
	257	7.3	7.3	7.0	9.0	3.7	7.0
Av.		7.2	7.5	7.0	8.5	4.2	6.5
X	277	6.0	4.3	5.7	7.7	4.3	5.7
	279	7.3	4.3	5.7	8.7	6.0	6.3
Av.		6.6	4.3	5.7	8.2	5.2	6.0
Av. (5 animals)		7.4	7.1	7.0	8.4	6.0	7.0

(continued)

Table 1 (continued)

Animal no.	Sample no.	Palatability scores					
		Aroma	Flavor		Tender-ness	Juici-ness	Tex-ture
			Meat	Liquid			

BICEPS FEMORIS MUSCLE (continued)

Lactic acid injection

Control samples

VI	181	8.0	7.7	6.3	8.3	6.3	7.7
	183	7.7	7.0	6.7	7.3	6.7	6.7
Av.		7.8	7.4	6.5	7.8	6.5	7.2
VII	204	8.0	7.3	6.0	8.7	6.7	6.7
	206	7.7	8.0	7.3	8.0	6.0	6.7
Av.		7.8	7.6	6.6	8.4	6.4	6.7
VIII	232	8.3	7.3	6.0	7.0	5.3	6.3
	233	7.3	7.3	6.3	7.7	5.3	5.7
Av.		7.8	7.3	6.2	7.4	5.3	6.0
Av. (3 animals)		7.8	7.4	6.4	7.9	6.1	6.6
IX	253	7.0	7.3	5.3	8.3	6.0	6.3
	255	7.0	7.0	5.3	7.3	3.3	4.3
Av.		7.0	7.2	5.3	7.8	4.6	5.3
X	276	5.0	3.7	5.0	7.0	3.3	4.0
	278	6.3	4.3	3.0	7.3	3.7	5.0
Av.		5.6	4.0	4.0	7.2	3.5	4.5
Av. (5 animals)		7.2	6.7	5.7	7.7	5.3	5.9

(continued)

Table 1 (continued)

Animal no.	Sample no.	Palatability scores					
		Aroma	Flavor		Tender-ness	Juici-ness	Tex-ture
			Meat	Liquid			
BICEPS FEMORIS MUSCLE (continued)							
<u>Lactic acid injection (continued)</u>							
Injected samples							
VI	181	7.7	7.3	6.7	8.7	7.0	7.7
	183	8.0	7.0	6.3	8.0	5.7	6.7
Av.		7.8	7.2	6.5	8.4	6.4	7.2
VII	204	8.0	8.0	7.3	8.7	7.7	7.0
	206	7.7	8.0	7.0	8.3	6.3	7.3
Av.		7.8	8.0	7.2	8.5	7.0	7.2
VIII	232	8.3	7.7	6.3	7.0	6.3	6.0
	233	7.3	7.7	5.7	7.7	5.7	6.3
Av.		7.8	7.7	6.0	7.4	6.0	6.2
Av. (3 animals)		7.8	7.6	6.6	8.1	6.5	6.9
IX	253	7.7	7.3	5.3	8.3	4.3	6.0
	255	7.0	6.3	5.7	7.0	2.7	5.0
Av.		7.4	6.8	5.5	7.6	3.5	5.5
X	276	5.7	3.7	4.3	7.0	4.3	4.0
	278	7.0	4.0	5.0	7.7	5.3	5.0
Av.		6.4	3.8	4.6	7.4	4.8	4.5
Av. (5 animals)		7.4	6.7	6.0	7.9	5.5	6.1

(continued)

Table 1 (continued)

Animal no.	Sample no.	Palatability scores					
		Aroma	Flavor		Tender-ness	Juici-ness	Tex-ture
			Meat	Liquid			
BICEPS FEMORIS MUSCLE (continued)							
<u>Sodium chloride and lactic acid injection</u>							
Control samples							
VI	180	7.7	8.0	7.0	9.0	7.7	8.3
	182	8.0	7.3	7.0	9.0	7.0	7.0
Av.		7.8	7.6	7.0	9.0	7.4	7.6
VII	208	7.7	7.3	6.7	7.0	7.3	6.3
	209	8.3	8.0	6.3	8.3	6.3	6.3
Av.		8.0	7.6	6.5	7.6	6.8	6.3
VIII	229	7.7	8.0	6.3	8.3	7.0	7.0
	231	7.7	7.0	5.7	7.3	5.7	5.7
Av.		7.7	7.5	6.0	7.8	6.4	6.4
Av. (3 animals)		7.8	7.6	6.5	8.1	6.9	6.8
IX	252	6.7	6.7	6.0	9.3	4.3	6.3
	254	7.7	7.0	5.0	8.0	5.7	5.7
Av.		7.2	6.8	5.5	8.6	5.0	6.0
X	280	5.3	4.3	3.7	8.0	3.0	5.0
	281	6.7	4.0	4.0	8.7	4.7	6.0
Av.		6.0	4.2	3.8	8.4	3.8	5.5
Av. (5 animals)		7.3	6.7	5.8	8.3	5.9	6.4

(continued)

Table 1 (continued)

Animal no.	Sample no.	Palatability scores					
		Aroma	Flavor		Tender-ness	Juici-ness	Tex-ture
			Meat	Liquid			
BICEPS FEMORIS MUSCLE (continued)							
<u>Sodium chloride and lactic acid injection (continued)</u>							
Injected samples							
VI	180	8.0	7.7	7.0	9.0	7.3	8.3
	182	8.0	8.0	7.7	8.7	7.3	8.0
Av.		8.0	7.8	7.4	8.8	7.3	8.2
VII	208	7.7	8.0	7.7	8.0	7.0	7.7
	209	8.3	7.7	7.7	8.7	6.7	7.3
Av.		8.0	7.8	7.7	8.4	6.8	7.5
VIII	229	7.7	7.7	8.0	9.0	6.0	7.7
	231	8.0	8.3	7.3	9.0	6.3	7.7
Av.		7.8	8.0	7.6	9.0	6.2	7.7
Av. (3 animals)		7.9	7.9	7.6	8.7	6.8	7.8
IX	252	7.0	8.0	7.0	9.7	4.7	7.0
	254	7.3	8.0	7.0	9.0	5.7	7.0
Av.		7.2	8.0	7.0	9.4	5.2	7.0
X	280	5.7	5.3	5.7	8.3	4.7	5.7
	281	7.0	4.3	4.7	8.7	6.0	6.3
Av.		6.4	4.8	5.2	8.5	5.4	6.0
Av. (5 animals)		7.5	7.3	7.0	8.8	6.2	7.3

Table 2. Slicing Quality of Canned Beef. The number of slices obtained grouped according to muscle used and processing time.

Animal* no.	Sample no.	No. slices obtained		Possible no. slices	Slices obtained		Total
		Firm	Crumbly		Firm %	Crumbly %	
LONGISSIMUS DORSI MUSCLE, LOIN PORTION							
<u>Sodium chloride injection</u>							
Control samples							
VI	162	4	4	10	40.0	40.0	80.0
VII	188	0	3	11	0	27.3	27.3
VIII	211	0	7	12	0	58.3	58.3
Av. (3 animals)		1.3	4.7	11.0	13.3	41.9	55.2
IX	234	0	3	11	0	27.3	27.3
X	260	0	6	11	0	54.5	54.5
Av. (5 animals)		0.8	4.6	11.0	8.0	41.5	49.5
Injected samples							
VI	162	5	3	11	45.4	27.3	72.7
VII	188	0	4	13	0	30.8	30.8
VIII	211	0	2	13	0	15.4	15.4
Av. (3 animals)		1.7	3.0	12.3	15.1	24.5	39.6
IX	234	0	0	11	0	0	0
X	260	8	1	12	66.7	8.3	75.0
Av. (5 animals)		2.6	2.0	12.0	22.4	16.4	38.8

(continued)

*Animals VI, VII, and VIII were steers, carcass grade Commercial, processed 65 minutes; Animal IX was a steer carcass grade Commercial, processed 90 minutes; Animal X was a cow, carcass grade Cutter, processed 90 minutes.

Table 2 (continued)

Animal no.	Sample no.	No. slices obtained		Possible no. slices	Slices obtained		Total
		Firm	Crumbly		Firm %	Crumbly %	

LONGISSIMUS DORSI MUSCLE, LOIN PORTION (continued)

Lactic acid injection

Control samples

VI	164	6	4	11	54.5	36.4	90.9
VII	187	0	0	13	0	0	0
VIII	210	0	0	12	0	0	0
Av. (3 animals)		2.0	1.3	12.0	18.2	12.1	30.3
IX	236	0	2	11	0	18.2	18.2
X	259	0	0	11	0	0	0
Av. (5 animals)		1.2	1.2	11.6	10.9	10.9	21.8

Injected samples

VI	164	6	4	12	50.0	33.3	83.3
VII	187	0	3	11	0	27.3	27.3
VIII	210	7	2	12	58.3	16.7	75.0
Av. (3 animals)		4.3	3.0	11.7	36.1	25.8	61.9
IX	236	0	2	12	0	16.7	16.7
X	259	4	3	11	36.4	27.3	63.7
Av. (5 animals)		3.4	2.8	11.6	28.9	24.3	53.2

(continued)

Table 2 (continued)

Animal no.	Sample no.	No. slices obtained		Possible no. slices	Slices obtained		
		Firm	Crumbly		Firm %	Crumbly %	Total %

LONGISSIMUS DORSI MUSCLE, LOIN PORTION (continued)

Sodium chloride and lactic acid injection

Control samples

VI	163	0	7	11	0	63.6	63.6
VII	186	2	6	11	18.2	54.5	72.7
VIII	212	0	5	13	0	38.5	38.5
Av. (3 animals)		0.7	6.0	11.7	6.1	52.2	58.3
IX	235	0	0	10	0	0	0
X	258	7	1	11	63.6	9.1	72.7
Av. (5 animals)		1.8	3.8	11.2	16.4	33.1	49.5

Injected samples

VI	163	8	2	12	66.7	16.7	83.4
VII	186	2	2	12	16.7	16.7	33.4
VIII	212	2	7	12	16.7	58.3	75.0
Av. (3 animals)		4.0	3.7	12.0	33.4	30.6	63.9
IX	235	0	0	12	0	0	0
X	258	2	6	11	18.2	54.5	72.7
Av. (5 animals)		2.8	3.4	11.8	23.7	29.2	52.9

(continued)

Table 2 (continued)

Animal no.	Sample no.	No. slices obtained		Possible no. slices	Slices obtained		
		Firm	Crumbly		Firm %	Crumbly %	Total %
LONGISSIMUS DORSI MUSCLE, RIB PORTION							
<u>Sodium chloride injection</u>							
Control samples							
VI	167	10	0	10	100.0	0	100.0
VII	189	2	5	10	20.0	50.0	70.0
VIII	214	8	1	11	72.7	9.1	81.8
Av. (3 animals)		6.7	2.0	10.3	64.2	19.7	83.9
IX	239	8	1	10	80.0	10.0	90.0
X	261	2	5	10	20.0	50.0	70.0
Av. (5 animals)		6.0	2.4	10.2	58.5	23.8	82.4
Injected samples							
VI	167	9	0	10	90.0	0	90.0
VII	189	3	2	10	30.0	20.0	50.0
VIII	214	1	7	10	10.0	70.0	80.0
Av. (3 animals)		4.3	3.0	10.0	43.3	30.0	73.3
IX	239	3	3	10	30.0	30.0	60.0
X	261	0	7	10	0	70.0	70.0
Av. (5 animals)		3.2	3.8	10.0	32.0	38.0	70.0

(continued)

Table 2 (continued)

Animal no.	Sample no.	No. slices obtained		Possible no. slices	Slices obtained		Total
		Firm	Crumbly		Firm	Crumbly	
					%	%	%

LONGISSIMUS DORSI MUSCLE, RIB PORTION (continued)

Lactic acid injection

Control samples

VI	165	1	7	11	9.1	63.6	72.7
VII	190	7	4	11	63.6	36.4	100.0
VIII	215	5	4	10	50.0	40.0	90.0
Av. (3 animals)		4.3	5.0	10.7	40.9	46.7	87.6
IX	237	5	4	11	45.4	36.4	81.8
X	262	6	2	10	60.0	20.0	80.0
Av. (5 animals)		4.8	4.2	10.6	45.6	39.3	84.9

Injected samples

VI	165	0	4	10	0	40.0	40.0
VII	190	9	2	11	81.8	18.2	100.0
VIII	215	9	2	11	81.8	18.2	100.0
Av. (3 animals)		6.0	2.7	10.7	54.5	25.5	80.0
IX	237	6	2	11	54.5	18.2	72.7
X	262	7	1	9	77.8	11.1	88.9
Av. (5 animals)		6.2	2.2	10.4	59.2	21.1	80.3

(continued)

Table 2 (continued)

Animal no.	Sample no.	No. slices obtained		Possible no. slices	Slices obtained		Total
		Firm	Crumbly		Firm %	Crumbly %	

LONGISSIMUS DORSI MUSCLE, RIB PORTION (continued)

Sodium chloride and lactic acid injection

Control samples

VI	166	0	5	10	0	50.0	50.0
VII	191	9	1	11	81.8	9.1	90.9
VIII	213	2	7	10	20.0	70.0	90.0
Av. (3 animals)		3.7	4.3	10.3	33.9	43.0	77.0
IX	238	0	2	10	0	20.0	20.0
X	263	0	1	12	0	8.3	8.3
Av. (5 animals)		2.2	3.2	10.6	20.4	31.5	51.8

Injected samples

VI	166	0	8	10	0	80.0	80.0
VII	191	1	8	10	10.0	80.0	90.0
VIII	213	2	7	10	20.0	70.0	90.0
Av. (3 animals)		1.0	7.7	10.0	10.0	76.7	86.7
IX	238	0	2	10	0	20.0	20.0
X	263	6	1	10	60.0	10.0	70.0
Av. (5 animals)		1.8	5.2	10.0	18.0	52.0	70.0

(continued)

Table 2 (continued)

Animal no.	Sample no.	No. slices obtained		Possible no. slices	Slices obtained		Total
		Firm	Crumbly		Firm %	Crumbly %	
PSOAS MAJOR AND PSOAS MINOR MUSCLES							
<u>Sodium chloride injection</u>							
Control samples							
VI	168	2	8	10	20.0	80.0	100.0
VII	193	0	6	11	0	54.5	54.5
VIII	218	4	5	11	36.4	45.4	81.8
Av. (3 animals)		2.0	6.3	10.7	18.8	60.0	78.8
IX	240	0	6	13	0	46.2	46.2
X	265	0	9	12	0	75.0	75.0
Av. (5 animals)		1.2	6.8	11.4	11.3	60.2	71.5
Injected samples							
VI	168	0	7	10	0	70.0	70.0
VII	193	0	2	10	0	20.0	20.0
VIII	218	1	3	11	9.1	27.3	36.4
Av. (3 animals)		0.3	4.0	10.3	3.0	39.1	42.1
IX	240	0	0	11	0	0	0
X	265	8	2	12	66.7	16.7	83.4
Av. (5 animals)		1.8	2.8	10.8	15.2	26.8	42.0

(continued)

Table 2 (continued)

Animal no.	Sample no.	No. slices obtained		Possible no. slices	Slices obtained		Total
		Firm	Crumbly		Firm %	Crumbly %	

PSOAS MAJOR AND PSOAS MINOR MUSCLES (continued)

Lactic acid injection

Control samples

VI	169	3	7	10	30.0	70.0	100.0
VII	194	0	6	10	0	60.0	60.0
VIII	216	1	4	12	8.3	33.3	41.6
Av. (3 animals)		1.3	5.7	10.7	12.8	54.4	67.2
IX	241	0	3	11	0	27.3	27.3
X	266	7	2	12	58.3	16.7	75.0
Av. (5 animals)		2.2	4.4	11.0	19.3	41.5	60.8

Injected samples

VI	169	0	8	10	0	80.0	80.0
VII	194	0	1	10	0	10.0	10.0
VIII	216	0	2	11	0	18.2	18.2
Av. (3 animals)		0	3.7	10.3	0	36.1	36.1
IX	241	0	2	9	0	22.2	22.2
X	266	5	4	11	45.4	36.4	81.8
Av. (5 animals)		1.0	3.4	10.2	9.1	33.4	42.4

(continued)

Table 2 (continued)

Animal no.	Sample no.	No. slices obtained		Possible no. slices	Slices obtained		
		Firm	Crumbly		Firm %	Crumbly %	Total %
PSOAS MAJOR AND PSOAS MINOR MUSCLES (continued)							
<u>Sodium chloride and lactic acid injection</u>							
Control samples							
VI	170	3	6	10	30.0	60.0	90.0
VII	192	0	3	11	0	27.3	27.3
VIII	217	0	7	12	0	58.3	58.3
Av. (3 animals)		1.0	5.3	11.0	10.0	48.5	58.5
IX	242	2	3	9	22.2	33.3	55.5
X	264	0	4	12	0	33.3	33.3
Av. (5 animals)		1.0	4.6	10.8	10.4	42.4	52.9
Injected samples							
VI	170	0	8	10	0	80.0	80.0
VII	192	0	2	10	0	20.0	20.0
VIII	217	1	5	10	10.0	50.0	60.0
Av. (3 animals)		0.3	5.0	10.0	3.3	50.0	53.3
IX	242	0	2	10	0	20.0	20.0
X	264	0	6	11	0	54.5	54.5
Av. (5 animals)		0.2	4.6	10.2	2.0	44.9	46.9

(continued)

Table 2 (continued)

Animal no.	Sample no.	No. slices obtained		Possible no. slices	Slices obtained		
		Firm	Crumbly		Firm %	Crumbly %	Total %
SEMITENDINOSUS MUSCLE							
<u>Sodium chloride injection</u>							
Control samples							
VI	172	8	1	10	80.0	10.0	90.0
VII	197	11	1	12	91.7	8.3	100.0
VIII	219	10	0	10	100.0	0	100.0
Av. (3 animals)		9.7	0.7	10.7	90.6	6.1	96.7
IX	244	8	1	10	80.0	10.0	90.0
X	269	11	1	12	91.7	8.3	100.0
Av. (5 animals)		9.6	0.8	10.8	88.7	7.3	96.0
Injected samples							
VI	172	9	0	9	100.0	0	100.0
VII	197	9	3	12	75.0	25.0	100.0
VIII	219	10	0	10	100.0	0	100.0
Av. (3 animals)		9.3	1.0	10.3	91.7	8.3	100.0
IX	244	8	1	9	88.9	11.1	100.0
X	269	11	1	12	91.7	8.3	100.0
Av. (5 animals)		9.4	1.0	10.4	91.1	8.9	100.0

(continued)

Table 2 (continued)

Animal no.	Sample no.	No. slices obtained		Possible no. slices	Slices obtained		Total
		Firm	Crumbly		Firm %	Crumbly %	
SEMITENDINOSUS MUSCLE (continued)							
<u>Lactic acid injection</u>							
Control samples							
VI	173	6	0	10	60.0	0	60.0
VII	195	11	0	11	100.0	0	100.0
VIII	220	10	0	10	100.0	0	100.0
Av. (3 animals)		9.0	0	10.3	86.7	0	86.7
IX	245	8	1	10	80.0	10.0	90.0
X	267	12	0	12	100.0	0	100.0
Av. (5 animals)		9.4	0.2	10.6	88.0	2.0	90.0
Injected samples							
VI	173	6	0	11	54.5	0	54.5
VII	195	9	2	11	81.8	18.2	100.0
VIII	220	10	0	10	100.0	0	100.0
Av. (3 animals)		8.3	0.7	10.7	78.8	6.1	84.8
IX	245	7	3	10	70.0	30.0	100.0
X	267	12	0	12	100.0	0	100.0
Av. (5 animals)		8.8	1.0	10.8	81.3	9.6	90.9

(continued)

Table 2 (continued)

Animal no.	Sample no.	No. slices obtained		Possible no. slices	Slices obtained		
		Firm	Crumbly		Firm %	Crumbly %	Total %
SEMITENDINOSUS MUSCLE (continued)							
<u>Sodium chloride and lactic acid injection</u>							
Control samples							
VI	171	9	2	11	81.8	18.2	100.0
VII	196	11	0	11	100.0	0	100.0
VIII	221	10	0	10	100.0	0	100.0
Av. (3 animals)		10.0	0.7	10.7	93.9	6.1	100.0
IX	243	9	1	10	90.0	10.0	100.0
X	268	12	0	12	100.0	0	100.0
Av. (5 animals)		10.2	0.6	10.8	94.4	5.6	100.0
Injected samples							
VI	171	9	1	10	90.0	10.0	100.0
VII	196	10	0	10	100.0	0	100.0
VIII	221	10	0	10	100.0	0	100.0
Av. (3 animals)		9.7	0.3	10.0	96.7	3.3	100.0
IX	243	6	2	9	66.7	22.2	88.9
X	268	11	1	12	91.7	8.3	100.0
Av. (5 animals)		9.2	0.8	10.2	89.7	8.1	97.8

(continued)

Table 2 (continued)

Animal no.	Sample no.	No. slices obtained		Possible no. slices	Slices obtained		
		Firm	Crumbly		Firm %	Crumbly %	Total %
SEMI MEMBRANOSUS MUSCLE							
<u>Sodium chloride injection</u>							
Control samples							
VI	177	2	5	11	18.2	45.4	63.6
	179	0	5	10	0	50.0	50.0
Av.		1.0	5.0	10.5	9.1	47.7	56.8
VII	199	0	2	12	0	16.7	16.7
	200	4	5	11	36.4	45.4	81.8
Av.		2.0	3.5	11.5	18.2	31.0	49.2
VIII	222	0	7	11	0	63.6	63.6
	226	2	5	11	18.2	45.4	63.6
Av.		1.0	6.0	11.0	9.1	54.5	63.6
Av. (3 animals)		1.3	4.8	11.0	12.1	44.4	56.5
IX	249	0	3	11	0	27.3	27.3
	251	2	4	11	18.2	36.4	54.6
Av.		1.0	3.5	11.0	9.1	31.8	41.0
X	271	2	7	12	16.7	58.3	75.0
	272	7	4	12	58.3	33.3	91.6
Av.		4.5	5.5	12.0	37.5	45.8	83.3
Av. (5 animals)		1.9	4.7	11.2	16.6	42.2	58.8

(continued)

Table 2 (continued)

Animal no.	Sample no.	No. slices obtained		Possible no.	Slices obtained		Total
		Firm	Crumbly		Firm %	Crumbly %	

SEMIMEMBRANOSUS MUSCLE (continued)

Sodium chloride injection (continued)

Injected samples

VI	177	6	4	11	54.5	36.4	90.9
	179	0	6	11	0	54.5	54.5
Av.		3.0	5.0	11.0	27.2	45.4	72.7
VII	199	4	5	12	33.3	41.7	75.0
	200	2	7	10	20.0	70.0	90.0
Av.		3.0	6.0	11.0	26.6	55.8	82.5
VIII	222	0	8	12	0	66.7	66.7
	226	0	3	11	0	27.3	27.3
Av.		0	5.5	11.5	0	47.0	47.0
Av. (3 animals)		2.0	5.5	11.2	17.9	49.4	67.4
IX	249	0	0	11	0	0	0
	251	0	2	11	0	18.2	18.2
Av.		0	1.0	11.0	0	9.1	9.1
X	271	5	4	12	41.7	33.3	75.0
	272	0	8	11	0	72.7	72.7
Av.		2.5	6.0	11.5	20.8	53.0	73.8
Av. (5 animals)		1.7	4.7	11.2	14.9	42.1	57.0

(continued)

Table 2 (continued)

Animal no.	Sample no.	No. slices obtained		Possible no. slices	Slices obtained		
		Firm	Crumbly		Firm %	Crumbly %	Total %
SEMIMEMBRANOSUS MUSCLE (continued)							
<u>Lactic acid injection</u>							
Control samples							
VI	175	1	3	10	10.0	30.0	40.0
	176	8	3	12	66.7	25.0	91.7
Av.		4.5	3.0	11.0	38.4	27.5	65.8
VII	198	0	3	11	0	27.3	27.3
	202	1	6	11	9.1	54.5	63.6
Av.		0.5	4.5	11.0	4.6	40.9	45.4
VIII	225	0	4	11	0	36.4	36.4
	227	0	1	12	0	8.3	8.3
Av.		0	2.5	11.5	0	22.4	22.4
Av. (3 animals)		1.7	3.3	11.2	14.3	30.3	44.5
IX	247	0	4	11	0	36.4	36.4
	248	0	1	11	0	9.1	9.1
Av.		0	2.5	11.0	0	22.8	22.8
X	270	2	8	11	18.2	72.7	90.9
	274	6	2	11	54.5	18.2	72.7
Av.		4.0	5.0	11.0	36.4	45.4	81.8
Av. (5 animals)		1.8	3.5	11.1	15.9	31.8	47.6

(continued)

Table 2 (continued)

Animal no.	Sample no.	No. slices obtained		Possible no. slices	Slices obtained		Total
		Firm	Crumbly		Firm %	Crumbly %	
SEMIMEMBRANOSUS MUSCLE (continued)							
<u>Lactic acid injection (continued)</u>							
Injected samples							
VI	175	2	6	11	18.2	54.5	72.7
	176	3	5	11	27.3	45.4	72.7
Av.		2.5	5.5	11.0	22.8	50.0	72.7
VII	198	0	4	10	0	40.0	40.0
	202	10	1	12	83.3	8.3	91.6
Av.		5.0	2.5	11.0	41.6	24.2	65.8
VIII	225	5	3	12	41.7	25.0	66.7
	227	2	5	12	16.7	41.7	58.4
Av.		3.5	4.0	12.0	29.2	33.4	62.6
Av. (3 animals)		3.7	4.0	11.3	31.2	35.9	67.0
IX	247	0	3	11	0	27.3	27.3
	248	0	0	11	0	0	0
Av.		0	1.5	11.0	0	13.6	13.6
X	270	0	9	12	0	75.0	75.0
	274	4	4	11	36.4	36.4	72.8
Av.		2.0	6.5	11.5	18.2	55.7	73.9
Av. (5 animals)		2.6	4.0	11.3	22.4	35.4	57.7

(continued)

Table 2 (continued)

Animal no.	Sample no.	No. slices obtained		Possible no. slices	Slices obtained		Total
		Firm	Crumbly		Firm	Crumbly	
					%	%	%
SEMIMEMBRANOSUS MUSCLE (continued)							
<u>Sodium chloride and lactic acid injection</u>							
Control samples							
VI	174	0	8	10	0	80.0	80.0
	178	5	5	12	41.7	41.7	83.4
Av.		2.5	6.5	11.0	20.8	60.8	81.7
VII	201	0	8	12	0	66.7	66.7
	203	2	7	12	16.7	58.3	75.0
Av.		1.0	7.5	12.0	8.4	62.5	70.8
VIII	223	0	7	12	0	58.3	58.3
	224	3	6	11	27.3	54.5	81.8
Av.		1.5	6.5	11.5	13.6	56.4	70.0
Av. (3 animals)		1.7	6.8	11.5	14.3	59.9	74.2
IX	246	0	2	11	0	18.2	18.2
	250	2	2	12	16.7	16.7	33.4
Av.		1.0	2.0	11.5	8.4	17.4	25.8
X	273	7	4	12	58.3	33.3	91.6
	275	0	6	12	0	50.0	50.0
Av.		3.5	5.0	12.0	29.2	41.6	70.8
Av. (5 animals)		1.9	5.5	11.6	16.1	47.7	63.8

(continued)

Table 2 (continued)

Animal no.	Sample no.	No. slices obtained		Possible no. slices	Slices obtained		Total
		Firm	Crumbly		Firm %	Crumbly %	
SEMIMEMBRANOSUS MUSCLE (continued)							
<u>Sodium chloride and lactic acid injection (continued)</u>							
Injected samples							
VI	174	7	2	11	63.6	18.2	81.8
	178	0	7	10	0	70.0	70.0
Av.		3.5	4.5	10.5	31.8	44.1	75.9
VII	201	1	8	10	10.0	80.0	90.0
	203	0	7	12	0	58.3	58.3
Av.		0.5	7.5	11.0	5.0	69.2	74.2
VIII	223	5	4	12	41.7	33.3	75.0
	224	1	5	11	9.1	45.4	54.5
Av.		3.0	4.5	11.5	25.4	39.4	64.8
Av. (3 animals)		2.3	5.5	11.0	20.7	50.9	71.6
IX	246	0	4	10	0	40.0	40.0
	250	3	4	11	27.3	36.4	63.7
Av.		1.5	4.0	10.5	13.6	38.2	51.8
X	273	3	5	11	27.3	45.4	72.7
	275	0	6	12	0	50.0	50.0
Av.		1.5	5.5	11.5	13.6	47.7	61.3
Av. (5 animals)		2.0	5.2	11.0	17.9	47.7	65.6

(continued)

Table 2 (continued)

Animal no.	Sample no.	No. slices obtained		Possible no. slices	Slices obtained		
		Firm	Crumbly		Firm %	Crumbly %	Total %
BICEPS FEMORIS MUSCLE							
<u>Sodium chloride injection</u>							
Control samples							
VI	184	3	8	12	25.0	66.7	91.7
	185	7	3	13	53.8	23.1	76.9
Av.		5.0	5.5	12.5	39.4	44.9	84.3
VII	205	8	2	11	72.7	18.2	90.9
	207	9	2	12	75.0	16.7	91.7
Av.		8.5	2.0	11.5	73.8	17.4	91.3
VIII	228	5	2	13	38.5	15.4	53.9
	230	6	4	11	54.5	36.4	90.9
Av.		5.5	3.0	12.0	46.5	25.9	72.4
Av. (3 animals)		6.3	3.5	12.0	53.2	29.4	82.7
IX	256	0	3	12	0	25.0	25.0
	257	6	5	11	54.5	45.4	99.9
Av.		3.0	4.0	11.5	27.2	35.2	62.4
X	277	10	1	12	83.3	8.3	91.6
	279	11	0	11	100.0	0	100.0
Av.		10.5	0.5	11.5	91.6	4.2	95.8
Av. (5 animals)		6.5	3.0	11.8	55.7	25.5	81.2

(continued)

Table 2 (continued)

Animal no.	Sample no.	No. slices obtained		Possible no. slices	Slices obtained		
		Firm	Crumbly		Firm %	Crumbly %	Total %

BICEPS FEMORIS MUSCLE (continued)

Sodium chloride injection (continued)

Injected samples

VI	184	9	2	11	81.8	18.2	100.0
	185	11	1	12	91.7	8.3	100.0
Av.		10.0	1.5	11.5	86.8	13.2	100.0
VII	205	10	2	12	83.3	16.7	100.0
	207	8	4	12	66.7	33.3	100.0
Av.		9.0	3.0	12.0	75.0	25.0	100.0
VIII	228	6	2	11	54.5	18.2	72.7
	230	4	6	11	36.4	54.5	90.9
Av.		5.0	4.0	11.0	45.4	36.4	81.8
Av. (3 animals)		8.0	2.8	11.5	69.1	24.9	93.9
IX	256	0	3	11	0	27.3	27.3
	257	6	6	12	50.0	50.0	100.0
Av.		3.0	4.5	11.5	25.0	38.6	63.6
X	277	6	5	11	54.5	45.4	99.9
	279	11	2	13	84.6	15.4	100.0
Av.		8.5	3.5	12.0	69.6	30.4	100.0
Av. (5 animals)		7.1	3.3	11.6	60.4	28.7	89.1

(continued)

Table 2 (continued)

Animal no.	Sample no.	No. slices obtained		Possible no. slices	Slices obtained		Total
		Firm	Crumbly		Firm %	Crumbly %	
BICEPS FEMORIS MUSCLE (continued)							
<u>Lactic acid injection</u>							
Control samples							
VI	181	11	0	11	100.0	0	100.0
	183	10	2	12	83.3	16.7	100.0
Av.		10.5	1.0	11.5	91.6	8.4	100.0
VII	204	4	3	13	30.8	23.1	53.9
	206	10	2	12	83.3	16.7	100.0
Av.		7.0	2.5	12.5	57.0	19.9	77.0
VIII	232	11	2	13	84.6	15.4	100.0
	233	0	9	12	0	75.0	75.0
Av.		5.5	5.5	12.5	42.3	45.2	87.5
Av. (3 animals)		7.7	3.0	12.2	63.6	24.5	88.2
IX	253	0	1	10	0	10.0	10.0
	255	0	3	11	0	27.3	27.3
Av.		0	2.0	10.5	0	18.6	18.6
X	276	2	4	12	16.7	33.3	50.0
	278	10	1	11	90.9	9.1	100.0
Av.		6.0	2.5	11.5	53.8	21.2	75.0
Av. (5 animals)		5.8	2.7	11.7	48.9	22.7	71.6

(continued)

Table 2 (continued)

Animal no.	Sample no.	No. slices obtained		Possible no. slices	Slices obtained		
		Firm	Crumbly		Firm %	Crumbly %	Total %
BICEPS FEMORIS MUSCLE (continued)							
<u>Lactic acid injection</u> (continued)							
Injected samples							
VI	181	11	0	11	100.0	0	100.0
	183	7	5	12	58.3	41.7	100.0
Av.		9.0	2.5	11.5	79.2	20.8	100.0
VII	204	3	3	12	25.0	25.0	50.0
	206	8	2	11	72.7	18.2	90.9
Av.		5.5	2.5	11.5	48.8	21.6	70.4
VIII	232	11	1	12	91.7	8.3	100.0
	233	8	2	11	72.7	18.2	90.9
Av.		9.5	1.5	11.5	82.2	13.2	95.4
Av. (3 animals)		8.0	2.2	11.5	70.1	18.5	88.6
IX	253	0	2	10	0	20.0	20.0
	255	0	4	11	0	36.4	36.4
Av.		0	3.0	10.5	0	28.2	28.2
X	276	5	5	12	41.7	41.7	83.4
	278	9	1	11	81.8	9.1	90.9
Av.		7.0	3.0	11.5	61.8	25.4	87.2
Av. (5 animals)		6.2	2.5	11.3	54.4	21.8	76.2

(continued)

Table 2 (continued)

Animal no.	Sample no.	No. slices obtained		Possible no. slices	Slices obtained		
		Firm	Crumbly		Firm %	Crumbly %	Total %
BICEPS FEMORIS MUSCLE (continued)							
<u>Sodium chloride and lactic acid injection</u>							
Control samples							
VI	180	3	3	9	33.3	33.3	66.6
	182	12	0	12	100.0	0	100.0
Av.		7.5	1.5	10.5	66.6	16.6	83.3
VII	208	4	5	12	33.3	41.7	75.0
	209	11	1	12	91.7	8.3	100.0
Av.		7.5	3.0	12.0	62.5	25.0	87.5
VIII	229	7	2	11	63.6	18.2	81.8
	231	5	6	12	41.7	50.0	91.7
Av.		6.0	4.0	11.5	52.6	34.1	86.8
Av. (3 animals)		7.0	2.8	11.3	60.6	25.2	85.9
IX	252	0	0	11	0	0	0
	254	0	9	11	0	81.8	81.8
Av.		0	4.5	11.0	0	40.9	40.9
X	280	9	1	11	81.8	9.1	90.9
	281	12	0	12	100.0	0	100.0
Av.		10.5	0.5	11.5	90.9	4.6	95.4
Av. (5 animals)		6.3	2.7	11.3	54.5	24.2	78.8

(continued)

Table 2 (continued)

Animal no.	Sample no.	No. slices obtained		Possible no. slices	Slices obtained		
		Firm	Crumbly		Firm %	Crumbly %	Total %
BICEPS FEMORIS MUSCLE (continued)							
<u>Sodium chloride and lactic acid injection (continued)</u>							
Injected samples							
VI	180	0	5	11	0	45.4	45.4
	182	11	0	11	100.0	0	100.0
Av.		5.5	2.5	11.0	50.0	22.7	72.7
VII	208	2	6	12	16.7	50.0	66.7
	209	12	0	12	100.0	0	100.0
Av.		7.0	3.0	12.0	58.4	25.0	83.4
VIII	229	5	6	11	45.4	54.5	99.9
	231	4	9	13	30.8	69.2	100.0
Av.		4.5	7.5	12.0	38.1	61.8	100.0
Av. (3 animals)		5.7	4.3	11.7	48.8	36.5	85.4
IX	252	0	4	11	0	36.4	36.4
	254	0	9	11	0	81.8	81.8
Av.		0	6.5	11.0	0	59.1	59.1
X	280	10	2	13	76.9	15.4	92.3
	281	10	1	11	90.9	9.1	100.0
Av.		10.0	1.5	12.0	83.9	12.2	96.2
Av. (5 animals)		5.4	4.2	11.6	46.1	36.2	82.3

Table 3. Slicing Quality of Canned Beef. Judges' ratings of sliceability (from sample used in scoring) and the sliceability calculated by difference between weight of the canned beef and the unsliceable portion. The range for sliceability scores is 10 for extremely good slicing quality to 1 for extremely poor quality.

Animal* no.	Sample no.	Slice- ability score	Sliceability on basis of wt.			
			Canned meat gm.	Unslice- able meat gm.	Sliceable meat (by difference) gm.	%
LONGISSIMUS DORSI MUSCLE, LOIN PORTION						
<u>Sodium chloride injection</u>						
Control samples						
VI	162	6.7	379	59	320	84.4
VII	188	6.3	392	230	162	41.3
VIII	211	7.7	364	145	219	60.2
Av. (3 animals)		6.9	378	145	234	62.0
IX	234	4.7	376	248	128	34.0
X	260	4.0	373	165	208	55.8
Av. (5 animals)		5.9	377	169	207	55.1
Injected samples						
VI	162	5.7	376	81	295	78.4
VII	188	2.7	408	240	168	41.2
VIII	211	2.7	362	276	86	23.8
Av. (3 animals)		3.7	382	199	183	47.8
IX	234	3.3	360	326	34	9.4
X	260	9.3	406	73	333	82.0
Av. (5 animals)		4.7	382	199	183	47.0

(continued)

*Animals VI, VII, and VIII were steers, carcass grade Commercial, processed 65 minutes; Animal IX was a steer, carcass grade Commercial, processed 90 minutes; Animal X was a cow, carcass grade Cutter, processed 90 minutes.

Table 3 (continued)

Animal no.	Sample no.	Slice-ability score	Sliceability on basis of wt.			
			Canned meat gm.	Unslice-able meat gm.	Sliceable meat gm.	(by difference) %
LONGISSIMUS DORSI MUSCLE, LOIN PORTION (continued)						
<u>Lactic acid injection</u>						
Control samples						
VI	164	9.3	391	26	365	93.4
VII	187	2.0	394	359	35	8.9
VIII	210	3.0	363	331	32	8.8
Av. (3 animals)		4.8	383	239	144	37.0
IX	236	4.0	378	279	99	26.2
X	259	2.3	378	334	44	11.6
Av. (5 animals)		4.1	381	266	115	29.8
Injected samples						
VI	164	7.7	380	52	328	86.3
VII	187	6.0	383	157	226	59.0
VIII	210	6.3	354	62	292	82.5
Av. (3 animals)		6.7	372	90	282	75.6
IX	236	3.0	365	254	111	30.4
X	259	5.7	364	134	230	63.2
Av. (5 animals)		5.7	369	132	237	64.3

(continued)

Table 3 (continued)

Animal no.	Sample no.	Slice-ability score	Sliceability on basis of wt.			
			Canned meat gm.	Unslice-able meat gm.	Sliceable meat (by difference) gm.	%

LONGISSIMUS DORSI MUSCLE, LOIN PORTION (continued)

Sodium chloride and lactic acid injection

Control samples

VI	163	5.3	387	126	261	67.4
VII	186	7.0	390	73	317	81.3
VIII	212	5.3	364	189	175	48.1
Av. (3 animals)		5.9	380	129	251	65.6
IX	235	2.3	366	332	34	9.3
X	258	8.0	372	76	296	79.6
Av. (5 animals)		5.6	376	159	217	57.1

Injected samples

VI	163	9.3	385	58	327	84.9
VII	186	3.7	384	240	144	37.5
VIII	212	7.0	353	70	283	80.2
Av. (3 animals)		6.7	374	123	251	67.5
IX	235	3.0	355	327	28	7.9
X	258	6.3	395	80	315	79.7
Av. (5 animals)		5.9	374	155	219	58.0

(continued)

Table 3 (continued)

Animal no.	Sample no.	Slice-ability score	Sliceability on basis of wt.			
			Canned meat gm.	Unslice-able meat gm.	Sliceable meat (by difference) gm.	%
LONGISSIMUS DORSI MUSCLE, RIB PORTION						
<u>Sodium chloride injection</u>						
Control samples						
VI	167	9.3	372	0	372	100.0
VII	189	8.0	391	85	306	78.3
VIII	214	7.0	378	49	329	87.0
Av. (3 animals)		8.1	380	45	336	88.4
IX	239	8.7	375	22	353	94.1
X	261	8.0	356	99	257	72.2
Av. (5 animals)		8.2	374	51	323	86.3
Injected samples						
VI	167	9.3	384	33	351	91.4
VII	189	6.7	384	173	211	54.9
VIII	214	8.0	357	66	291	81.5
Av. (3 animals)		8.0	375	91	284	75.9
IX	239	5.0	368	109	259	70.4
X	261	8.0	356	82	274	77.0
Av. (5 animals)		7.4	370	93	277	75.0

(continued)

Table 3 (continued)

Animal no.	Sample no.	Slice-ability score	Sliceability on basis of wt.			
			Ganned meat gm.	Unslice-able meat gm.	Sliceable meat gm.	(by difference) %

LONGISSIMUS DORSI MUSCLE, RIB PORTION (continued)

Lactic acid injection

Control samples

VI	165	6.3	368	81	287	78.0
VII	190	8.3	424	0	424	100.0
VIII	215	9.0	379	35	344	90.8
Av. (3 animals)		7.9	390	39	352	89.6
IX	237	8.3	364	45	319	87.6
X	262	8.7	383	58	325	84.8
Av. (5 animals)		8.1	384	44	340	88.2

Injected samples

VI	165	4.7	363	183	180	49.6
VII	190	7.3	390	0	390	100.0
VIII	215	8.7	359	0	359	100.0
Av. (3 animals)		6.9	371	61	310	83.2
IX	237	5.7	343	66	277	80.8
X	262	9.0	363	35	328	90.4
Av. (5 animals)		7.1	364	57	307	84.2

(continued)

Table 3 (continued)

Animal no.	Sample no.	Slice-ability score	Sliceability on basis of wt.			
			Canned meat gm.	Unslice-able meat gm.	Sliceable meat (by difference) gm.	%

LONGISSIMUS DORSI MUSCLE, RIB PORTION (continued)

Sodium chloride and lactic acid injection

Control samples

VI	166	7.0	378	145	233	61.6
VII	191	9.0	387	27	360	93.0
VIII	213	7.0	383	20	363	94.8
Av. (3 animals)		7.7	383	64	319	83.1
IX	238	3.3	374	266	108	28.9
X	263	3.7	378	320	58	15.3
Av. (5 animals)		6.0	380	156	224	58.7

Injected samples

VI	166	7.3	388	62	326	84.0
VII	191	6.3	394	36	358	90.9
VIII	213	6.3	359	26	333	92.8
Av. (3 animals)		6.6	380	41	339	89.2
IX	238	5.3	356	256	100	28.1
X	263	8.3	385	83	302	78.4
Av. (5 animals)		6.7	376	93	284	74.8

(continued)

Table 3 (continued)

Animal no.	Sample no.	Slice-ability score	Sliceability on basis of wt.			
			Canned meat gm.	Unslice-able meat gm.	Sliceable meat gm.	(by difference) %
PSOAS MAJOR AND PSOAS MINOR MUSCLES						
<u>Sodium chloride injection</u>						
Control samples						
VI	168	5.3	414	0	414	100.0
VII	193	7.0	425	149	276	64.9
VIII	218	9.0	379	51	328	86.5
Av. (3 animals)		7.1	406	67	339	83.8
IX	240	5.7	422	189	233	55.2
X	265	4.7	373	94	279	74.8
Av. (5 animals)		6.3	403	97	306	76.3
Injected samples						
VI	168	3.7	388	75	313	80.7
VII	193	4.7	423	289	134	31.7
VIII	218	6.3	359	226	133	37.0
Av. (3 animals)		4.9	390	197	193	49.8
IX	240	2.7	387	348	39	10.1
X	265	6.7	362	58	304	84.0
Av. (5 animals)		4.8	384	199	185	48.7

(continued)

Table 3 (continued)

Animal no.	Sample no.	Slice-ability score	Sliceability on basis of wt.			
			Canned meat gm.	Unslice-able meat gm.	Sliceable meat (by difference) gm.	%

PSOAS MAJOR AND PSOAS MINOR MUSCLES (continued)

Lactic acid injection

Control samples

VI	169	6.7	418	0	418	100.0
VII	194	5.7	436	147	289	66.3
VIII	216	5.0	413	207	206	49.9
Av. (3 animals)		5.8	422	118	304	72.1
IX	241	5.0	408	272	136	33.3
X	266	7.3	383	86	297	77.5
Av. (5 animals)		5.9	412	142	269	65.4

Injected samples

VI	169	6.7	379	50	329	86.8
VII	194	3.7	369	295	74	20.0
VIII	216	4.7	380	231	149	39.2
Av. (3 animals)		5.0	376	192	184	48.7
IX	241	4.7	385	284	101	26.2
X	266	6.0	353	55	298	84.4
Av. (5 animals)		5.2	373	183	190	51.3

(continued)

Table 3 (continued)

Animal no.	Sample no.	Slice-ability score	Sliceability on basis of wt.			
			Canned meat gm.	Unslice-able meat gm.	Sliceable meat (by difference) gm.	%

PSOAS MAJOR AND PSOAS MINOR MUSCLES (continued)

Sodium chloride and lactic acid injection

Control samples

VI	170	8.7	396	25	371	93.7
VII	192	5.3	415	260	155	37.3
VIII	217	5.3	403	129	274	68.0
Av. (3 animals)		6.4	405	138	267	66.3
IX	242	8.0	382	126	256	67.0
X	264	5.3	393	264	129	32.8
Av. (5 animals)		6.5	398	161	237	59.8

Injected samples

VI	170	8.0	391	44	347	88.7
VII	192	3.3	414	277	137	33.1
VIII	217	6.7	380	138	242	63.7
Av. (3 animals)		6.0	395	153	242	61.8
IX	242	4.0	373	282	91	24.4
X	264	7.0	363	133	230	63.4
Av. (5 animals)		5.8	384	175	209	54.7

(continued)

Table 3 (continued)

Animal no.	Sample no.	Slice-ability score	Sliceability on basis of wt.			
			Canned meat gm.	Unslice-able meat gm.	Sliceable meat gm.	(by difference) %

SEMITENDINOSUS MUSCLE

Sodium chloride injection

Control samples

VI	172	10.0	369	39	330	89.4
VII	197	9.3	405	0	405	100.0
VIII	219	10.0	371	0	371	100.0
Av. (3 animals)		9.8	382	13	369	96.5
IX	244	9.0	369	37	332	90.0
X	269	10.0	363	0	363	100.0
Av. (5 animals)		9.7	375	15	360	95.9

Injected samples

VI	172	10.0	358	0	358	100.0
VII	197	9.7	384	0	384	100.0
VIII	219	9.7	374	0	374	100.0
Av. (3 animals)		9.8	372	0	372	100.0
IX	244	9.7	360	0	360	100.0
X	269	10.0	377	0	377	100.0
Av. (5 animals)		9.8	371	0	371	100.0

(continued)

Table 3 (continued)

Animal no.	Sample no.	Slice-ability score	Sliceability on basis of wt.			
			Canned meat gm.	Unslice-able meat gm.	Sliceable meat (by difference) gm.	%

SEMITENDINOSUS MUSCLE (continued)

Lactic acid injection

Control samples

VI	173	10.0	363	121	242	66.7
VII	195	9.3	375	0	375	100.0
VIII	220	10.0	362	0	362	100.0
Av. (3 animals)		9.8	367	40	326	88.9
IX	245	8.7	377	38	339	89.9
X	267	10.0	362	0	362	100.0
Av. (5 animals)		9.6	368	32	336	91.3

Injected samples

VI	173	10.0	401	141	260	64.8
VII	195	10.0	372	0	372	100.0
VIII	220	9.7	360	0	360	100.0
Av. (3 animals)		9.9	378	47	331	88.3
IX	245	9.3	366	0	366	100.0
X	267	10.0	347	0	347	100.0
Av. (5 animals)		9.8	369	28	341	93.0

(continued)

Table 3 (continued)

Animal no.	Sample no.	Slice-ability score	Sliceability on basis of wt.			
			Canned meat gm.	Unslice-able meat gm.	Sliceable meat gm.	(by difference) %

SEMITENDINOSUS MUSCLE (continued)

Sodium chloride and lactic acid injection

Control samples

VI	171	9.7	372	0	372	100.0
VII	196	10.0	380	0	380	100.0
VIII	221	10.0	369	0	369	100.0
Av. (3 animals)		9.9	374	0	374	100.0
IX	243	8.7	364	0	364	100.0
X	268	10.0	365	0	365	100.0
Av. (5 animals)		9.7	370	0	370	100.0

Injected samples

VI	171	10.0	370	0	370	100.0
VII	196	9.7	365	0	365	100.0
VIII	221	10.0	355	0	355	100.0
Av. (3 animals)		9.9	363	0	363	100.0
IX	243	9.0	370	36	334	90.3
X	268	10.0	359	0	359	100.0
Av. (5 animals)		9.7	364	7	357	98.1

(continued)

Table 3 (continued)

Animal no.	Sample no.	Slice-ability source	Sliceability on basis of wt.			
			Canned meat gm.	Unslice-able meat gm.	Sliceable meat (by difference) gm.	%
SEMI MEMBRANOSUS MUSCLE						
<u>Sodium chloride injection</u>						
Control samples						
VI	177	4.7	363	119	244	67.2
	179	5.3	374	147	227	60.7
Av.		5.0	368	133	236	64.0
VII	199	4.7	355	283	72	20.3
	200	7.7	360	60	300	83.3
Av.		6.2	358	172	186	51.8
VIII	222	7.3	367	124	243	66.2
	226	7.7	355	121	234	65.9
Av.		7.5	361	122	238	66.0
Av. (3 animals)		6.2	362	142	220	60.6
IX	249	5.0	355	226	129	36.3
	251	7.3	367	166	201	54.8
Av.		6.2	361	196	165	45.6
X	271	8.0	327	64	263	80.4
	272	6.3	330	23	307	93.0
Av.		7.2	328	44	285	86.7
Av. (5 animals)		6.4	355	133	222	62.8

(continued)

Table 3 (continued)

Animal no.	Sample no.	Slice-ability score	Sliceability on basis of wt.			
			Canned meat gm.	Unslice-able meat gm.	Sliceable meat gm.	(by difference) %

SEMIMEMBRANOSUS MUSCLE (continued)

Sodium chloride injection (continued)

Injected samples

VI	177	8.0	345	36	309	89.6
	179	6.3	352	127	225	63.9
Av.		7.2	348	82	267	76.8
VII	199	8.7	346	78	268	77.4
	200	7.3	350	20	330	94.3
Av.		8.0	348	49	299	85.8
VIII	222	5.3	350	101	249	71.1
	226	5.3	330	174	156	47.3
Av.		5.3	340	138	202	59.2
Av. (3 animals)		6.8	345	90	256	73.9
IX	249	1.7	342	313	29	8.5
	251	3.3	345	261	84	24.3
Av.		2.5	344	287	56	16.4
X	271	7.3	344	75	269	78.2
	272	5.7	324	63	261	80.6
Av.		6.5	334	69	265	79.4
Av. (5 animals)		5.9	343	125	218	63.5

(continued)

Table 3 (continued)

Animal no.	Sample no.	Slice-ability scores	Sliceability on basis of wt.			
			Canned meat gm.	Unslice-able meat gm.	Sliceable meat (by difference) gm. %	
SEMIMEMBRANOSUS MUSCLE (continued)						
<u>Lactic acid injection</u>						
Control samples						
VI	175	6.7	374	201	173	46.2
	176	8.0	358	29	329	91.9
Av.		7.4	366	115	251	69.0
VII	198	4.7	364	228	136	37.4
	202	6.0	362	106	256	70.7
Av.		5.4	363	167	196	54.0
VIII	225	4.3	345	221	124	35.9
	227	4.3	358	307	51	14.2
Av.		4.3	352	264	88	25.0
Av. (3 animals)		5.7	360	182	178	49.3
IX	247	6.0	348	206	142	40.8
	248	3.3	359	301	58	16.2
Av.		4.6	354	254	100	28.5
X	270	7.0	318	31	287	90.2
	274	9.0	328	65	263	80.2
Av.		8.0	323	48	275	85.2
Av. (5 animals)		5.9	352	170	182	52.3

(continued)

Table 3 (continued)

Animal no.	Sample no.	Slice-ability score	Sliceability on basis of wt.			
			Canned meat gm.	Unslice-able meat gm.	Sliceable meat gm.	(by difference) %

SEMIMEMBRANOSUS MUSCLE (continued)

Lactic acid injection (continued)

Injected samples

VI	175	7.3	376	85	291	77.4
	176	7.3	350	68	282	80.6
Av.		7.3	363	76	286	79.0
VII	198	4.7	342	164	178	52.0
	202	9.3	358	30	328	91.6
Av.		7.0	350	97	253	71.8
VIII	225	6.7	333	81	252	75.7
	227	4.7	351	134	217	61.8
Av.		5.7	342	108	234	68.8
Av. (3 animals)		6.7	352	94	258	73.2
IX	247	3.7	343	229	114	33.2
	248	3.3	342	316	26	7.6
Av.		3.5	342	272	70	20.4
X	270	6.7	302	70	232	76.8
	274	5.0	316	73	243	76.9
Av.		5.8	309	72	238	76.8
Av. (5 animals)		5.9	341	125	216	63.4

(continued)

Table 3 (continued)

Animal no.	Sample no.	Slice-ability score	Sliceability on basis of wt.			
			Canned meat gm.	Unslice-able meat gm.	Sliceable meat gm.	(by difference) %

SEMIMEMBRANOSUS MUSCLE (continued)

Sodium chloride and lactic acid injection

Control samples

VI	174	6.7	368	77	291	79.1
	178	7.3	363	54	309	85.1
Av.		7.0	366	66	300	82.1
VII	201	4.3	363	104	259	71.3
	203	7.0	368	71	297	80.7
Av.		5.6	366	88	278	76.0
VIII	223	5.7	352	124	228	64.8
	224	7.3	347	63	284	81.8
Av.		6.5	350	94	256	73.3
Av. (3 animals)		6.4	361	83	278	77.1
IX	246	2.0	364	275	89	24.4
	250	7.3	355	214	141	39.7
Av.		4.6	360	244	115	32.0
X	273	9.3	332	20	312	94.0
	275	6.7	336	117	219	65.2
Av.		8.0	334	68	266	79.6
Av. (5 animals)		6.3	355	112	243	68.6

(continued)

Table 3 (continued)

Animal no.	Sample no.	Slice-ability score	Sliceability on basis of wt.			
			Canned meat gm.	Unslice-able meat gm.	Sliceable meat gm.	(by difference) %

SEMIMEMBRANOSUS MUSCLE (continued)

Sodium chloride and lactic acid injection (continued)

Injected samples

VI	174	8.7	366	51	315	86.1
	178	6.3	344	97	247	71.8
Av.		7.5	355	74	281	79.0
VII	201	7.7	352	30	322	91.5
	203	5.3	375	110	265	70.7
Av.		6.5	364	70	294	81.1
VIII	223	7.3	345	70	275	79.7
	224	7.0	336	138	198	58.9
Av.		7.2	340	104	236	69.3
Av. (3 animals)		7.1	353	83	270	76.5
IX	246	6.7	353	175	178	50.4
	250	9.0	346	93	253	73.1
Av.		7.8	350	134	216	61.8
X	273	7.3	326	66	260	79.8
	275	7.0	333	128	205	61.6
Av.		7.2	330	97	232	70.7
Av. (5 animals)		7.2	348	96	252	72.4

(continued)

Table 3 (continued)

Animal no.	Sample no.	Slice-ability score	Sliceability on basis of wt.			
			Canned meat gm.	Unslice-able meat gm.	Sliceable meat (by difference)	
			gm.	gm.	gm.	%
BICEPS FEMORIS MUSCLE						
<u>Sodium chloride injection</u>						
Control samples						
VI	184	9.7	401	20	381	95.0
	185	9.0	394	68	326	82.7
Av.		9.4	398	44	354	88.8
VII	205	9.0	392	35	357	91.1
	207	8.7	373	28	345	92.5
Av.		8.8	382	32	351	91.8
VIII	228	4.0	383	118	265	69.2
	230	8.3	344	22	322	93.6
Av.		6.2	364	70	294	81.4
Av. (3 animals)		8.1	381	49	333	87.3
IX	256	6.3	362	200	162	44.8
	257	9.3	359	0	359	100.0
Av.		7.8	360	100	260	72.4
X	277	8.7	356	26	330	92.7
	279	10.0	345	0	345	100.0
Av.		9.4	350	13	338	96.4
Av. (5 animals)		8.3	371	52	319	86.2

(continued)

Table 3 (continued)

Animal no.	Sample no.	Slice-ability score	Sliceability on basis of wt.			
			Canned meat gm.	Unslice-able meat gm.	Sliceable meat gm.	(by difference) %

BICEPS FEMORIS MUSCLE (continued)

Sodium chloride injection (continued)

Injected samples

VI	184	9.0	360	0	360	100.0
	185	10.0	373	0	373	100.0
Av.		9.5	366	0	366	100.0
VII	205	9.3	367	0	367	100.0
	207	9.0	350	0	350	100.0
Av.		9.2	358	0	358	100.0
VIII	228	9.3	347	64	283	81.6
	230	7.3	342	30	312	91.2
Av.		8.3	344	47	298	86.4
Av. (3 animals)		9.0	356	16	341	95.5
IX	256	4.7	366	235	131	35.8
	257	9.3	359	0	359	100.0
Av.		7.0	362	118	245	67.9
X	277	8.7	360	0	360	100.0
	279	10.0	337	0	337	100.0
Av.		9.4	348	0	348	100.0
Av. (5 animals)		8.7	356	33	323	90.9

(continued)

Table 3 (continued)

Animal no.	Sample no.	Slice-ability score	Sliceability on basis of wt.			
			Canned meat gm.	Unslice-able meat gm.	Sliceable meat gm.	(by difference) %
BICEPS FEMORIS MUSCLE (continued)						
<u>Lactic acid injection</u>						
Control samples						
VI	181	9.7	372	0	372	100.0
	183	10.0	364	0	364	100.0
Av.		9.8	368	0	368	100.0
VII	204	5.7	391	138	253	64.7
	206	7.7	379	0	379	100.0
Av.		6.7	385	69	316	82.4
VIII	232	9.7	354	0	354	100.0
	233	7.3	357	74	283	79.3
Av.		8.5	356	37	318	89.6
Av. (3 animals)		8.3	370	35	334	90.7
IX	253	3.7	367	316	51	13.9
	255	5.3	364	205	159	43.7
Av.		4.5	366	260	105	28.8
X	276	5.7	375	156	219	58.4
	278	9.3	344	0	344	100.0
Av.		7.5	360	78	282	79.2
Av. (5 animals)		7.4	367	89	278	76.0

(continued)

Table 3 (continued)

Animal no.	Sample no.	Slice-ability score	Sliceability on basis of wt.			
			Canned meat gm.	Unslice-able meat gm.	Sliceable meat (by difference) gm.	%
BICEPS FEMORIS MUSCLE (continued)						
<u>Lactic acid injection (continued)</u>						
Injected samples						
VI	181	9.7	343	0	343	100.0
	183	8.7	340	0	340	100.0
Av.		9.2	342	0	342	100.0
VII	204	7.3	370	197	173	46.8
	206	9.0	354	37	317	89.5
Av.		8.2	362	117	245	68.2
VIII	232	9.0	348	0	348	100.0
	233	8.3	350	23	327	93.4
Av.		8.6	349	12	338	96.7
Av. (3 animals)		8.7	351	43	308	88.3
IX	253	4.7	345	243	102	29.6
	255	6.7	337	185	152	45.1
Av.		5.7	341	214	127	37.4
X	276	7.7	348	54	294	84.5
	278	10.0	321	22	299	93.1
Av.		8.8	334	38	296	88.8
Av. (5 animals)		8.1	346	76	270	78.2

(continued)

Table 3 (continued)

Animal no.	Sample no.	Slice-ability score	Sliceability on basis of wt.			
			Canned meat gm.	Unslice-able meat gm.	Sliceable meat gm.	(by difference) %

BICEPS FEMORIS MUSCLE (continued)

Sodium chloride and lactic acid injection

Control samples

VI	180	8.0	417	105	312	74.8
	182	10.0	362	0	362	100.0
Av.		9.0	390	52	337	87.4
VII	208	6.7	382	89	293	76.7
	209	9.7	385	0	385	100.0
Av.		8.2	384	45	339	88.4
VIII	229	9.3	358	66	292	81.6
	231	9.0	370	27	343	92.7
Av.		9.2	364	46	318	87.2
Av. (3 animals)		8.8	379	48	331	87.7
IX	252	3.0	376	336	40	10.6
	254	7.3	358	55	303	84.6
Av.		5.2	367	196	172	47.6
X	280	10.0	355	36	319	89.8
	281	9.7	365	0	365	100.0
Av.		9.8	360	18	342	94.9
Av. (5 animals)		8.3	373	71	302	81.1

(continued)

Table 3 (continued)

Animal no.	Sample no.	Slice-ability score	Sliceability on basis of wt.			
			Canned meat gm.	Unslice-able meat gm.	Sliceable meat gm.	(by difference) %
BICEPS FEMORIS MUSCLE (continued)						
<u>Sodium chloride and lactic acid injection (continued)</u>						
Injected samples						
VI	180	7.0	403	198	205	50.9
	182	9.3	368	0	368	100.0
Av.		8.2	386	99	286	75.4
VII	208	5.0	382	110	272	71.2
	209	9.7	378	0	378	100.0
Av.		7.4	380	55	325	85.6
VIII	229	9.3	354	0	354	100.0
	231	7.7	349	0	349	100.0
Av.		8.5	352	0	352	100.0
Av. (3 animals)		8.0	373	51	321	87.0
IX	252	7.0	363	202	161	44.4
	254	8.3	363	65	298	82.1
Av.		7.6	363	134	230	63.2
X	280	8.0	369	25	344	93.2
	281	9.7	364	0	364	100.0
Av.		8.8	366	12	354	96.6
Av. (5 animals)		8.1	369	60	309	84.2

Table 4. pH of Beef. pH of beef muscles aged one day and before dividing into cuts. Anterior, middle, and posterior positions are represented for muscles of trunk; proximal, middle, and distal positions for muscles of leg. (Left muscles were used for control cuts; right muscles for injected cuts.)

Animal no.	Position in animal	Position in muscle			Average pH
		Anterior (Proximal) pH	Middle pH	Posterior (Distal) pH	
LONGISSIMUS DORSI MUSCLE, LOIN PORTION					
VI	Left	5.35	5.35	5.35	5.35
	Right	5.35	5.35	5.40	5.37
VII	Left	5.40	5.45	5.40	5.42
	Right	5.45	5.40	5.40	5.42
VIII	Left	5.40	5.45	5.40	5.42
	Right	5.50	5.55	5.40	5.48
IX	Left	5.45	5.45	5.35	5.42
	Right	5.35	5.45	5.35	5.38
X	Left	5.70	5.80	5.60	5.70
	Right	5.70	5.75	5.55	5.67
LONGISSIMUS DORSI MUSCLE, RIB PORTION					
VI	Left	5.40	5.40	5.40	5.40
	Right	5.45	5.40	5.40	5.42
VII	Left	5.50	5.45	5.45	5.47
	Right	5.55	5.50	5.45	5.50
VIII	Left	5.45	5.40	5.45	5.43
	Right	5.45	5.45	5.45	5.45
IX	Left	5.45	5.40	5.45	5.43
	Right	5.45	5.35	5.35	5.38
X	Left	5.65	5.60	5.40	5.55
	Right	5.65	5.60	5.40	5.55

(continued)

Table 4 (continued)

Animal no.	Position in animal	Position in muscle			Average
		Anterior (Proximal)	Middle	Posterior (Distal)	
		pH	pH	pH	pH
PSOAS MAJOR AND PSOAS MINOR MUSCLES					
VI	Left	5.35	5.40	5.50	5.42
	Right	5.35	5.40	5.50	5.42
VII	Left	5.35	5.40	5.50	5.42
	Right	5.40	5.45	5.50	5.45
VIII	Left	5.40	5.40	5.50	5.43
	Right	5.40	5.40	5.50	5.43
IX	Left	5.35	5.40	5.55	5.43
	Right	5.40	5.40	5.55	5.45
X	Left	5.65	5.70	5.65	5.67
	Right	5.55	5.65	5.65	5.62
SEMITENDINOSUS MUSCLE					
VI	Left	5.35	5.35	5.40	5.37
	Right	5.35	5.35	5.45	5.38
VII	Left	5.35	5.35	5.40	5.37
	Right	5.40	5.40	5.50	5.43
VIII	Left	5.35	5.40	5.45	5.40
	Right	5.35	5.40	5.40	5.38
IX	Left	5.30	5.30	5.35	5.32
	Right	5.30	5.30	5.40	5.33
X	Left	5.50	5.50	5.60	5.53
	Right	5.50	5.50	5.60	5.53

(continued)

Table 4 (continued)

Animal no.	Position in animal	Position in muscle			Average pH
		Anterior (Proximal)	Middle	Posterior (Distal)	
		pH	pH	pH	pH
SEMIMEMBRANOSUS MUSCLE					
VI	Left	5.30	5.30	5.30	5.30
	Right	5.30	5.35	5.30	5.32
VII	Left	5.35	5.30	5.35	5.33
	Right	5.35	5.30	5.35	5.33
VIII	Left	5.40	5.30	5.35	5.35
	Right	5.40	5.30	5.35	5.35
IX	Left	5.40	5.30	5.30	5.33
	Right	5.50	5.30	5.30	5.37
X	Left	5.55	5.50	5.55	5.53
	Right	5.55	5.50	5.60	5.55
BICEPS FEMORIS MUSCLE					
VI	Left	5.45	5.40	5.40	5.42
	Right	5.45	5.35	5.45	5.42
VII	Left	5.50	5.55	5.50	5.52
	Right	5.55	5.55	5.50	5.53
VIII	Left	5.40	5.35	5.40	5.38
	Right	5.40	5.35	5.45	5.40
IX	Left	5.40	5.35	5.35	5.37
	Right	5.45	5.40	5.35	5.40
X	Left	5.75	5.60	5.65	5.67
	Right	5.75	5.60	5.65	5.67

Table 5. pH of Beef. pH of beef muscles (average of three determinations) before dividing into cuts but after aging one day; of beef cuts aged eight days (one determination); and of the canned beef (one determination).

Animal* no.	Beef muscles (uninjected) aged one day pH	Sample no.	Beef cuts aged eight days pH	Canned beef pH
LONGISSIMUS DORSI MUSCLE, LOIN PORTION				
<u>Sodium chloride injection</u>				
Left muscle		Control samples		
VI	5.35	162	5.45	5.75
VII	5.42	188	5.40	5.60
VIII	5.42	211	5.35	5.75
Av. (3 animals)	5.40		5.40	5.70
IX	5.42	234	5.30	5.65
X	5.70	260	5.50	5.90
Av. (5 animals)	5.46		5.40	5.73
Right muscle		Injected samples		
VI	5.37	162	5.35	5.70
VII	5.42	188	5.40	5.60
VIII	5.48	211	5.35	5.70
Av. (3 animals)	5.42		5.37	5.65
IX	5.38	234	5.40	5.65
X	5.67	260	5.55	5.85
Av. (5 animals)	5.46		5.41	5.70

(continued)

*Animals VI, VII, and VIII were steers, carcass grade Commercial, processed 65 minutes; Animal IX was a steer, carcass grade Commercial, processed 90 minutes; Animal X was a cow, carcass grade Cutter, processed 90 minutes.

Table 5 (continued)

Animal no.	Beef muscles (uninjected) aged one day pH	Sample no.	Beef cuts aged eight days pH	Canned beef pH
LONGISSIMUS DORSI MUSCLE, LOIN PORTION (continued)				
<u>Lactic acid injection</u>				
Left muscle	(same as for sodium chloride injection)	Control samples		
VI		164	5.30	5.70
VII		187	5.45	5.60
VIII		210	5.30	5.70
Av. (3 animals)			5.35	5.67
IX		236	5.30	5.60
X		259	5.50	5.90
Av. (5 animals)			5.37	5.70
Right muscle	(same as for sodium chloride injection)	Injected samples		
VI		164	5.40	5.70
VII		187	5.45	5.60
VIII		210	5.30	5.70
Av. (3 animals)			5.38	5.67
IX		236	5.30	5.60
X		259	5.50	5.90
Av. (5 animals)			5.39	5.70

(continued)

Table 5 (continued)

Animal no.	Beef muscles (uninjected) aged one day pH	Sample no.	Beef cuts aged eight days pH	Canned beef pH
LONGISSIMUS DORSI MUSCLE, LOIN PORTION (continued)				
<u>Sodium chloride and lactic acid injection</u>				
Left muscle	(same as for sodium chloride injection)	Control samples		
VI		163	5.40	5.70
VII		186	5.40	5.60
VIII		212	5.30	5.65
Av. (3 animals)			5.37	5.65
IX		235	5.35	5.60
X		258	5.55	5.85
Av. (5 animals)			5.40	5.68
Right muscle	(same as for sodium chloride injection)	Injected samples		
VI		163	5.40	5.70
VII		186	5.45	5.55
VIII		212	5.35	5.65
Av. (3 animals)			5.40	5.63
IX		235	5.40	5.60
X		258	5.55	5.85
Av. (5 animals)			5.43	5.67

(continued)

Table 5 (continued)

Animal no.	Beef muscles (uninjected) aged one day pH	Sample no.	Beef cuts aged eight days pH	Canned beef pH
LONGISSIMUS DORSI MUSCLE, RIB PORTION				
<u>Sodium chloride injection</u>				
Left muscle		Control samples		
VI	5.40	167	5.50	5.80
VII	5.47	189	5.55	5.80
VIII	5.43	214	5.40	5.75
Av. (3 animals)	5.43		5.48	5.78
IX	5.43	239	5.35	5.65
X	5.55	261	5.55	6.00
Av. (5 animals)	5.46		5.47	5.80
Right muscle		Injected samples		
VI	5.42	167	5.50	5.75
VII	5.50	189	5.65	5.75
VIII	5.45	214	5.50	5.80
Av. (3 animals)	5.46		5.55	5.77
IX	5.38	239	5.40	5.65
X	5.55	261	5.60	5.90
Av. (5 animals)	5.46		5.53	5.77

(continued)

Table 5 (continued)

Animal no.	Beef muscles (uninjected) aged one day pH	Sample no.	Beef cuts aged eight days pH	Canned beef pH
LONGISSIMUS DORSI MUSCLE, RIB PORTION (continued)				
<u>Lactic acid injection</u>				
Left muscle	(same as for sodium chloride injection)	Control samples		
VI		165	5.45	5.85
VII		190	5.55	5.75
VIII		215	5.45	5.75
Av. (3 animals)			5.48	5.78
IX		237	5.40	5.75
X		262	5.50	5.90
Av. (5 animals)			5.47	5.80
Right muscle	(same as for sodium chloride injection)	Injected samples		
VI		165	5.50	6.00
VII		190	5.60	5.75
VIII		215	5.45	5.75
Av. (3 animals)			5.52	5.83
IX		237	5.40	5.75
X		262	5.60	5.95
Av. (5 animals)			5.51	5.84

(continued)

Table 5 (continued)

Animal no.	Beef muscles (uninjected) aged one day pH	Sample no.	Beef cuts aged eight days pH	Canned beef pH
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LONGISSIMUS DORSI MUSCLE, RIB PORTION (continued)

Sodium chloride and lactic acid injection

Left muscle	(same as for sodium chloride injection)	Control samples		
VI		166	5.45	5.80
VII		191	5.50	5.70
VIII		213	5.40	5.85
Av. (3 animals)			5.45	5.78
IX		238	5.35	5.65
X		263	5.50	5.90
Av. (5 animals)			5.44	5.78
Right muscle	(same as for sodium chloride injection)	Injected samples		
VI		166	5.60	5.80
VII		191	5.60	5.70
VIII		213	5.40	5.80
Av. (3 animals)			5.53	5.77
IX		238	5.35	5.65
X		263	5.65	5.90
Av. (5 animals)			5.52	5.77

(continued)

Table 5 (continued)

Animal no.	Beef muscles (uninjected) aged one day pH	Sample no.	Beef cuts aged eight days pH	Canned beef pH
PSOAS MAJOR AND PSOAS MINOR MUSCLES				
<u>Sodium chloride injection</u>				
Left muscle		Control samples		
VI	5.42	168	5.50	5.85
VII	5.42	193	5.40	5.65
VIII	5.43	218	5.55	5.85
Av. (3 animals)	5.42		5.48	5.78
IX	5.43	240	5.40	5.60
X	5.67	265	5.65	6.00
Av. (5 animals)	5.47		5.50	5.79
Right muscle		Injected samples		
VI	5.42	168	5.55	5.85
VII	5.45	193	5.50	5.65
VIII	5.43	218	5.50	5.80
Av. (3 animals)	5.43		5.52	5.77
IX	5.45	240	5.45	5.60
X	5.62	265	5.50	5.85
Av. (5 animals)	5.47		5.50	5.75

(continued)

Table 5 (continued)

Animal no.	Beef muscles (uninjected) aged one day pH	Sample no.	Beef cuts aged eight days pH	Canned beef pH
PSOAS MAJOR AND PSOAS MINOR MUSCLES (continued)				
<u>Lactic acid injection</u>				
Left muscle	(same as for sodium chloride injection)	Control samples		
VI		169	5.60	5.85
VII		194	5.55	5.75
VIII		216	5.45	5.80
Av. (3 animals)			5.53	5.80
IX		241	5.45	5.65
X		266	5.65	6.00
Av. (5 animals)			5.54	5.81
Right muscle	(same as for sodium chloride injection)	Injected samples		
VI		169	5.65	5.80
VII		194	5.60	5.80
VIII		216	5.40	5.75
Av. (3 animals)			5.55	5.78
IX		241	5.45	5.65
X		266	5.65	6.05
Av. (5 animals)			5.55	5.81

(continued)

Table 5 (continued)

Animal no.	Beef muscles (uninjected) aged one day	Sample no.	Beef cuts aged eight days	Canned beef
	pH		pH	pH
PSOAS MAJOR AND PSOAS MINOR MUSCLES (continued)				
<u>Sodium chloride and lactic acid injection</u>				
Left muscle	(same as for sodium chloride injection)	Control samples		
VI		170	5.60	5.85
VII		192	5.40	5.65
VIII		217	5.45	5.80
Av. (3 animals)			5.48	5.77
IX		242	5.55	5.75
X		264	5.60	6.00
Av. (5 animals)			5.52	5.81
Right muscle	(same as for sodium chloride injection)	Injected samples		
VI		170	5.60	5.85
VII		192	5.50	5.65
VIII		217	5.45	5.75
Av. (3 animals)			5.52	5.75
IX		242	5.55	5.75
X		264	5.60	5.90
Av. (5 animals)			5.54	5.78

(continued)

Table 5 (continued)

Animal no.	Beef muscles (uninjected) aged one day pH	Sample no.	Beef cuts aged eight days pH	Canned beef pH
SEMITENDINOSUS MUSCLE				
<u>Sodium chloride injection</u>				
Left muscle		Control samples		
VI	5.37	172	5.45	5.80
VII	5.37	197	5.45	5.75
VIII	5.40	219	5.35	5.75
Av. (3 animals)	5.38		5.42	5.77
IX	5.32	244	5.40	5.65
X	5.53	269	5.55	5.90
Av. (5 animals)	5.40		5.44	5.77
Right muscle		Injected samples		
VI	5.38	172	5.40	5.80
VII	5.43	197	5.50	5.70
VIII	5.38	219	5.35	5.75
Av. (3 animals)	5.40		5.42	5.75
IX	5.33	244	5.40	5.65
X	5.53	269	5.60	5.85
Av. (5 animals)	5.41		5.45	5.75

(continued)

Table 5 (continued)

Animal no.	Beef muscles (uninjected) aged one day pH	Sample no.	Beef cuts aged eight days pH	Canned beef pH
SEMITENDINOSUS MUSCLE (continued)				
<u>Lactic acid injection</u>				
Left muscle	(same as for sodium chloride injection)	Control samples		
VI		173	5.40	5.85
VII		195	5.50	5.65
VIII		220	5.40	5.75
Av. (3 animals)			5.43	5.75
IX		245	5.40	5.70
X		267	5.50	5.90
Av. (5 animals)			5.44	5.77
Right muscle	(same as for sodium chloride injection)	Injected samples		
VI		173	5.50	5.85
VII		195	5.40	5.65
VIII		220	5.50	5.75
Av. (3 animals)			5.47	5.75
IX		245	5.45	5.70
X		267	5.55	5.90
Av. (5 animals)			5.48	5.77

(continued)

Table 5 (continued)

Animal no.	Beef muscles (uninjected) aged one day pH	Sample no.	Beef cuts aged eight days pH	Canned beef pH
SEMITENDINOSUS MUSCLE (continued)				
<u>Sodium chloride and lactic acid injection</u>				
Left muscle	(same as for sodium chloride injection)	Control samples		
VI		171	5.45	5.80
VII		196	5.40	5.65
VIII		221	5.50	5.80
Av. (3 animals)			5.45	5.75
IX		243	5.40	5.65
X		268	5.50	5.95
Av. (5 animals)			5.45	5.77
Right muscle	(same as for sodium chloride injection)	Injected samples		
VI		171	5.40	5.75
VII		196	5.45	5.65
VIII		221	5.45	5.75
Av. (3 animals)			5.43	5.72
IX		243	5.40	5.65
X		268	5.60	5.85
Av. (5 animals)			5.46	5.73

(continued)

Table 5 (continued)

Animal no.	Beef muscles (uninjected) aged one day pH	Sample no.	Beef cuts aged eight days pH	Canned beef pH
SEMIMEMBRANOSUS MUSCLE				
<u>Sodium chloride injection</u>				
Left muscle		Control samples		
VI	5.30	177	5.35	5.80
		179	5.35	5.75
Av.			5.35	5.78
VII	5.33	199	5.45	5.60
		200	5.45	5.60
Av.			5.45	5.60
VIII	5.35	222	5.30	5.65
		226	5.35	5.70
Av.			5.32	5.68
Av. (3 animals)	5.33		5.37	5.69
IX	5.33	249	5.35	5.60
		251	5.40	5.60
Av.			5.38	5.60
X	5.53	271	5.45	5.85
		272	5.50	5.90
Av.			5.48	5.88
Av. (5 animals)	5.37		5.40	5.71

(continued)

Table 5 (continued)

Animal no.	Beef muscles (uninjected) aged one day pH	Sample no.	Beef cuts aged eight days pH	Canned beef pH
SEMIMEMBRANOSUS MUSCLE (continued)				
<u>Sodium chloride injection (continued)</u>				
Right muscle		Injected samples		
VI	5.32	177	5.40	5.75
		179	5.35	5.70
Av.			5.38	5.72
VII	5.33	199	5.50	5.60
		200	5.45	5.60
Av.			5.48	5.60
VIII	5.35	222	5.35	5.65
		226	5.40	5.65
Av.			5.38	5.65
Av. (3 animals)	5.33		5.41	5.66
IX	5.37	249	5.40	5.55
		251	5.40	5.55
Av.			5.40	5.55
X	5.55	271	5.50	5.80
		272	5.50	5.80
Av.			5.50	5.80
Av. (5 animals)	5.38		5.43	5.66

(continued)

Table 5 (continued)

Animal no.	Beef muscles (uninjected) aged one day pH	Sample no.	Beef cuts aged eight days pH	Canned beef pH
SEMIMEMBRANOSUS MUSCLE (continued)				
<u>Lactic acid injection</u>				
Left muscle	(same as for sodium chloride injections)	Control samples		
VI		175	5.35	5.75
		176	5.35	5.75
Av.			5.35	5.75
VII		198	5.45	5.60
		202	5.40	5.60
Av.			5.42	5.60
VIII		225	5.40	5.65
		227	5.35	5.65
Av.			5.38	5.65
Av. (3 animals)			5.38	5.67
IX		247	5.30	5.60
		248	5.40	5.55
Av.			5.35	5.58
X		270	5.45	5.80
		274	5.70	5.95
Av.			5.59	5.88
Av. (5 animals)			5.42	5.69

(continued)

Table 5 (continued)

Animal no.	Beef muscles (uninjected) aged one day pH	Sample no.	Beef cuts aged eight days pH	Canned beef pH
SEMIMEMBRANOSUS MUSCLE (continued)				
<u>Lactic acid injection</u> (continued)				
Right muscle	(same as for sodium chloride injection)	Injected samples		
VI		175	5.35	5.80
		176	5.35	5.75
Av.			5.35	5.78
VII		198	5.45	5.60
		202	5.40	5.60
Av.			5.42	5.60
VIII		225	5.40	5.70
		227	5.35	5.65
Av.			5.38	5.68
Av. (3 animals)			5.38	5.69
IX		247	5.30	5.60
		248	5.35	5.55
Av.			5.32	5.58
X		270	5.45	5.80
		274	5.70	6.05
Av.			5.58	5.92
Av. (5 animals)			5.41	5.71

(continued)

Table 5 (continued)

Animal no.	Beef muscles (uninjected) aged one day pH	Sample no.	Beef cuts aged eight days pH	Canned beef pH
SEMIMEMBRANOSUS MUSCLE (continued)				
<u>Sodium chloride and lactic acid injection</u>				
Left muscle	(same as for sodium chloride injection)	Control samples		
VI		174	5.35	5.70
		178	5.35	5.85
Av.			5.35	5.78
VII		201	5.45	5.60
		203	5.40	5.60
Av.			5.42	5.60
VIII		223	5.35	5.65
		224	5.35	5.60
Av.			5.35	5.62
Av. (3 animals)			5.37	5.67
IX		246	5.30	5.60
		250	5.35	5.55
Av.			5.32	5.58
X		273	5.50	5.85
		275	5.55	6.00
Av.			5.52	5.92
Av. (5 animals)			5.39	5.70

(continued)

Table 5 (continued)

Animal no.	Beef muscles (uninjected) aged one day pH	Sample no.	Beef cuts aged eight days pH	Canned beef pH
SEMIMEMBRANOSUS MUSCLE (continued)				
<u>Sodium chloride and lactic acid injection (continued)</u>				
Right muscle	(same as for sodium chloride injection)	Injected samples		
VI		174	5.35	5.70
		178	5.35	5.80
Av.			5.35	5.75
VII		201	5.40	5.60
		203	5.40	5.60
Av.			5.40	5.60
VIII		223	5.35	5.65
		224	5.40	5.60
Av.			5.38	5.62
Av. (3 animals)			5.38	5.66
IX		246	5.35	5.55
		250	5.40	5.60
Av.			5.38	5.58
X		273	5.55	5.85
		275	5.75	5.95
Av.			5.65	5.90
Av. (5 animals)			5.43	5.69

(continued)

Table 5 (continued)

Animal no.	Beef muscles (uninjected) aged one day pH	Sample no.	Beef cuts aged eight days pH	Canned beef pH
BICEPS FEMORIS MUSCLE				
<u>Sodium chloride injection</u>				
Left muscle		Control samples		
VI	5.42	184	5.40	5.80
		185	5.45	5.80
Av.			5.42	5.80
VII	5.52	205	5.45	5.70
		207	5.40	5.60
Av.			5.42	5.65
VIII	5.38	228	5.35	5.75
		230	5.30	5.65
Av.			5.32	5.70
Av. (3 animals)	5.44		5.39	5.72
IX	5.37	256	5.35	5.65
		257	5.45	5.70
Av.			5.40	5.68
X	5.67	277	5.65	5.95
		279	5.50	5.90
Av.			5.58	5.92
Av. (5 animals)	5.47		5.43	5.75

(continued)

Table 5 (continued)

Animal no.	Beef muscles (uninjected) aged one day pH	Sample no.	Beef cuts aged eight days pH	Canned beef pH
BICEPS FEMORIS MUSCLE (continued)				
<u>Sodium chloride injection (continued)</u>				
Right muscle		Injected samples		
VI	5.42	184	5.40	5.85
		185	5.45	5.80
Av.			5.42	5.82
VII	5.53	205	5.45	5.65
		207	5.40	5.60
Av.			5.42	5.62
VIII	5.40	228	5.40	5.70
		230	5.35	5.65
Av.			5.38	5.68
Av. (3 animals)	5.45		5.41	5.71
IX	5.40	256	5.40	5.60
		257	5.50	5.65
Av.			5.45	5.62
X	5.67	277	5.55	5.95
		279	5.50	5.80
Av.			5.52	5.88
Av. (5 animals)	5.48		5.44	5.72

(continued)

Table 5 (continued)

Animal no.	Beef muscles (uninjected) aged one day pH	Sample no.	Beef cuts aged eight days pH	Canned beef pH
BICEPS FEMORIS MUSCLE (continued)				
<u>Lactic acid injection</u>				
Left muscle	(same as for sodium chloride injection)	Control samples		
VI		181	5.50	5.85
		183	5.40	5.70
Av.			5.45	5.78
VII		204	5.45	5.70
		206	5.40	5.65
Av.			5.42	5.68
VIII		232	5.35	5.75
		233	5.45	5.75
Av.			5.40	5.75
Av. (3 animals)			5.42	5.74
IX		253	5.30	5.65
		255	5.40	5.60
Av.			5.35	5.62
X		276	5.60	6.00
		278	5.45	5.90
Av.			5.52	5.95
Av. (5 animals)			5.43	5.76

(continued)

Table 5 (continued)

Animal no.	Beef muscles (uninjected) aged one day pH	Sample no.	Beef cuts aged eight days pH	Canned beef pH
BICEPS FEMORIS MUSCLE (continued)				
<u>Lactic acid injection (continued)</u>				
Right muscle	(same as for sodium chloride injection)	Injected samples		
VI		181	5.40	5.90
		183	5.40	5.70
Av.			5.40	5.80
VII		204	5.45	5.70
		206	5.40	5.60
Av.			5.42	5.65
VIII		232	5.35	5.75
		233	5.50	5.75
Av.			5.42	5.75
Av. (3 animals)			5.41	5.73
IX		253	5.40	5.65
		255	5.35	5.60
Av.			5.38	5.62
X		276	5.50	6.00
		278	5.50	5.85
Av.			5.50	5.92
Av. (5 animals)			5.42	5.75

(continued)

Table 5 (continued)

Animal no.	Beef muscles (uninjected) aged one day pH	Sample no.	Beef cuts aged eight days pH	Canned beef pH
BICEPS FEMORIS MUSCLE (continued)				
<u>Sodium chloride and lactic acid injection</u>				
Left muscle	(same as for sodium chloride injection)	Control samples		
VI		180	5.50	5.85
		182	5.35	5.85
Av.			5.42	5.85
VII		208	5.40	5.60
		209	5.40	5.65
Av.			5.40	5.62
VIII		229	5.35	5.70
		231	5.30	5.65
Av.			5.32	5.68
Av. (3 animals)			5.38	5.72
IX		252	5.45	5.70
		254	5.30	5.65
Av.			5.38	5.68
X		280	5.55	5.95
		281	5.45	6.00
Av.			5.50	5.98
Av. (5 animals)			5.40	5.76

(continued)

Table 5 (continued)

Animal no.	Beef muscles (uninjected) aged one day pH	Sample no.	Beef cuts aged eight days pH	Canned beef pH
BICEPS FEMORIS MUSCLE (continued)				
<u>Sodium chloride and lactic acid injection (continued)</u>				
Right muscle	(same as for	Injected samples		
VI	sodium chloride injection	180	5.50	5.80
		182	5.40	5.80
Av.			5.45	5.80
VII		208	5.45	5.60
		209	5.45	5.65
Av.			5.45	5.62
VIII		229	5.45	5.70
		231	5.40	5.65
Av.			5.42	5.68
Av. (3 animals)			5.44	5.70
IX		252	5.45	5.65
		254	5.45	5.60
Av.			5.45	5.62
X		280	5.55	5.80
		281	5.50	5.85
Av.			5.52	5.82
Av. (5 animals)			5.46	5.71

(continued)

Table 6. Weight Changes of Beef before Canning. Initial weight of beef and weight of beef plus injecting solution one day after slaughter of animal; weight of aged beef eight days after slaughter; percentage weight after aging based on initial weight of sample.

Animal* no.	Sample no.	Initial wt. gm.	Beef plus injecting solution gm.	Wt. beef after aging gm.	%	
LONGISSIMUS DORSI MUSCLE, LOIN PORTION						
<u>Sodium chloride injection</u>						
Control samples						
	VI	162	606	598	98.7	
	VII	188	601	587	97.7	
	VIII	211	605	595	98.3	
	Av. (3 animals)		604	593	98.2	
	IX	234	605	587	97.0	
	X	260	604	593	98.2	
	Av. (5 animals)		604	592	98.0	
Injected samples						
	VI	162	608	668	631	103.8
	VII	188	602	662	617	102.5
	VIII	211	606	666	593	97.8
	Av. (3 animals)		605	665	614	101.4
	IX	234	606	666	635	104.8
	X	260	606	666	634	104.6
	Av. (5 animals)		606	666	622	102.7

(continued)

*Animals VI, VII, and VIII were steers, carcass grade Commercial, processed 65 minutes; Animal IX was a steer, carcass grade Commercial, processed 90 minutes; Animal X was a cow, carcass grade Cutter, processed 90 minutes.

Table 6 (continued)

Animal no.	Sample no.	Initial wt. gm.	Beef plus injecting solution gm.	Wt. beef after aging	
				gm.	%
LONGISSIMUS DORSI MUSCLE, LOIN PORTION (continued)					
<u>Lactic acid injection</u>					
Control samples					
VI	164	604		597	98.8
VII	187	604		588	97.4
VIII	210	606		597	98.5
Av. (3 animals)		605		594	98.2
IX	236	606		594	98.0
X	259	607		598	98.5
Av. (5 animals)		605		595	98.2
Injected samples					
VI	164	606	666	605	99.8
VII	187	608	668	599	98.5
VIII	210	606	666	602	99.3
Av. (3 animals)		607	667	602	99.2
IX	236	606	666	607	100.2
X	259	607	667	633	104.3
Av. (5 animals)		606	666	609	100.4

(continued)

Table 6 (continued)

Animal no.	Sample no.	Initial wt.	Beef plus injecting solution	Wt. beef after aging	
		gm.	gm.	gm.	%
LONGISSIMUS DORSI MUSCLE, LOIN PORTION (continued)					
<u>Sodium chloride and lactic acid injection</u>					
Control samples					
VI	163	605		590	97.5
VII	186	600		588	98.0
VIII	212	606		592	97.7
Av. (3 animals)		604		590	97.7
IX	235	600		591	98.5
X	258	601		587	97.7
Av. (5 animals)		602		590	97.9
Injected samples					
VI	163	608	668	630	103.6
VII	186	604	664	622	103.0
VIII	212	607	667	630	103.8
Av. (3 animals)		606	666	627	103.5
IX	235	605	665	619	102.3
X	258	604	664	634	105.0
Av. (5 animals)		606	666	627	103.5

(continued)

Table 6 (continued)

Animal no.	Sample no.	Initial wt.	Beef plus injecting solution	Wt. beef after aging		
		gm.	gm.	gm.	%	
LONGISSIMUS DORSI MUSCLE, RIB PORTION						
<u>Sodium chloride injection</u>						
Control samples						
	VI	167	605	596	98.5	
	VII	189	601	594	98.8	
	VIII	214	606	603	99.5	
	Av. (3 animals)		604	598	98.9	
	IX	239	608	597	98.2	
	X	261	608	603	99.2	
	Av. (5 animals)		606	599	98.8	
Injected samples						
	VI	167	600	660	640	106.7
	VII	189	608	668	661	108.7
	VIII	214	605	665	643	106.3
	Av. (3 animals)		604	664	648	107.2
	IX	239	605	665	646	106.8
	X	261	607	667	661	108.9
	Av. (5 animals)		605	665	650	107.5

(continued)

Table 6 (continued)

Animal no.	Sample no.	Initial wt. gm.	Beef plus injecting solution gm.	Wt. beef after aging		
				gm.	%	
LONGISSIMUS DORSI MUSCLE, RIB PORTION (continued)						
<u>Lactic acid injection</u>						
Control samples						
	VI	165	607	606	99.8	
	VII	190	603	593	98.3	
	VIII	215	606	598	98.7	
	Av. (3 animals)		605	599	98.9	
	IX	237	601	590	98.2	
	X	262	606	600	99.0	
	Av. (5 animals)		605	597	98.8	
Injected samples						
	VI	165	605	665	648	107.1
	VII	190	602	662	619	102.8
	VIII	215	605	665	616	101.8
	Av. (3 animals)		604	664	628	103.9
	IX	237	605	665	626	103.5
	X	262	605	665	645	106.6
	Av. (5 animals)		604	664	631	104.4

(continued)

Table 6 (continued)

Animal no.	Sample no.	Initial wt. gm.	Beef plus injecting solution gm.	Wt. beef after aging gm.	%
LONGISSIMUS DORSI MUSCLE, RIB PORTION (continued)					
<u>Sodium chloride and lactic acid injection</u>					
Control samples					
VI	166	605		599	99.0
VII	191	601		589	98.0
VIII	213	607		603	99.3
Av. (3 animals)		604		597	98.8
IX	238	604		587	97.2
X	263	608		596	98.0
Av. (5 animals)		605		595	98.3
Injected samples					
VI	166	607	667	642	105.8
VII	191	605	665	622	102.8
VIII	213	607	667	646	106.4
Av. (3 animals)		606	666	637	105.0
IX	238	608	668	634	104.3
X	263	608	668	645	106.1
Av. (5 animals)		607	667	638	105.1

(continued)

Table 6 (continued)

Animal no.	Sample no.	Initial wt. gm.	Beef plus injecting solution gm.	Wt. beef after aging	
				gm.	%
PSOAS MAJOR AND PSOAS MINOR MUSCLES					
<u>Sodium chloride injection</u>					
Control samples					
VI	168	608		606	99.7
VII	193	604		594	98.3
VIII	218	605		597	98.7
Av. (3 animals)		606		599	98.9
IX	240	602		585	97.2
X	265	604		600	99.3
Av. (5 animals)		605		596	98.6
Injected samples					
VI	168	606	666	645	106.4
VII	193	606	665	646	106.8
VIII	218	606	666	650	107.3
Av. (3 animals)		606	666	647	106.8
IX	240	606	666	637	105.1
X	265	607	667	648	106.8
Av. (5 animals)		606	666	645	106.5

(continued)

Table 6 (continued)

Animal no.	Sample no.	Initial wt. gm.	Beef plus injecting solution gm.	Wt. beef after aging	
				gm.	%
PSOAS MAJOR AND PSOAS MINOR MUSCLES (continued)					
<u>Lactic acid injection</u>					
Control samples					
VI	169	600		599	99.8
VII	194	604		596	98.7
VIII	216	606		595	98.2
Av. (3 animals)		603		597	98.9
IX	241	607		601	99.0
X	266	602		596	99.0
Av. (5 animals)		604		597	98.9
Injected samples					
VI	169	601	661	630	104.8
VII	194	601	661	648	107.8
VIII	216	606	666	631	104.1
Av. (3 animals)		603	663	636	105.5
IX	241	604	664	639	105.8
X	266	605	665	636	105.1
Av. (5 animals)		603	663	637	105.5

(continued)

Table 6 (continued)

Animal no.	Sample no.	Initial wt.	Beef plus injecting solution	Wt. beef after aging	
		gm.	gm.	gm.	%
PSOAS MAJOR AND PSOAS MINOR MUSCLES (continued)					
<u>Sodium chloride and lactic acid injection</u>					
Control samples					
VI	170	605		600	99.2
VII	192	603		595	98.7
VIII	217	606		600	99.0
Av. (3 animals)		605		598	99.0
IX	242	608		599	98.5
X	264	605		599	99.0
Av. (5 animals)		605		599	98.9
Injected samples					
VI	170	607	667	639	105.3
VII	192	602	662	658	109.3
VIII	217	606	666	645	106.4
Av. (3 animals)		605	665	647	107.0
IX	242	602	662	636	105.6
X	264	605	665	636	105.1
Av. (5 animals)		604	664	643	106.3

(continued)

Table 6 (continued)

Animal no.	Sample no.	Initial wt.	Beef plus injecting solution	Wt. beef after aging	
		gm.	gm.	gm.	%
SEMITENDINOSUS MUSCLE					
<u>Sodium chloride injection</u>					
Control samples					
VI	172	608		605	99.5
VII	197	601		594	98.8
VIII	219	607		593	97.7
Av. (3 animals)		605		597	98.7
IX	244	605		593	98.0
X	269	604		601	99.5
Av. (5 animals)		605		597	98.7
Injected samples					
VI	172	607	667	634	104.4
VII	197	603	663	644	106.8
VIII	219	606	666	624	103.0
Av. (3 animals)		605	665	634	104.7
IX	244	604	664	638	105.6
X	269	605	665	638	105.4
Av. (5 animals)		605	665	636	105.0

(continued)

Table 6 (continued)

Animal no.	Sample no.	Initial wt. gm.	Beef plus injecting solution gm.	Wt. beef after aging gm.	%
SEMITENDINOSUS MUSCLE (continued)					
<u>Lactic acid injection</u>					
Control samples					
VI	173	608		607	99.8
VII	195	603		597	99.0
VIII	220	608		602	99.0
Av. (3 animals)		606		602	99.3
IX	245	609		602	98.8
X	267	604		600	99.3
Av. (5 animals)		606		602	99.2
Injected samples					
VI	173	601	661	630	104.8
VII	195	600	660	616	102.7
VIII	220	607	667	606	99.8
Av. (3 animals)		603	663	617	102.4
IX	245	602	662	600	99.7
X	267	608	668	633	104.1
Av. (5 animals)		604	664	617	102.2

(continued)

Table 6 (continued)

Animal no.	Sample no.	Initial wt.	Beef plus injecting solution	Wt. beef after aging	
		gm.	gm.	gm.	%
SEMITENDINOSUS MUSCLE (continued)					
<u>Sodium chloride and lactic acid injection</u>					
Control samples					
VI	171	607		600	98.8
VII	196	606		604	99.7
VIII	221	606		597	98.5
Av. (3 animals)		606		600	99.0
IX	243	604		588	97.4
X	268	606		602	99.3
Av. (5 animals)		606		598	98.7
Injected samples					
VI	171	606	666	636	105.0
VII	196	604	664	635	105.1
VIII	221	607	667	642	105.8
Av. (3 animals)		606	666	638	105.3
IX	243	604	664	617	102.2
X	268	602	662	634	105.3
Av. (5 animals)		605	665	633	104.7

(continued)

Table 6 (continued)

Animal no.	Sample no.	Initial wt.	Beef plus injecting solution	Wt. beef after aging	
		gm.	gm.	gm.	%
SEMIMEMBRANOSUS MUSCLE					
<u>Sodium chloride injection</u>					
Control samples					
VI	177	604		597	98.8
	179	609		597	98.0
Av.		606		597	98.4
VII	199	605		594	98.2
	200	600		586	97.7
Av.		602		590	98.0
VIII	222	606		591	97.5
	226	606		594	98.0
Av.		606		592	97.8
Av. (3 animals)		605		593	98.1
IX	249	607		592	97.5
	251	600		578	96.3
Av.		604		585	96.9
X	271	606		600	99.0
	272	604		596	98.7
Av.		605		598	98.8
Av. (5 animals)		605		592	98.0

(continued)

Table 6 (continued)

Animal no.	Sample no.	Initial wt.	Beef plus injecting solution	Wt. beef after aging	
		gm.	gm.	gm.	%
SEMIMEMBRANOSUS MUSCLE (continued)					
<u>Sodium chloride injection (continued)</u>					
Injected samples					
VI	177	604	664	649	107.4
	179	602	662	625	103.8
Av.		603	663	637	105.6
VII	199	601	661	628	104.5
	200	606	666	644	106.3
Av.		604	664	636	105.4
VIII	222	606	666	633	104.4
	226	606	666	644	106.3
Av.		606	666	638	105.4
Av. (3 animals)		604	664	637	105.5
IX	249	608	668	637	104.8
	251	608	668	636	104.6
Av.		608	668	636	104.7
X	271	606	666	637	105.1
	272	604	664	635	105.1
Av.		605	665	636	105.1
Av. (5 animals)		605	665	637	105.2

(continued)

Table 6 (continued)

Animal no.	Sample no.	Initial wt. gm.	Beef plus injecting solution gm.	Wt. beef after aging	
				gm.	%
SEMIMEMBRANOSUS MUSCLE (continued)					
<u>Lactic acid injection</u>					
Control samples					
VI	175	600		589	98.2
	176	605		596	98.5
Av.		602		592	98.4
VII	198	608		595	97.9
	202	605		595	98.3
Av.		606		595	98.1
VIII	225	607		597	98.4
	227	606		583	96.2
Av.		606		590	97.3
Av. (3 animals)		605		592	97.9
IX	247	603		588	97.5
	248	607		604	99.5
Av.		605		596	98.5
X	270	605		602	99.5
	274	606		599	98.8
Av.		606		600	99.2
Av. (5 animals)		605		595	98.3

(continued)

Table 6 (continued)

Animal no.	Sample no.	Initial wt. gm.	Beef plus injecting solution gm.	Wt. beef after aging	
				gm.	%
SEMIMEMBRANOSUS MUSCLE (continued)					
<u>Lactic acid injection (continued)</u>					
Injected samples					
VI	175	604	664	606	100.3
	176	605	665	603	99.7
Av.		604	664	604	100.0
VII	198	604	664	625	103.5
	202	602	662	613	101.8
Av.		603	663	619	102.6
VIII	225	606	666	612	101.0
	227	605	665	606	100.2
Av.		606	666	609	100.6
Av. (3 animals)		604	664	611	101.1
IX	247	604	664	604	100.0
	248	601	661	603	100.3
Av.		602	662	604	100.2
X	270	605	665	621	102.6
	274	606	666	628	103.6
Av.		606	666	625	103.1
Av. (5 animals)		604	664	612	101.3

(continued)

Table 6 (continued)

Animal no.	Sample no.	Initial wt. gm.	Beef plus injecting solution gm.	Wt. beef after aging	
				gm.	%
SEMIMEMBRANOSUS MUSCLE (continued)					
<u>Sodium chloride and lactic acid injection</u>					
Control samples					
VI	174	606		596	98.3
	178	608		594	97.7
Av.		607		595	98.0
VII	201	600		582	97.0
	203	606		594	98.0
Av.		603		588	97.5
VIII	223	608		596	98.0
	224	608		596	98.0
Av.		608		596	98.0
Av. (3 animals)		606		593	97.8
IX	246	604		593	98.2
	250	600		581	96.8
Av.		602		587	97.5
X	273	605		590	97.5
	275	604		600	99.3
Av.		604		595	98.4
Av. (5 animals)		605		592	97.9

(continued)

Table 6 (continued)

Animal no.	Sample no.	Initial wt.	Beef plus injecting solution	Wt. beef after aging	
		gm.	gm.	gm.	%
SEMIMEMBRANOSUS MUSCLE (continued)					
<u>Sodium chloride and lactic acid injection (continued)</u>					
Injected samples					
VI	174	600	660	625	104.2
	178	600	660	622	103.7
Av.		600	660	624	104.0
VII	201	606	666	637	105.1
	203	602	662	641	106.5
Av.		604	664	639	105.8
VIII	223	606	666	623	102.8
	224	607	667	619	102.0
Av.		606	666	621	102.4
Av. (3 animals)		603	663	628	104.1
IX	246	600	660	625	104.2
	250	608	668	630	103.6
Av.		604	664	628	103.9
X	273	605	665	641	106.0
	275	604	664	645	106.8
Av.		604	664	643	106.4
Av. (5 animals)		604	664	631	104.5

(continued)

Table 6 (continued)

Animal no.	Sample no.	Initial wt. gm.	Beef plus injecting solution gm.	Wt. beef after aging	
				gm.	%
BICEPS FEMORIS MUSCLE					
<u>Sodium chloride injection</u>					
Control samples					
VI	184	605		600	99.2
	185	606		601	99.2
Av.		606		600	99.2
VII	205	601		598	99.5
	207	607		601	99.0
Av.		604		600	99.2
VIII	228	605		596	98.5
	230	606		596	98.3
Av.		606		596	98.4
Av. (3 animals)		605		599	98.9
IX	256	607		595	98.0
	257	603		595	98.7
Av.		605		595	98.4
X	277	603		597	99.0
	279	607		600	98.8
Av.		605		598	98.9
Av. (5 animals)		605		598	98.8

(continued)

Table 6 (continued)

Animal no.	Sample no.	Initial wt.	Beef plus injecting solution	Wt. beef after aging	
		gm.	gm.	gm.	%
BICEPS FEMORIS MUSCLE (continued)					
<u>Sodium chloride injection (continued)</u>					
Injected samples					
VI	184	601	661	639	106.3
	185	606	666	643	106.1
Av.		604	664	641	106.2
VII	205	603	663	652	108.3
	207	607	667	664	109.4
Av.		605	665	658	108.8
VIII	228	605	665	638	105.4
	230	608	668	635	104.4
Av.		606	666	636	104.9
Av. (3 animals)		605	665	645	106.6
IX	256	603	663	635	105.3
	257	603	663	632	104.8
Av.		603	663	634	105.0
X	277	608	668	629	103.4
	279	600	660	641	106.8
Av.		604	664	635	105.1
Av. (5 animals)		604	664	641	106.0

(continued)

Table 6 (continued)

Animal no.	Sample no.	Initial wt.	Beef plus injecting solution	Wt. beef after aging	
		gm.	gm.	gm.	%
BICEPS FEMORIS MUSCLE (continued)					
<u>Lactic acid injection</u>					
Control samples					
VI	181	607		604	99.5
	183	609		600	98.5
Av.		608		602	99.0
VII	204	606		598	98.7
	206	602		595	98.8
Av.		604		596	98.8
VIII	232	605		596	98.5
	233	607		602	99.2
Av.		606		599	98.8
Av. (3 animals)		606		599	98.9
IX	253	608		601	98.8
	255	605		592	97.8
Av.		606		596	98.3
X	276	600		591	98.5
	278	608		597	98.2
Av.		604		594	98.4
Av. (5 animals)		606		597	98.7

(continued)

Table 6 (continued)

Animal no.	Sample no.	Initial wt.	Beef plus injecting solution	Wt. beef after aging	
		gm.	gm.	gm.	%
BICEPS FEMORIS MUSCLE (continued)					
<u>Lactic acid injection</u> (continued)					
Injected samples					
VI	181	603	663	630	104.5
	183	606	666	620	102.3
Av.		604	664	625	103.4
VII	204	603	663	637	105.6
	206	600	660	618	103.0
Av.		602	662	628	104.3
VIII	232	606	666	623	102.8
	233	605	665	624	103.1
Av.		606	666	624	103.0
Av. (3 animals)		604	664	626	103.6
IX	253	605	665	604	99.8
	255	607	667	623	102.6
Av.		606	666	614	101.2
X	276	605	665	633	104.6
	278	607	667	625	103.0
Av.		606	666	629	103.8
Av. (5 animals)		605	665	624	103.1

(continued)

Table 6 (continued)

Animal no.	Sample no.	Initial wt. gm.	Beef plus injecting solution gm.	Wt. beef after aging	
				gm.	%
BICEPS FEMORIS MUSCLE (continued)					
<u>Sodium chloride and lactic acid injection</u>					
Control samples					
VI	180	608		603	99.2
	182	600		592	98.7
Av.		604		598	99.0
VII	208	601		594	98.8
	209	606		602	99.3
Av.		604		598	99.0
VIII	229	606		603	99.5
	231	606		590	97.4
Av.		606		596	98.4
Av. (3 animals)		605		597	98.8
IX	252	603		589	97.7
	254	607		598	98.5
Av.		605		594	98.1
X	280	605		601	99.3
	281	607		605	99.7
Av.		606		603	99.5
Av. (5 animals)		605		598	98.8

(continued)

Table 6 (continued)

Animal no.	Sample no.	Initial wt. gm.	Beef plus injecting solution gm.	Wt. beef after aging gm.	%
BICEPS FEMORIS MUSCLE (continued)					
<u>Sodium chloride and lactic acid injection (continued)</u>					
Injected samples					
VI	180	604	664	656	108.6
	182	610	670	640	104.9
Av.		607	667	648	106.8
VII	208	603	663	642	106.5
	209	605	665	633	104.6
Av.		604	664	638	105.6
VIII	229	607	667	637	104.9
	231	606	666	632	104.3
Av.		606	666	634	104.6
Av. (3 animals)		606	666	640	105.7
IX	252	604	664	637	105.5
	254	604	664	629	104.1
Av.		604	664	633	104.8
X	280	608	668	648	106.6
	281	605	665	649	107.3
Av.		606	666	648	107.0
Av. (5 animals)		605	665	640	105.8

Table 7. Weight of Canned Meat and of Liquid. Weight of beef before processing; weight of meat and of liquid after processing; percentage weight of meat and of liquid based on weight of beef before processing.

Animal* no.	Sample no.	Wt. before processing gm.	Weight after processing			
			Meat		Liquid	
			gm.	%	gm.	%
LONGISSIMUS DORSI MUSCLE, LOIN PORTION						
<u>Sodium chloride injection</u>						
Control samples						
VI	162	568	379	66.7	188	33.1
VII	188	570	392	68.8	174	30.5
VIII	211	568	364	64.1	199	35.0
Av. (3 animals)		569	378	66.5	187	32.9
IX	234	568	376	66.2	188	33.1
X	260	569	373	65.6	192	33.7
Av. (5 animals)		569	377	66.3	188	33.1
Injected samples						
VI	162	570	376	66.0	192	33.7
VII	188	572	408	71.3	158	27.6
VIII	211	569	362	63.6	203	35.7
Av. (3 animals)		570	382	67.0	184	32.3
IX	234	568	360	63.4	138	24.3
X	260	569	406	71.4	146	25.6
Av. (5 animals)		570	382	67.1	167	29.4

(continued)

*Animals VI, VII, and VIII were steers, carcass grade Commercial, processed 65 minutes; Animal IX was a steer, carcass grade Commercial, processed 90 minutes; Animal X was a cow, carcass grade Cutter, processed 90 minutes.

Table 7 (continued)

Animal no.	Sample no.	Wt. before processing gm.	Weight after processing			
			Meat		Liquid	
			gm.	%	gm.	%
LONGISSIMUS DORSI MUSCLE, LOIN PORTION (continued)						
<u>Lactic acid injection</u>						
Control samples						
VI	164	568	391	68.8	176	31.0
VII	187	570	394	69.1	174	30.5
VIII	210	568	363	63.9	197	34.7
Av. (3 animals)		569	383	67.3	182	32.1
IX	236	570	378	66.3	182	31.9
X	259	568	378	66.5	189	33.3
Av. (5 animals)		569	381	66.9	184	32.3
Injected samples						
VI	164	569	380	66.8	188	33.0
VII	187	570	383	67.2	186	32.6
VIII	210	568	354	62.3	202	35.6
Av. (3 animals)		569	372	65.4	192	33.7
IX	236	568	365	64.3	196	34.5
X	259	568	364	64.1	188	33.1
Av. (5 animals)		569	369	64.9	192	33.8

(continued)

Table 7 (continued)

Animal no.	Sample no.	Wt. before processing gm.	Weight after processing			
			Meat		Liquid	
			gm.	%	gm.	%
LONGISSIMUS DORSI MUSCLE, LOIN PORTION (continued)						
<u>Sodium chloride and lactic acid injection</u>						
Control samples						
VI	163	568	387	68.1	178	31.4
VII	186	568	390	68.7	168	29.6
VIII	212	570	364	63.8	206	36.1
Av. (3 animals)		569	380	66.9	184	32.4
IX	235	569	366	64.3	194	34.1
X	258	568	372	65.5	185	32.6
Av. (5 animals)		569	376	66.1	186	32.8
Injected samples						
VI	163	569	385	67.7	185	32.5
VII	186	569	384	67.5	184	32.3
VIII	212	570	353	61.9	202	35.4
Av. (3 animals)		569	374	65.7	190	33.4
IX	235	568	355	62.5	210	37.0
X	258	569	395	69.5	156	27.4
Av. (5 animals)		569	374	65.8	187	32.9

(continued)

Table 7 (continued)

Animal no.	Sample no.	Wt. before processing gm.	Weight after processing			
			Meat		Liquid	
			gm.	%	gm.	%
LONGISSIMUS DORSI MUSCLE, RIB PORTION						
<u>Sodium chloride injection</u>						
Control samples						
VI	167	568	372	65.5	195	34.3
VII	189	573	391	68.2	165	28.8
VIII	214	570	378	66.3	181	31.8
Av. (3 animals)		570	380	66.7	180	31.6
IX	239	570	375	65.8	193	33.8
X	261	568	356	62.7	135	23.8
Av. (5 animals)		570	374	65.7	174	30.5
Injected samples						
VI	167	571	384	67.2	188	32.9
VII	189	573	384	67.0	159	27.7
VIII	214	570	357	62.6	209	36.7
Av. (3 animals)		571	375	65.6	185	32.4
IX	239	569	368	64.7	191	33.6
X	261	570	356	62.4	191	33.5
Av. (5 animals)		571	370	64.8	188	32.9

(continued)

Table 7 (continued)

Animal no.	Sample no.	Wt. before processing gm.	Weight after processing			
			Meat		Liquid	
			gm.	%	gm.	%
LONGISSIMUS DORSI MUSCLE, RIB PORTION (continued)						
<u>Lactic acid injection</u>						
Control samples						
VI	165	570	368	64.6	202	35.4
VII	190	571	424	74.2	143	25.0
VIII	215	570	379	66.5	192	33.7
Av. (3 animals)		570	390	68.4	179	31.4
IX	237	568	364	64.1	197	34.7
X	262	569	383	67.3	172	30.2
Av. (5 animals)		570	384	67.3	181	31.8
Injected samples						
VI	165	570	363	63.7	200	35.1
VII	190	568	390	68.7	168	29.6
VIII	215	569	359	63.1	201	35.3
Av. (3 animals)		569	371	65.2	190	33.3
IX	237	569	343	60.3	213	37.4
X	262	569	363	63.8	148	26.0
Av. (5 animals)		569	364	63.9	186	32.7

(continued)

Table 7 (continued)

Animal no.	Sample no.	Wt. before processing gm.	Weight after processing			
			Meat		Liquid	
			gm.	%	gm.	%
LONGISSIMUS DORSI MUSCLE, RIB PORTION (continued)						
<u>Sodium chloride and lactic acid injection</u>						
Control samples						
VI	166	570	378	66.3	191	33.5
VII	191	571	387	67.8	178	31.2
VIII	213	570	383	67.2	185	32.4
Av. (3 animals)		570	383	67.1	185	32.4
IX	238	569	374	65.7	187	32.9
X	263	570	378	66.3	168	29.5
Av. (5 animals)		570	380	66.7	182	31.9
Injected samples						
VI	166	570	388	68.1	183	32.1
VII	191	571	394	69.0	178	31.2
VIII	213	570	359	63.0	208	36.5
Av. (3 animals)		570	380	66.7	190	33.3
IX	238	569	356	62.6	206	36.2
X	263	569	385	67.7	153	26.9
Av. (5 animals)		570	376	66.1	186	32.6

(continued)

Table 7 (continued)

Animal no.	Sample no.	Wt. before processing gm.	Weight after processing			
			Meat		Liquid	
			gm.	%	gm.	%
PSOAS MAJOR AND PSOAS MINOR MUSCLES						
<u>Sodium chloride injection</u>						
Control samples						
VI	168	571	414	72.5	155	27.1
VII	193	568	425	74.8	138	24.3
VIII	218	568	379	66.7	189	33.3
Av. (3 animals)		569	406	71.3	161	28.2
IX	240	570	422	74.0	143	25.1
X	265	570	373	65.4	171	30.0
Av. (5 animals)		569	403	70.7	159	28.0
Injected samples						
VI	168	572	388	67.8	178	31.1
VII	193	569	423	74.3	145	25.5
VIII	218	568	359	63.2	207	36.4
Av. (3 animals)		570	390	68.4	177	31.0
IX	240	570	387	67.9	161	28.2
X	265	568	362	63.7	162	28.5
Av. (5 animals)		569	384	67.4	171	29.9

(continued)

Table 7 (continued)

Animal no.	Sample no.	Wt. before processing gm.	Weight after processing			
			Meat		Liquid	
			gm.	%	gm.	%
PSOAS MAJOR AND PSOAS MINOR MUSCLES (continued)						
<u>Lactic acid injection</u>						
Control samples						
VI	169	571	418	73.2	159	27.8
VII	194	568	436	76.8	127	22.4
VIII	216	569	413	72.6	151	26.5
Av. (3 animals)		569	422	74.2	146	25.6
IX	241	568	408	71.8	153	26.9
X	266	568	383	67.4	238	41.9
Av. (5 animals)		569	412	72.4	166	29.1
Injected samples						
VI	169	571	379	66.4	186	32.6
VII	194	570	369	64.7	189	33.2
VIII	216	568	380	66.9	182	32.0
Av. (3 animals)		570	376	66.0	186	32.6
IX	241	568	385	67.8	149	26.2
X	266	568	353	62.1	120	21.1
Av. (5 animals)		569	373	65.6	165	29.0

(continued)

Table 7 (continued)

Animal no.	Sample no.	Wt. before processing gm.	Weight after processing			
			Meat		Liquid	
			gm.	%	gm.	%
PSOAS MAJOR AND PSOAS MINOR MUSCLES (continued)						
<u>Sodium chloride and lactic acid injection</u>						
Control samples						
VI	170	570	396	69.5	175	30.7
VII	192	568	415	73.2	142	25.0
VIII	217	570	403	70.7	153	26.8
Av. (3 animals)		569	405	71.1	157	27.5
IX	242	568	382	67.2	151	26.6
X	264	570	393	68.9	139	24.4
Av. (5 animals)		569	398	69.9	152	26.7
Injected samples						
VI	170	572	391	68.4	183	32.0
VII	192	571	414	72.5	141	24.7
VIII	217	571	380	66.5	175	30.6
Av. (3 animals)		571	395	69.1	166	29.1
IX	242	569	373	65.6	175	30.8
X	264	568	363	63.9	180	31.7
Av. (5 animals)		570	384	67.4	171	30.0

(continued)

Table 7 (continued)

Animal no.	Sample no.	Wt. before processing gm.	Weight after processing			
			Meat		Liquid	
			gm.	%	gm.	%
SEMITENDINOSUS MUSCLE						
<u>Sodium chloride injection</u>						
Control samples						
VI	172	568	369	65.0	198	34.8
VII	197	569	405	71.2	162	28.5
VIII	219	570	371	65.1	198	34.7
Av. (3 animals)		569	382	67.1	186	32.7
IX	244	570	369	64.7	183	32.1
X	269	568	363	63.9	205	36.1
Av. (5 animals)		569	375	66.0	189	33.2
Injected samples						
VI	172	571	358	62.7	201	35.2
VII	197	570	384	67.4	187	32.8
VIII	219	570	374	65.6	170	29.8
Av. (3 animals)		570	372	65.2	186	32.6
IX	244	568	360	63.4	177	31.2
X	269	568	377	66.4	178	31.3
Av. (5 animals)		569	371	65.1	183	32.1

(continued)

Table 7 (continued)

Animal no.	Sample no.	Wt. before processing gm.	Weight after processing			
			Meat		Liquid	
			gm.	%	gm.	%
SEMITENDINOSUS MUSCLE (continued)						
<u>Lactic acid injection</u>						
Control samples						
VI	173	568	363	63.9	186	32.7
VII	195	568	375	66.0	187	32.9
VIII	220	568	362	63.7	195	34.3
Av. (3 animals)		568	367	64.5	189	33.3
IX	245	568	377	66.4	132	23.2
X	267	569	362	63.6	203	35.7
Av. (5 animals)		568	368	64.7	181	31.8
Injected samples						
VI	173	568	401	70.6	145	25.5
VII	195	569	372	65.4	191	33.6
VIII	220	568	360	63.4	203	35.7
Av. (3 animals)		568	378	66.5	180	31.6
IX	245	568	366	64.4	179	31.5
X	267	568	347	61.1	170	29.9
Av. (5 animals)		568	369	65.0	178	31.2

(continued)

Table 7 (continued)

Animal no.	Sample no.	Wt. before processing gm.	Weight after processing			
			Meat		Liquid	
			gm.	%	gm.	%
SEMITENDINOSUS MUSCLE (continued)						
<u>Sodium chloride and lactic acid injection</u>						
Control samples						
VI	171	568	372	65.5	193	34.0
VII	196	569	380	66.8	189	33.2
VIII	221	568	369	65.0	187	32.9
Av. (3 animals)		568	374	65.8	190	33.4
IX	243	569	364	64.0	194	34.1
X	268	568	365	64.3	204	35.9
Av. (5 animals)		568	370	65.1	193	34.0
Injected samples						
VI	171	570	370	64.9	122	21.4
VII	196	569	365	64.1	201	35.3
VIII	221	568	355	62.5	214	37.7
Av. (3 animals)		569	363	63.8	179	31.5
IX	243	568	370	65.1	170	29.9
X	268	570	359	63.0	187	32.8
Av. (5 animals)		569	364	63.9	179	31.4

(continued)

Table 7 (continued)

Animal no.	Sample no.	Wt. before processing gm.	Weight after processing			
			Meat		Liquid	
			gm.	%	gm.	%
SEMIMEMBRANOSUS MUSCLE						
<u>Sodium chloride injection</u>						
Control samples						
VI	177	570	363	63.7	208	36.5
	179	568	374	65.8	198	34.8
Av.		569	368	64.8	203	35.6
VII	199	568	355	62.5	213	37.5
	200	569	360	63.3	204	35.8
Av.		568	358	62.9	208	36.6
VIII	222	568	367	64.6	200	35.2
	226	571	355	62.2	213	37.3
Av.		570	361	63.4	206	36.2
Av. (3 animals)		569	362	63.7	206	36.1
IX	249	569	355	62.4	196	34.4
	251	568	367	64.6	199	35.0
Av.		568	361	63.5	198	34.7
X	271	568	327	57.6	171	30.1
	272	570	330	57.9	238	41.8
Av.		569	328	57.8	205	36.0
Av. (5 animals)		569	355	62.5	204	35.8

(continued)

Table 7 (continued)

Animal no.	Sample no.	Wt. before processing gm.	Weight after processing			
			Meat		Liquid	
			gm.	%	gm.	%
SEMIMEMBRANOSUS MUSCLE (continued)						
<u>Sodium chloride injection (continued)</u>						
Injected samples						
VI	177	571	345	60.4	226	39.6
	179	568	352	62.0	217	38.2
Av.		570	348	61.2	222	38.9
VII	199	568	346	60.9	220	38.7
	200	570	350	61.4	216	37.9
Av.		569	348	61.2	218	38.3
VIII	222	570	350	61.4	221	38.8
	226	570	330	57.9	221	38.8
Av.		570	340	59.6	221	38.8
Av. (3 animals)		570	345	60.7	220	38.7
IX	249	568	342	60.2	209	36.8
	251	568	345	60.7	206	36.3
Av.		568	344	60.4	208	36.6
X	271	568	344	60.6	219	38.6
	272	570	324	56.8	223	39.1
Av.		569	334	58.7	221	38.8
Av. (5 animals)		569	343	60.2	218	38.3

(continued)

Table 7 (continued)

Animal no.	Sample no.	Wt. before processing gm.	Weight after processing			
			Meat		Liquid	
			gm.	%	gm.	%
SEMIMEMBRANOSUS MUSCLE (continued)						
<u>Lactic acid injection</u>						
Control samples						
VI	175	568	374	65.8	190	33.4
	176	570	358	62.8	205	36.0
Av.		569	366	64.3	198	34.7
VII	198	568	364	64.1	200	35.2
	202	570	362	63.5	205	36.0
Av.		569	363	63.8	202	35.6
VIII	225	568	345	60.7	223	39.3
	227	568	358	63.0	212	37.3
Av.		568	352	61.8	218	38.3
Av. (3 animals)		569	360	63.3	206	36.2
IX	247	568	348	61.3	215	37.8
	248	569	359	63.1	162	28.5
Av.		568	354	62.2	188	33.2
X	270	569	318	55.9	233	40.9
	274	569	328	57.6	231	40.6
Av.		569	323	56.8	232	40.8
Av. (5 animals)		569	352	61.8	208	36.5

(continued)

Table 7 (continued)

Animal no.	Sample no.	Wt. before processing gm.	Weight after processing			
			Meat		Liquid	
			gm.	%	gm.	%
SEMIMEMBRANOSUS MUSCLE (continued)						
<u>Lactic acid injection (continued)</u>						
Injected samples						
VI	175	569	376	66.1	191	33.6
	176	568	350	61.6	214	37.7
Av.		568	363	63.8	202	35.6
VII	198	568	342	60.2	217	38.2
	202	570	358	62.8	209	36.7
Av.		569	350	61.5	213	37.4
VIII	225	568	333	58.6	231	40.7
	227	570	351	61.6	218	38.2
Av.		569	342	60.1	224	39.4
Av. (3 animals)		569	352	61.8	213	37.5
IX	247	568	343	60.4	215	37.8
	248	569	342	60.1	180	31.6
Av.		568	342	60.2	198	34.7
X	270	570	302	53.0	217	38.1
	274	570	316	55.4	174	30.5
Av.		570	309	54.2	196	34.3
Av. (5 animals)		569	341	60.0	207	36.3

(continued)

Table 7 (continued)

Animal no.	Sample no.	Wt. before processing gm.	Weight after processing			
			Meat		Liquid	
			gm.	%	gm.	%
SEMIMEMBRANOSUS MUSCLE (continued)						
<u>Sodium chloride and lactic acid injection</u>						
Control samples						
VI	174	570	368	64.6	198	34.7
	178	568	363	63.9	208	36.6
Av.		569	366	64.2	203	35.6
VII	201	568	363	63.9	204	35.9
	203	568	368	64.8	202	35.6
Av.		568	366	64.4	203	35.8
VIII	223	571	352	61.6	219	38.4
	224	570	347	60.9	218	38.2
Av.		570	350	61.2	218	38.3
Av. (3 animals)		569	361	63.3	208	36.6
IX	246	568	364	64.1	177	31.2
	250	568	355	62.5	211	37.1
Av.		568	360	63.3	194	34.2
X	273	568	332	58.4	237	41.7
	275	568	336	59.2	209	36.8
Av.		568	334	58.8	223	39.2
Av. (5 animals)		569	355	62.4	208	36.6

(continued)

Table 7 (continued)

Animal no.	Sample no.	Wt. before processing gm.	Weight after processing			
			Meat		Liquid	
			gm.	%	gm.	%
SEMI MEMBRANOSUS MUSCLE (continued)						
<u>Sodium chloride and lactic acid injection (continued)</u>						
Injected samples						
VI	174	572	366	64.0	209	36.5
	178	568	344	60.6	222	39.1
Av.		570	355	62.3	216	37.8
VII	201	569	352	61.9	216	38.0
	203	568	375	66.0	190	33.4
Av.		568	364	64.0	203	35.7
VIII	223	568	345	60.7	219	38.6
	224	570	336	58.9	218	38.2
Av.		569	340	59.8	218	38.4
Av. (3 animals)		569	353	62.0	212	37.3
IX	246	570	353	61.9	170	29.8
	250	569	346	60.8	222	39.0
Av.		570	350	61.4	196	34.4
X	273	568	326	57.4	217	38.2
	275	569	333	58.5	172	30.2
Av.		568	330	58.0	194	34.2
Av. (5 animals)		569	348	61.1	205	36.1

(continued)

Table 7 (continued)

Animal no.	Sample no.	Wt. before processing gm.	Weight after processing			
			Meat		Liquid	
			gm.	%	gm.	%
BICEPS FEMORIS MUSCLE						
<u>Sodium chloride injection</u>						
Control samples						
VI	184	568	401	70.6	170	29.9
	185	569	394	69.2	174	30.6
Av.		568	398	69.9	172	30.2
VII	205	569	392	68.9	180	31.6
	207	569	373	65.6	199	35.0
Av.		569	382	67.2	190	33.3
VIII	228	570	383	67.2	187	32.8
	230	570	344	60.4	210	36.8
Av.		570	364	63.8	198	34.8
Av. (3 animals)		569	381	67.0	187	32.8
IX	256	568	362	63.7	200	35.2
	257	568	359	63.2	204	35.9
Av.		568	360	63.4	202	35.6
X	277	570	356	62.4	212	37.2
	279	570	345	60.5	227	39.8
Av.		570	350	61.5	220	38.5
Av. (5 animals)		569	371	65.2	196	34.5

(continued)

Table 7 (continued)

Animal no.	Sample no.	Wt. before processing gm.	Weight after processing			
			Meat		Liquid	
			gm.	%	gm.	%
BICEPS FEMORIS MUSCLE (continued)						
<u>Sodium chloride injection (continued)</u>						
Injected samples						
VI	184	568	360	63.4	205	36.1
	185	568	373	65.7	195	34.3
Av.		568	366	64.6	200	35.2
VII	205	569	367	64.5	204	35.8
	207	568	350	61.6	221	38.9
Av.		568	358	63.0	212	37.4
VIII	228	570	347	60.9	223	39.1
	230	570	342	60.0	227	39.8
Av.		570	344	60.4	225	39.4
Av. (3 animals)		569	356	62.7	212	37.3
IX	256	569	366	64.3	197	34.6
	257	568	359	63.2	207	36.4
Av.		568	363	63.8	202	35.5
X	277	568	360	63.4	180	31.7
	279	570	337	59.1	241	42.3
Av.		569	348	61.2	210	37.0
Av. (5 animals)		569	356	62.6	210	36.9

(continued)

Table 7 (continued)

Animal no.	Sample no.	Wt. before processing gm.	Weight after processing			
			Meat		Liquid	
			gm.	%	gm.	%
BICEPS FEMORIS MUSCLE (continued)						
<u>Lactic acid injection</u>						
Control samples						
VI	181	569	372	65.4	196	34.4
	183	570	364	63.8	207	36.3
Av.		570	368	64.6	202	35.4
VII	204	568	391	68.8	178	31.3
	206	568	379	66.7	192	33.8
Av.		568	385	67.8	185	32.6
VIII	232	570	354	62.1	207	36.3
	233	569	357	62.7	212	37.2
Av.		570	356	62.4	210	36.8
Av. (3 animals)		569	370	64.9	199	34.9
IX	253	570	367	64.4	188	33.0
	255	569	364	64.0	205	36.0
Av.		570	366	64.2	197	34.5
X	276	569	375	65.9	195	34.3
	278	570	344	60.4	229	40.2
Av.		570	360	63.2	212	37.2
Av. (5 animals)		570	367	64.4	201	35.3

(continued)

Table 7 (continued)

Animal no.	Sample no.	Wt. before processing gm.	Weight after processing			
			Meat		Liquid	
			gm.	%	gm.	%
BICEPS FEMORIS MUSCLE (continued)						
<u>Lactic acid injection (continued)</u>						
Injected samples						
VI	181	570	343	60.2	223	39.1
	183	568	340	59.8	227	40.0
Av.		569	342	60.0	225	39.6
VII	204	569	370	65.0	196	34.4
	206	568	354	62.3	214	37.7
Av.		568	362	63.6	205	36.0
VIII	232	569	348	61.2	220	38.7
	233	569	350	61.5	216	38.0
Av.		569	349	61.4	218	38.4
Av. (3 animals)		569	351	61.7	216	38.0
IX	253	568	345	60.7	208	36.6
	255	568	337	59.3	227	40.0
Av.		568	341	60.0	218	38.3
X	276	569	348	61.2	222	39.0
	278	569	321	56.4	251	44.1
Av.		569	335	58.8	236	41.6
Av. (5 animals)		569	346	60.8	220	38.8

(continued)

Table 7 (continued)

Animal no.	Sample no.	Wt. before processing gm.	Weight after processing			
			Meat		Liquid	
			gm.	%	gm.	%
BICEPS FEMORIS MUSCLE (continued)						
<u>Sodium chloride and lactic acid injection</u>						
Control samples						
VI	180	568	417	73.4	140	24.6
	182	569	362	63.6	181	31.8
Av.		568	390	68.5	160	28.2
VII	208	568	382	67.2	194	34.2
	209	569	385	67.7	199	35.0
Av.		568	384	67.4	196	34.6
VIII	229	569	358	62.9	210	36.9
	231	570	370	64.9	199	34.9
Av.		570	364	63.9	204	35.9
Av. (3 animals)		569	379	66.6	187	32.9
IX	252	569	376	66.1	187	32.9
	254	570	358	62.8	213	37.4
Av.		570	367	64.5	200	35.2
X	280	569	355	62.4	193	33.9
	281	568	365	64.3	205	36.1
Av.		568	360	63.4	199	35.0
Av. (5 animals)		569	373	65.5	192	33.8

(continued)

Table 7 (continued)

Animal no.	Sample no.	Wt. before processing gm.	Weight after processing			
			Meat		Liquid	
			gm.	%	gm.	%
BICEPS FEMORIS MUSCLE (continued)						
<u>Sodium chloride and lactic acid injection (continued)</u>						
Injected samples						
VI	180	570	403	70.7	159	27.9
	182	570	368	64.6	203	35.6
Av.		570	386	67.6	181	31.8
VII	208	569	382	67.1	186	32.7
	209	568	378	66.5	188	33.1
Av.		568	380	66.8	187	32.9
VIII	229	571	354	62.0	220	38.5
	231	570	349	61.2	226	39.6
Av.		570	352	61.6	223	39.0
Av. (3 animals)		569	373	65.3	197	34.6
IX	252	569	363	63.8	185	32.5
	254	570	363	63.7	197	34.6
Av.		570	363	63.8	191	33.6
X	280	568	369	65.0	203	35.7
	281	569	364	64.0	195	34.3
Av.		568	366	64.5	199	35.0
Av. (5 animals)		569	369	64.9	196	34.5

Table 8. Analysis of Variance of Scores for Flavor of Canned Beef. Sodium chloride injection.

Source of variation	Degrees of freedom	Sum of squares	Mean square
ANIMALS VI, VII, AND VIII			
Total	35	111.2222	
Animals	2	2.9305	1.4652
Muscles	5	32.2222	6.4444
Error (a)	10	26.5695	2.6570
Treatments	1	36.0000	36.0000**
T x M	5	3.0000	.6000
Error (b)	12	10.5000	.8750
ANIMALS IX AND X			
Total	23	388.8333	
Animals	1	273.3750	273.3750**
Muscles	5	66.9583	13.3917
Error (a)	5	17.2500	3.4500
Treatments	1	26.0416	26.0416**
T x M	5	4.0834	.8167
Error (b)	6	1.1250	.1875

* - Significant.

** - Highly significant.

Table 9. Analysis of Variance of Scores for Flavor of Canned Beef. Sodium chloride and lactic acid injection.

Source of variation	Degrees of freedom	Sum of squares	Mean square
ANIMALS VI, VII, AND VIII			
Total	35	94.2431	
Animals	2	.5973	0.2986
Muscles	5	22.3681	4.4736
Error (a)	10	30.6527	3.0653
Treatments	1	25.8403	25.8403**
T x M	5	10.2014	2.0403**
Error (b)	12	4.5833	.3819
ANIMALS IX AND X			
Total	23	346.8333	
Animals	1	234.3750	234.3750**
Muscles	5	28.2083	5.6417
Error (a)	5	41.5000	8.3000
Treatments	1	30.3750	30.3750**
T x M	5	6.5000	1.3000
Error (b)	6	5.8750	.9792

* - Significant.

** - Highly significant.

Table 10. Analysis of Variance of Scores for Flavor of Canned Beef. Lactic acid injection.

Source of variation	Degrees of freedom	Sum of squares	Mean square
ANIMALS VI, VII, AND VIII			
Total	35	59.1875	
Animals	2	0.3750	0.1875
Muscles	5	22.8125	4.5625
Error (a)	10	17.6250	1.7625
Treatments	1	0.1736	.1736
T x M	5	7.7014	1.5403
Error (b)	12	10.5000	.8750
ANIMALS IX AND X			
Total	23	350.8333	
Animals	1	247.0416	247.0416**
Muscles	5	47.2083	9.4417
Error (a)	5	44.8334	8.9667
Treatments	1	.3750	.3750
T x M	5	3.0000	.6000
Error (b)	6	8.3750	1.3958

* - Significant.

** - Highly significant.

Table 11. Analysis of Variance of Scores for Tenderness of Canned Beef. Sodium chloride injection.

Source of variation	Degrees of freedom	Sum of squares	Mean square
ANIMALS VI, VII, AND VIII			
Total	35	685.5764	
Animals	2	12.0972	6.0486
Muscles	5	355.3681	71.0736*
Error (a)	10	164.2361	16.4236
Treatments	1	79.5070	79.5070**
T x M	5	26.5346	5.3069
Error (b)	12	47.8334	3.9861
ANIMALS IX AND X			
Total	23	251.9062	
Animals	1	5.5104	5.5104*
Muscles	5	147.9687	29.5937**
Error (a)	5	2.3021	.4604
Treatments	1	71.7604	71.7604**
T x M	5	10.8021	2.1604
Error (b)	6	13.5625	2.2604

* - Significant.

** - Highly significant.

Table 12. Analysis of Variance of Scores for Tenderness of Canned Beef. Sodium chloride and lactic acid injection.

Source of variation	Degrees of freedom	Sum of squares	Mean square
ANIMALS VI, VII, AND VIII			
Total	35	378.3056	
Animals	2	12.1806	6.0903
Muscles	5	185.1389	37.0278*
Error (a)	10	68.7361	6.8736
Treatments	1	84.0278	84.0278**
T x M	5	6.8056	1.3611
Error (b)	12	21.4166	1.7847
ANIMALS IX AND X			
Total	23	243.9896	
Animals	1	.5104	.5104
Muscles	5	100.8021	20.1604**
Error (a)	5	9.0521	1.8104
Treatments	1	102.0938	102.0938**
T x M	5	26.2187	5.2437*
Error (b)	6	5.3125	.8854

* - Significant.

** - Highly significant.

Table 13. Analysis of Variance of Scores for Tenderness of Canned Beef. Lactic acid injection.

Source of variation	Degrees of freedom	Sum of squares	Mean square
ANIMALS VI, VII, AND VIII			
Total	35	555.7222	
Animals	2	70.6805	35.3402**
Muscles	5	421.5555	84.3111**
Error (a)	10	35.7362	3.5736
Treatments	1	0.6972	0.6972
T x M	5	12.1362	2.4272
Error (b)	12	14.9166	1.2430
ANIMALS IX AND X			
Total	23	190.9896	
Animals	1	3.0104	3.0104
Muscles	5	142.5521	28.5104*
Error (a)	5	27.0521	5.4104
Treatments	1	.5104	.5104
T x M	5	9.3021	1.8604
Error (b)	6	8.5625	1.4271

* - Significant.

** - Highly significant.

Table 14. Analysis of Variance of Scores for Juiciness of Canned Beef. Sodium chloride injection.

Source of variation	Degrees of freedom	Sum of squares	Mean square
ANIMALS VI, VII, AND VIII			
Total	35	257.74	
Animals	2	6.51	3.255
Muscles	5	105.53	21.106
Error (a)	10	79.20	7.920
Treatments	1	29.34	29.34**
M x T	5	13.37	2.674
Error (b)	12	23.79	1.983
ANIMALS IX AND X			
Total	23	126.74	
Animals	1	3.01	3.01
Muscles	5	51.80	10.360
Error (a)	5	21.31	4.262
Treatments	1	19.26	19.26
M x T	5	4.56	0.912
Error (b)	6	26.80	4.467

* - Significant.

** - Highly significant.

Table 15. Analysis of Variance of Scores for Juiciness of Canned Beef. Sodium chloride and lactic acid injection.

Source of variation	Degrees of freedom	Sum of squares	Mean square
ANIMALS VI, VII, AND VIII			
Total	35	233.24	
Animals	2	2.05	1.025
Muscles	5	132.53	26.506**
Error (a)	10	34.79	3.479
Treatments	1	9.56	9.56*
M x T	5	37.98	7.596**
Error (b)	12	16.33	1.361
ANIMALS IX AND X			
Total	23	113.24	
Animals	1	1.26	1.26
Muscles	5	33.55	6.710
Error (a)	5	32.56	6.512
Treatments	1	17.51	17.51*
M x T	5	11.81	2.362
Error (b)	6	16.55	2.758

* - Significant.
 ** - Highly significant.

Table 16. Analysis of Variance of Scores for Juiciness of Canned Beef. Lactic acid injection.

Source of variation	Degrees of freedom	Sum of squares	Mean square
ANIMALS VI, VII, AND VIII			
Total	35	275.22	
Animals	2	25.43	12.715
Muscles	5	183.80	36.760**
Error (a)	10	37.99	3.799
Treatment	1	0.25	0.25
M x T	5	8.67	1.734
Error (b)	12	19.08	1.590
ANIMALS IX AND X			
Total	23	175.33	
Animals	1	2.04	2.04
Muscles	5	91.71	18.342
Error (a)	5	45.83	9.166
Treatments	1	1.04	1.04
M x T	5	4.08	0.816
Error (b)	6	31.08	5.180

* - Significant.

** - Highly significant.

Table 17. Analysis of Variance of Scores for Texture of Canned Beef. Sodium chloride injection.

Source of variation	Degrees of freedom	Sum of squares	Mean square
ANIMALS VI, VII, AND VIII			
Total	35	681.4722	
Animals	2	11.7222	5.8611
Muscles	5	337.1389	67.4278*
Error (a)	10	138.6111	13.8611
Treatments	1	106.7778	106.7778**
T x M	5	40.8888	8.1778
Error (b)	12	46.3334	3.8611
ANIMALS IX AND X			
Total	23	262.4583	
Animals	1	37.5000	37.5000*
Muscles	5	113.5833	22.7167*
Error (a)	5	16.1250	3.2250
Treatments	1	84.3750	84.3750**
T x M	5	9.2500	1.8500*
Error (b)	6	1.6250	0.2708

* - Significant.

** - Highly significant.

Table 18. Analysis of Variance of Scores for Texture of Canned Beef. Sodium chloride and lactic acid injection.

Source of variation	Degrees of freedom	Sum of squares	Mean square
ANIMALS VI, VII, AND VIII			
Total	35	492.50	
Animals	2	4.50	2.25
Muscles	5	263.00	52.60**
Error (a)	10	74.00	7.40
Treatments	1	106.78	106.78**
T x M	5	18.22	3.6440
Error (b)	12	26.00	2.1667
ANIMALS IX AND X			
Total	23	265.4583	
Animals	1	42.6666	42.6666**
Muscles	5	87.8333	17.5667*
Error (a)	5	12.7084	2.5417
Treatments	1	100.0416	100.0416**
T x M	5	10.3334	2.0667
Error (b)	6	11.8750	1.9792

* - Significant.
 ** - Highly significant.

Table 19. Analysis of Variance of Scores for Texture of Canned Beef. Lactic acid injection.

Source of variation	Degrees of freedom	Sum of squares	Mean square
ANIMALS VI, VII, AND VIII			
Total	35	546.7431	
Animals	2	75.3473	37.6736**
Muscles	5	406.5348	81.3070**
Error (a)	10	41.2360	4.1236
Treatments	1	0.8403	0.8403
T x M	5	12.5347	2.5069
Error (b)	12	10.2500	0.8542
ANIMALS IX AND X			
Total	23	262.8333	
Animals	1	51.0416	51.0416*
Muscles	5	154.2083	30.8417*
Error (a)	5	27.8334	5.5667
Treatments	1	3.3750	3.3750
T x M	5	2.5000	.5000
Error (b)	6	23.8750	3.9792

* - Significant.

** - Highly significant.

Table 20. Analysis of Variance of Scores for Slicing Quality of Canned Beef. Sodium chloride injection.

Source of variation	Degrees of freedom	Sum of squares	Mean square
ANIMALS VI, VII, AND VIII			
Total	35	1349.4097	
Animals	2	8.1805	4.0902
Muscles	5	790.3680	158.0736**
Error (a)	10	211.7362	21.1736
Treatments	1	37.0069	37.0069
T x M	5	185.7015	37.1403*
Error (b)	12	116.4166	9.7014
ANIMALS IX AND X			
Total	23	1168.24	
Animals	1	162.76	162.76**
Muscles	5	658.31	131.66**
Error (a)	5	24.54	4.908
Treatments	1	10.01	10.01
T x M	5	100.79	20.16
Error (b)	6	211.83	17.65

* - Significant.
 ** - Highly significant.

Table 21. Analysis of Variance of Scores for Slicing Quality of Canned Beef. Sodium chloride and lactic acid injection.

Source of variation	Degrees of freedom	Sum of squares	Mean square
ANIMALS VI, VII, AND VIII			
Total	35	985.00	
Animals	2	84.50	42.24
Muscles	5	557.3333	111.4667**
Error (a)	10	130.6667	13.0667
Treatments	1	1.3611	1.3611
T x M	5	37.1389	7.4278
Error (b)	12	174.0000	14.5000
ANIMALS IX AND X			
Total	23	1138.41	
Animals	1	207.10	207.10*
Muscles	5	529.72	105.94
Error (a)	5	105.22	21.04
Treatments	1	21.10	21.10
T x M	5	110.97	22.19
Error (b)	6	164.30	27.38

* - Significant.

** - Highly significant.

Table 22. Analysis of Variance of Scores for Slicing
Quality of Canned Beef. Lactic acid injection.

Source of variation	Degrees of freedom	Sum of squares	Mean square
ANIMALS VI, VII, AND VIII			
Total	35	1477.0000	
Animals	2	97.1250	48.5625
Muscles	5	820.0000	164.0000*
Error (a)	10	354.1250	35.4125
Treatments	1	5.4444	5.4444
T x M	5	79.5556	15.9111
Error (b)	12	120.7500	10.0625
ANIMALS IX AND X			
Total	23	1093.74	
Animals	1	184.26	184.26*
Muscles	5	691.57	138.31*
Error (a)	5	79.04	15.81
Treatments	1	1.26	1.26
T x M	5	103.29	20.66
Error (b)	6	34.32	5.72

* - Significant.

** - Highly significant.

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Sample No. _____

Date _____

SCORE CARD FOR MEAT

Slicing quality

Factor	10	9	8	7	6	5	4	3	2	1	Remarks
	Extremely good	Very good	Good	Medium		minus	Fair	Poor	Very poor	Extremely poor	
Aroma				plus		minus					
Flavor											
Liquid											
Lean											
Tenderness	Extremely tender	Very tender	Tender	Medium		minus	Fair	Tough	Very tough	Extremely tough	
				plus							
Juiciness	Extremely juicy	Very juicy	Juicy	Medium		minus	Fair	Dry	Very dry	Extremely dry	- 511 -
				plus							

Texture

Descriptive Terms

- Aroma
1. Mild -----
 2. Sharp -----
 3. Strong -----
 4. Faint -----
 5. Foreign ----
 6. -----
 7. -----
 8. -----

- Flavor
1. Flat -----
 2. Mild -----
 3. Mellowed -----
 4. Rich -----
 5. Strong -----
 6. Old -----
 7. Bitter -----
 8. Acid -----
 9. Salty -----
 10. Sweet -----

- Color of Lean
1. Light brown -----
 2. Dark brown -----
 3. Red and brown -----
 4. Gray -----
 5. Irridescent -----

- Texture
1. Stringy -----
 2. Dense, compact -----
 3. -----
 4. -----
 5. -----

Preference _____
 (among samples judged at one time)

Scorer _____